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Welcome

On behalf of the Fish Health Section/Asian Fisheries Society, I am delighted to welcome all delegates to the 5th Symposium on Diseases in Asian Aquaculture here on the beautiful Gold Coast.

The previous four Symposia held throughout Asian countries have each brought together over 170 fish health scientists, students, government workers and industry practitioners from some 20 countries to discuss problems affecting aquaculture production and their solutions. The Symposium is an important event for the aquaculture industry in the Asia-Pacific region. This is the first time that the symposium has been held in Australia with the theme of Healthy, Wealthy and Wise. Biosecurity and risk assessment have particularly been included since they have become critical issues in aquaculture at all levels of production and marketing as well as for all aspects of fisheries world wide.

I would like to take this opportunity to thank you for all your kind contributions to the Symposium. Finally, I hope this meeting will be a time to renew your interaction among colleagues and friends and hope you all enjoy the social events and the beauty of Gold Coast.

Supanee Chinabut

Chair, Fish Health Section /Asian Fisheries Society
(1999-2002)

AFS Fish Health Section Committee FHS Executive Committee (1999-2002)

Chairperson	Supanee Chinabut, Thailand
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Jiang Yulin, People's Republic of China
CV Mohan, India
Masud Khan, Bangladesh
Phan Thi Van, Vietnam

Past Chairperson and Editor of FHS Newsletter:

Celia Lavilla-Pitogo, Philippines

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Program Chair	Peter Walker	CSIRO, Australia
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Program Committee

Peter Walker	CSIRO, Australia
Melba Bondad-Reantaso	Network of Aquaculture Centres Asia-Pacific, Thailand
Timothy Flegel	Mahidol University, Thailand
Rohana Subasinghe	Food & Agricultural Organization of the UN, Italy
Peter Beers	Biosecurity Australia
Mike Hine	Ministry of Agriculture and Fisheries, New Zealand
Barry Munday	University of Tasmania, Australia
Barry Hill	Centre for Environment, Fisheries and Aquaculture Sciences, UK
Celia Lavilla-Pitogo	SEAFDEC, Philippines
Angus Cameron	AusVet, Australia

Keynote Speakers

Dr David Alderman

David Alderman is President of the European Association of Fish Pathologists, Senior Microbiologist at the CEFAS Weymouth Laboratory in UK and editor of the Disease Section of the international journal, *Aquaculture*. His research interests include a wide range of aquatic diseases and in more recent years have centred around pharmaceuticals and immunologicals in aquaculture. David has published more than 100 papers on these areas. A member of the Scientific Secretariat and Biologics Committees he provides expert advice to UK Veterinary Medicines Directorate on data submitted for Marketing Authorisations and has contributed extensively to international bodies such as FAO, WHO and ICES.

Dr Takashi Aoki

Takashi Aoki is a Professor in Fish Genetics and Biochemistry at the Tokyo University of Fisheries. Here he teaches graduate and undergraduate courses and directs a research program dealing with fish and shellfish diseases and immunology. Takashi Aoki has published extensively in the fish and shellfish diseases and immunology on topics including cloning and characterization of fish and shellfish immune-related genes, characterization of virulence and drug resistance genes of fish pathogenic bacteria, studies of vaccines for aquaculture and molecular diagnosis techniques for fish pathogens.

Dr Chris Baldock

Chris Baldock is one of Australia's leading veterinary epidemiologists. He is a specialist in the design and implementation of livestock disease control and surveillance programs within a business planning framework as well as in epidemiological research and training. He has undertaken numerous consultancies both in Australia and in Asia, has authored numerous scientific articles, is co-author for a number of epidemiology texts, and is on the editorial boards of two international scientific journals.

Dr Peter Beers

Peter Beers is Manager of Aquatic Animal Biosecurity at Biosecurity Australia in the federal Department of Agriculture, Fisheries and Forestry. He leads a team of seven professional staff who are responsible for developing Australia's aquatic animal health policies for the import and export of aquatic animals and their products. Peter has been closely involved in Australia's approach to risk assessment methodologies, quarantine legislation and the WTO's SPS Agreement. He has been directly involved in aquatic animal import risk analyses and the development of Australia's biosecurity policies for the importation of aquatic animals and their products since 1990.

Dr Franck C.J. Berthe

Franck Berthe, DVM, PhD, has been working, since 1995, at the IFREMER laboratory of La Tremblade, France. His responsibilities are: (a) research on *martelliosis*, a parasitic disease of molluscs, (b) coordinator of disease research and control programs in collaboration with mainland and overseas IFREMER teams, (c) head of the Reference Laboratory for Mollusc Diseases for the OIE and European Union, and (d) member of the Aquaculture Department Directorate in IFREMER. Franck is also a regular observer at the Fish Diseases Commission of the OIE. He has published research articles; he has been involved in organising training courses, workshops, conferences, etc. on fish and shellfish health.

Dr Melba Bondad-Reantaso

Melba B. Reantaso, was Aquatic Animal Health Specialist of NACA from 1999 to 2002 before taking up a post as Aquatic Animal Research Pathologist at the Cooperative Oxford Laboratory, Maryland Department of Natural Resources in September 2002. Prior to that Melba retired after 20 years of service at the Fish Health Section of the Philippine Bureau of Fisheries and Aquatic Resources (BFAR). While at NACA, she was in charge of the Asia-Pacific Regional Program on Aquatic Animal Health and worked closely with Asian governments, APEC, ASEAN, FAO, OIE and SEAFDEC and other aquatic animal health institutes in Asia-Pacific developing national strategies and regional projects and policies on various aspects of responsible health management. She is also currently Secretary/Treasurer of the Fish Health Section of the Asian Fisheries Society. Melba initiated the Molluscan Health Management Program, did some work on pearl oyster health while at BFAR and now currently involved in an annual active targeted surveillance of *Haplosporidium nelsoni* in Chesapeake Bay. Melba earned her PhD at University of Tokyo and post-doctoral studies at Nippon Veterinary and Animal Science University in Tokyo, Japan.

Dr Pornlerd Chanratchakool

Pornlerd Chanratchakool is currently working as the pathologist at The Aquatic Animal Health Research Institute (AAHRI), Department of Fisheries, Thailand. He has more than 10 years experience in both research and field work, in shrimp diseases and farm management. He is the chief author of the shrimp farm manual "Health Management in Shrimp Ponds". He has been invited to visit many countries and participate in seminars, workshops and conferences.

Keynote Speakers

Dr Albert (Kwang-Sik) Choi

Albert Choi - 1979 to 1983 Department of Oceanography, Inha University, Incheon Korea, Bs
1984 to 1992; Ms and Ph.D. from Dept. Oceanography, Texas A&M University, USA.

1995 to Present; Associate Professor at Cheju National University, Cheju Korea at the Department of Aquaculture. Research Interest; reproductive physiology and parasitic disease problems in marine bivalves including oysters and clams.

Dr Barry Hill

Barry Hill is the Chief Advisor for Fish and Shellfish Health to the UK government's Department of Environment, Food and Rural Affairs. For over 30 years, he has played a lead role in advising on scientific aspects of new UK legislation and policy on fish and shellfish diseases. He also provides specialist advice to the European Commission on EU legislation on the animal health conditions for trade in aquaculture animals. Since 1988, he has been a member of the OIE Fish Diseases Commission and has represented OIE at numerous scientific and technical meetings with other international organisations, particularly FAO and NACA.

Dr Alex Hyatt

Alex Hyatt is a Senior Principal Research Scientist and Project Leader of Electron Microscopy & Iridoviruses at the CSIRO Australian Animal Health Laboratory. He is an OIE recognised expert in ranaviruses and has developed electron microscopy as a key and core activity within Australia for the identification of new and emerging viruses. He is internationally known for his work in immunoelectron microscopy, virus morphogenesis, identification of new and emerging viruses and the development of assays for the detection of ranaviruses and chytrids. Alex has approximately 156 publications including many contributed chapters and one book. He has also received three major awards including the CSIRO Chairman's Medal, 1995, CSIRO Medal 2000 and the 2000 AQIS Quarantine Award 's for Science and Research.

Dr Chu-Fang Lo

Chu-Fang Lo is Professor of Zoology at National Taiwan University. Respected by her academic colleagues and appreciated by the aquaculture industry, she has won acclaim both locally and internationally for her work on the white spot syndrome virus (WSSV). Over the last decade, Dr Lo and her associates have conducted research on the virus itself, on the associated disease and on the development of control measures. She is currently investigating gene-based strategies against the disease.

Dr Toshihiro Nakai

Toshihiro Nakai teaches a graduate course at the Hiroshima University, Japan and directs a research program dealing with fish & shellfish pathology. Particularly, Nakai and colleagues concentrated their works on betanodavirus infections of marine fish for 10 years, and published many papers on this subject in scientific journals. Dr. Kazuhiro Nakajima, co-author of this symposium's presentation, who belongs to Fisheries Research Agency, has extensively studied iridovirus infections of fish, published many papers, and contributed to establish the current vaccination system against RSIV infections in Japan.

Dr Sirirat Rengpipat

Sirirat Rengpipat is an Associate Professor in Department of Microbiology, Faculty of Science, Chulalongkorn University, Bangkok, Thailand. Here she teaches both undergraduate and graduate courses dealing with Food Microbiology, Advance Bacterial Physiology and Metabolism, Microbial Enzyme, and Industrial Microbiology. Probiotic bacteria in shrimp feed is her interest of research topic for 8 years, she has isolated the effective probiont and demonstrated its benefits after shrimp taken as food supplement to *Penaeus monodon*. Competition exclusion, immune enhancement are the evidences found in shrimp fed probiotic fortified feed and using probiotic as prophylactic treatment is also possible.

Dr Rohana Subashinge

Rohana Subashinge is an experienced aquatic animal health specialist, gaining experience from many years of research and development work in Asia. He is currently connected with the Food and Agriculture Organization of the UN (FAO) since 2000, in-charge of managing the aquaculture programme of the Fishery Resources Division of the FAO Fisheries Department, which includes a major programme on aquatic animal health management. He has been instrumental in developing an Asia Regional Programme on Aquatic Animal Health Management in Asia, in collaboration with the Network of Aquaculture Centres in Asia-Pacific (NACA) and the Office International des Epizooties (OIE), and recently in Latin America. His work now covers all regions of the world.

Keynote Speakers

Prof Dr Willem B Van Muiswinkel

Willem Van Muiswinkel is a professor in Cell Biology and Immunology at the Department of Animal Sciences, Wageningen University, The Netherlands. He is teaching undergraduate courses in Cell Biology and graduate courses in Comparative Immunology. For more than 25 years Willem and his group have performed extensive studies on the immune system of fish and shrimp. Special focus is on cell characterisation, ontogeny, mucosal immunity and neuro-endocrine regulation. The results have been used in genetic selection programs to increase disease resistance, development of oral vaccines or for understanding the impact of stress on animal health and welfare in co-operation with (inter)national scientists, agencies and industries.

Invited Speakers

Dr Ellen Ariel

Ellen Ariel is a senior consultant at the European Community Reference Laboratory for Fish Diseases. She co-ordinates the daily activities of the reference laboratory and is in charge of the planning and execution of the annual inter-laboratory proficiency tests and the epidemiological surveys for notifiable diseases in the EU. Ellen organises Annual Meetings for National Reference Laboratories in the EU and associated countries and provides consultancy for fish disease laboratories in the EU predominantly. She is also a member of the private expert group advising to the Commission on legislative matters relating to the health management and disease control in aquaculture.

Dr Flavio Corsin

Flavio Corsin is a post-doctorate research assistant at the Institute of Aquaculture of the University of Stirling (UK). After he completed an MSc in Aquaculture, he joined a collaborative project with the University of Liverpool aimed at identifying the risk factors for White Spot Disease outbreaks. He conducted epidemiological studies in both Vietnam and India. Some of his work on the epidemiology of White Spot Disease is already available in peer-reviewed journals, while other results are either in press or in preparation.

Dr Iain East

Iain East is a scientific specialist with the Aquatic Animal Health section within Agriculture, Fisheries and Forestry - Australia. Iain gained his PhD from the University of Melbourne and worked in research in both the United States and Australia for 20 years and has published over 70 scientific books and papers. Iain's current role involves provision of scientific advice on the prevention and management of emergency disease incursions in fisheries and aquaculture, the development of resources to assist management of emergency disease incursions and the design and conduct of training in emergency management.

Prof Timothy W Flegel

Tim Flegel is the head of the Center of Excellence for Shrimp Molecular Biology and Biotechnology at the Faculty of Science, Mahidol University, Bangkok, Thailand. He is primarily a shrimp pathologist and is particularly interested in the shrimp response to viral pathogens. His group has worked on a number of important pathogens of the black tiger shrimp, including viruses, bacteria and parasites, for which they have developed a number of rapid diagnostic techniques. Together with Leigh Owens' group in Australia, they have pioneered work on phage induced virulence in the shrimp pathogen, *Vibrio harveyi*.

Dr Luc Grisez

Luc Grisez is research manager for Intervet Norbio Singapore. He is responsible for research on diseases in Asian aquaculture as well as for the development of aquatic animal health products. At present 14 scientists are employed in Intervet Norbio Singapore. Luc has been working on fish diseases throughout his career first during his studies at the University of Leuven in Belgium mainly involving freshwater fish species and later involving the Mediterranean fish species such as European sea bass and sea bream. He is with Intervet, an animal health company with HQ in The Netherlands, for the last 5 years and for the last 3 years he is based in Singapore.

Dr Lachlan Harris

Lachlan Harris originally studied microbiology at the University of Queensland, Brisbane, Australia. He then continued at postgraduate level at James Cook University, Townsville, Australia with Dr Leigh Owens, specializing in the understanding and control of bacterial diseases of penaeid larvae, in particular the interaction between larvae and luminous strains of *Vibrio harveyi*. Lachlan then worked for three years as Head of Technical Services for Seafarm Pty Ltd, Australia's largest prawn farming operation, continuing to investigate the control of bacterial and viral diseases in order to improve production, and also supervising Seafarm's selective breeding program for *Penaeus monodon* and *Penaeus merguensis*. In 2001, Lachlan travelled to Ecuador to commence working for Acuabiotec LLC, a company specializing in the control of bacterial diseases through the use of microbial technology. Between 2001 and 2002 Lachlan was responsible for implementing this technology and providing advice to a variety of shrimp farms in Ecuador severely affected by WSSV and bacterial diseases. Since July 2002 Lachlan has been working as the General Manager of Seaquest S. A., a program of maturation, genetic improvement and research and investigation responsible for an association of three shrimp producers in Ecuador, comprising 5000 hectares of extensive ponds cultivating *Penaeus vannamei*.

Invited Speakers

Dr Brian Jones

Brian Jones is senior fish pathologist at the Department of Fisheries, Government of Western Australia, Adjunct Professor of Fish Health at the Muresk Institute and Adjunct Associate Professor at Murdoch University School of Veterinary and Biomedical Sciences. Brian is author and co-author of over 80 scientific papers and technical reports and has broad international experience of both freshwater and marine shellfish aquaculture. For the past 7 years Brian has provided a disease diagnostic and surveillance service to the pearl oyster industry and was a key participant in a major 3 year survey to identify oyster diseases across northern Australia.

Professor Indrani Karunasagar

Indrani Karunasagar is an Associate Professor in Microbiology and Director of the UNESCO MIRCEN for Marine Biotechnology at the College of Fisheries of the University of Agriculture Sciences, Mangalore. She is also a scientific advisor to the International Foundation for Science, Sweden and a Member of the Task Force of the Department of Biotechnology, Govt. of India. Her research interests include: molecular diagnostics, bioremediation and probiotics, bacteriophages and shellfish toxins, use of biotechnological tools in health management, public health related pathogens.

Dr CV Mohan

CV Mohan is an Associate Professor of Fish Pathology at the College of Fisheries, University of Agricultural Sciences, Mangalore, India. He teaches several courses in the field of aquatic animal health to undergraduate and postgraduate students. Mohan and his associates, over the past 15 years, have completed several national and international funded research projects on various aspects of aquatic animal health. Mohan has published extensively in international (23) and national (25) journals on aquatic animal health topics including fish and shrimp pathology, diagnostics, immunology and epidemiology. He has been involved as an invited expert from India in many of the expert consultations and workshops organised by international agencies like NACA, DFID, FAO, World Bank and ACIAR.

Dr Barry Munday

Barry Munday is presently a Research Fellow in the School of Human Life Sciences at the University of Tasmania. Until 2000 he was a Reader in the School of Aquaculture with responsibility for the fish health program in that school. Since the early 1990s he has been involved in investigating disease problems in the Australian tuna Aquaculture industry and has also visited the Kinki University tuna research centre in Japan to obtain information on their propagation program.

Dr Antonius Suwanto

Antonius Suwanto is a Professor in the Department of Biology, Faculty of Science and Mathematics, Bogor Agricultural University, and a coordinator on Research Center for Microbial Diversity. In addition he is also the head of Molecular Biology Laboratory in the Southeast Asian Regional Center for Tropical Biology (SEAMEO-BIOTROP). He has been working on bacteriology and molecular genetics of Vibriosis in shrimp hatcheries in Indonesia for more than 10 years. He has published several papers on his work in this field. He is also a recipient for a number of scientific and academic awards from both national and international institutions, such as Rockefeller Foundation (USA), International Foundation for Science (Sweden), Indonesian Science and Technology Award (ITSF), and Kehati Award (Indonesian Biodiversity Award).

Prof Just M Vlak

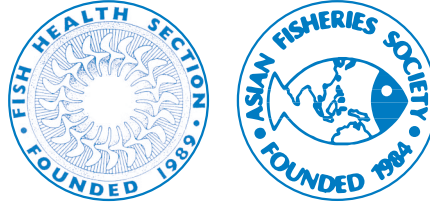
Just Vlak is a Professor of Virology at Wageningen University, The Netherlands. He is Chairman of the Biology Program and Director of the Graduate School 'Production Ecology and Resource Conservation'. His research program includes studies on the biology and genetics of baculoviruses (insect viruses) and their application as biocontrol agents of insect pests and expression vectors for eukaryotic proteins. Furthermore, his group studies the genetics of nimaviruses (White Spot Syndrome Virus) of crustaceans. At present he is Vice-President of the Society for Invertebrate Pathology. Over the past 25 years he published over 300 papers in international journals and books.

Prof Jiang Yulin

Jiang Yulin is a Professor of Fish diseases and director of the key Lab of aquatic animal diseases in Chinese quarantine system. Over past 20 years, he directed and finished 23 research programs dealing with aquatic pathology, immunology and detection methods such as hemorrhagic disease of grass carp, pox disease of carp, white spot disease of shrimp, lymphocystis, turtle diseases etc. He published about 70 papers on these fields.

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* Please note that this program is subject to change.

Sunday 24 November 2002

- 4.30pm Registration commences
- 6.30pm Welcome Reception at the Gold Coast International Hotel proudly sponsored by CSIRO

Monday 25 November 2002

- 8.00am Registration commences
- Opening Session proudly sponsored by CSIRO**
Chair: Peter Walker
- 8.30am Opening Ceremony
- 8.35am Dr Kamonporn Tonguthai (representing AFS)
- 8.40am Dr Supranee Chinabut (representing FHS)
- 8.45am Dr Richard Callinan (representing LOC)
- 8.50am Keynote: *Aquatic animal health management in Asia.* **Melba Bondad-Reantaso**, Cooperative Oxford Laboratory, USA
- 9.25am Keynote: *Food safety in aquaculture.* **David Alderman**, CEFAS Weymouth Laboratory, United Kingdom

10.00am Chief Veterinary Officer's Morning Tea

Session 1: Biosecurity and Risk Assessment – proudly sponsored by Biosecurity

Co-chairs: Barry Hill and Chris Baldock

- 10.30am Keynote Address: *Biosecurity: A new word for an old concept.* **Peter Beers**, Biosecurity, Australia
- 11.05am Invited Speaker: *Farm Level biosecurity in aquaculture.* **C.V Mohan**, College of Fisheries, India
- 11.30am *'To hazard or not to hazard that is the question.'* *How unknowns in science affect the identification of hazards in an import risk analysis.* **Sarah Kleeman**, Aquatic Animal Biosecurity, Australia
- 11.45am *The role of risk analysis and epidemiology in the development of biosecurity for aquaculture.* **Edmund Peeler**, Cefas, United Kingdom
- 12.00pm *Import risk analysis: Philippine experience.* **Joselito Somga**, Fish Health Section, Bureau Of Fisheries And Aquatic Resources, Philippines
- 12.15pm *A national survey to demonstrate freedom from white spot virus and yellow head virus in Australian crustaceans.* **Iain East**, Agriculture Fisheries & Forestry – Australia

12.30pm Lunch

Session 2: Diseases of aquatic vertebrates
Co-chairs: Jiang Yulin and Tina Hawkesford

- 1.30pm Keynote: *Recent advances of betanodaviruses and iridoviruses in Asian aquaculture.* **Toshihiro Nakai**, Hiroshima University, Japan
- 2.05pm Invited Speaker: *Diseases of Thunnus spp.: Emerging aquaculture species.* **Barry Munday**, School of Human Life Sciences - University of Tasmania, Australia
- 2.30pm *Susceptibility of marine fish species to Piscine nodavirus from orange-spotted grouper, Epinephelus coioides in the Philippines.* **Yukio Maeno**
- 2.45pm *Characterisation of an iridovirus isolated from diseased marble goby Oxyeleotris marmoratus (Bleeker, 1852).* **Pongpun Prasankok**, Chulalongkorn University, Thailand
- 3.00pm *Molecular characterization of a novel ranavirus isolated from grouper, Epinephelus spp.* **Qiwei Qin**, National University of Singapore, Singapore
- 3.15pm *Viral DNA sequences of genes encoding the ATPase and the major capsid protein of tropical iridovirus isolates which are pathogenic to fishes in Japan, South China Sea and Southeast Asian countries.* **Teruo Miyazaki**, Faculty Bioresources, Mie University, Japan
- 3.30pm Afternoon Tea

Session 3: Diseases of aquatic vertebrates – proudly sponsored by Queensland Department of Primary Industries

Co-chairs: Kamonporn Tonguthai and Kishio Hatai

- 4.00pm Keynote: *Ranavirus of fish, amphibians and reptiles.* **Alex Hyatt**, CSIRO Livestock Industries, Australia
- 4.35pm Invited Speaker: *Emerging diseases of soft-shell turtles.* **Jiang Yulin**, The Key Lab of Aquatic Animal Diseases, China
- 5.00pm *New and emerging disease in reef and Estuarine fishes from North Queensland, Australia.* **Rachel Bowater**, Queensland Department of Primary Industries, Australia
- 5.15pm *A virological survey in diseased grouper in Thailand using virus isolation and polymerase chain reaction (PCR) technique.* **Somkiat Kanchanakhan** Aquatic Animal Health Research Institute, Department of Fisheries, Thailand
- 5.30pm *Isolation and identification of Edwardsiella ictaluri from diseased Pangasius hypophthalmus (Sauvage) cultured in Vietnam.* **Margaret Crumlish**, Stirling University, Scotland
- 5.45pm *Viral hemorrhagic septicemia virus (VHSV) infection in Japanese flounder Paralichthys olivaceus.* **Kiyokuni Muroga**, Hiroshima University, Japan

6.00pm Sessions conclude

6.30pm EUS Workshop

**Moderators: Chris Baldock
Melba Bondad-Reantaso**

Invited Experts: **Vicki Blazer
Richard Callinan
Kishio Hatai
Indrani Karunasagar
CV Mohan**

The aim of the workshop is to achieve some agreement on "What is EUS?" Invited experts will each present their case definition with supporting evidence, followed by questions and answers from the audience. The workshop will (hopefully) conclude with an agreed case definition.

8.00pm Student Reception

**Proudly sponsored by
World Aquaculture Society**

Tuesday 26 November 2002

Session 4: Epidemiology
Co-chairs: CV Mohan and Angus Cameron

8.15am Keynote: *International trade and risk analysis.* **Chris Baldock**, AusVet Animal Health Services Australia

8.50am Invited Speaker: *Farm level risk factors for white spot disease outbreaks.* **Flavio Corsin**, University of Stirling, United Kingdom

9.15am *Application of epidemiology to support better health management in black tiger shrimp Penaeus monodon aquaculture: An experience from India.* **Arun Padiyar**, Network Of Aquaculture Centres In Asia-Pacific, Thailand

9.30am *Patterns of occurrence of the helminth parasites of the bullseye pufferfish (Sphoeroides annulatus) from Sinaloa, Mexico.* **Ana Roque**, Ciad,Ac, Mexico

9.45am *Epizootic haematopoietic necrosis virus – epidemiology and uncertainty.* **Richard Whittington** University of Sydney, Australia

10.00am Morning Tea **proudly sponsored by INVE**

Session 5: Shrimp Health 1
Co-chairs: Guang-Hsiung Kou and Boonsirm Withyachumnarnkul

10.30am Keynote: *Key farm management issues to reduce loss from diseases.* **Pornlerd Chanratchakool**, Kasetsart University Campus, Thailand

11.05am Invited Speaker: *Phage induced virulence in the shrimp pathogen Vibrio harveyi* **Timothy W Flegel**, Centex Shrimp, Thailand

11.30am *Experimental transmission of Hepatopancreatic parvovirus (HPV) infection in Penaeus monodon postlarvae* **Elena Catap**, Fish Health Section, Aquaculture Department-Southeast Asian Fisheries Development, Philippines

11.45am *Does spawning stress trigger the replication of white spot syndrome virus in shrimp?* **Guang-Hsiung Kou**, Department Of Zoology, National Taiwan University, ROC

12.00noon *Polychaetes not carries of white spot syndrome virus.* **Pisit Poltana**, Mahidol University, Thailand

12.15am *Predicting outbreaks of White Spot Disease in a semi-intensive Penaeus monodon culture system in Karnataka, India.* **James Turnbull**, Institute Of Aquaculture, United Kingdom

12.30pm Lunch

Session 6: Shrimp Health 2
Co-Chairs: Tim Flegel and Indrani Karunasagar

1.30pm Keynote: *White spot syndrome: What we have learned about the virus and the disease.* **Chu-Fang Lo**, National Taiwan University, Taiwan

2.05pm Invited Speaker: *Molecular genetics of white spot syndrome virus.* **Just M Vlank**, Laboratory Of Virology, The Netherlands

2.30pm *Breeding shrimp for disease resistance: A panacea or pariah* **Shaun Moss**, The Oceanic Institute, USA

2.45pm *B-type virus of Carconus mediterraneus and WSSV of Penaeid shrimp: Similarities and possible relationships.* **Jean-Robert Bonami**, CNRS, France

3.00pm *Use of WSSV cDNA microarray for gene profiling during WSSV infection in shrimps.* **Jimmy Kwang**, The National University Of Singapore, Singapore

3.15pm *Variations in tandem repeat DNA segments in the ribonucleotide reductase gene of white spot syndrome virus (WSSV) isolates from Vietnam.* **Hoa Tran Thi Tuyet**, CSIRO Livestock Industries, Australia

3.30pm Afternoon Tea

Session 7: Emerging Technologies
Co-chairs: Jimmy Kwang and Peter Walker

4.00pm Keynote: *Characterization of gene expression of biodefense related genes of Kuruma shrimp, Penaeus japonicus using real-time PCR technology.* **Takashi Aoki**, CSIRO Livestock Industries, Australia

4.35pm Invited Speaker: *Construction of recombinant Vibrio harveyi to study its adherence and pathogenicity in shrimp larvae.* **Antonius Suwanto**, Bogor Agricultural University Indonesia

- 5.00pm *Recent advances of studies on molecular genetics concerning defense mechanism and control disease in the Chinese shrimp, *Litopenaeus chinensis*. Jianhai Xiang, Chinese Academy of Sciences, China*
- 5.15pm *A hypothetical model for VHML bacteriophage conversion of *Vibrio harveyi*. Jane Oakey, Queensland Department Of Primary Industries, Australia*
- 5.30pm *Co-detection and differentiation of yellow head complex viruses using monoclonal antibodies. Chumporn Soowannayan CENTEX Shrimp, Thailand*
- 5.45pm *Molecular approach to the identification of virulence genes involved in *Edwardsiella tarda* pathogenesis. Srinivasa Rao P S, National University Of Singapore, Singapore*
- Chair: Supraanee Chinabut**
- 6.00pm Sixth Triennial General Meeting of the Fish Health Section – Asian Fisheries Society
- 8.00pm Hard Rock Café – optional evening
- 11.30am Invited Speaker: *Biocontrol of bacterial pathogens in aquaculture with emphasis on phage therapy. Indrani Karunasagar, University of Agricultural Sciences, College of Fisheries, India*
- 11.45am *Treatment of bacillary necrosis of larval Pacific oyster *Crassostrea gigas* with bacteriophages. Toshihiro Nakai, Hiroshima University, Japan*
- 12.00noon *The probiotic potential of *Vibrio alginolyticus* (Val 1) in the oyster hatchery. Cheok Keong Tan, University of Technology Sydney, Australia*
- 12.15pm *Antagonistic activity of *Aeromonas media* strain A199 against *Saprolegnia parasitica* in two species of finfish, the eel *Anguilla australis* and silver perch *Bidyanus bidyanus*. Josie Lategan, University Of Technology Sydney*
-
- 12.30pm Lunch
-
- 1.00pm Optional tours:
1. Aquaculture Farm Tour
 2. Golf at Sanctuary Cove
 3. Currumbin Bird Sanctuary & Wildlife Park
- Free evening

Wednesday 27 November 2002

- Session 8: Immunology**
Co-chairs: Ellen Ho and Barry Munday
- 8.15am Keynote: *Innate immunity in vertebrates and invertebrates. Willem van Muiswinkel, Wageningen University, The Netherlands*
- 8.50am Invited Speaker: *Vaccine development for Asian aquaculture. Luc Grisez, Intervet Norbio Singapore Pty Ltd Singapore*
- 9.15am *Immunostimulants induced immunity and its quantification in tiger shrimp, *Penaeus monodon* (fabrics) through 'challenge' and 'pro-PO assay'. Sugantham Felix, Fisheries Biotechnology Centre, India*
- 9.30am *Immunological properties of the phagocytosis and serum lectin in *Scaphara subcrenata*. Zhihong Liu, Chinese Academy of Fishery Sciences, China*
- 9.45am *Vaccination against white spot syndrome virus in shrimp. Jeroen Witteveldt, Wageningen University, The Netherlands*

10.00am Morning Tea

- Session 9: Probiotics and Therapeutics**
Co-chairs: Celia Lavilla-Pitogo and Leigh Owens
- 10.30am Keynote: *Probiotic bacteria: Are they beneficial? Sirirat Rengpipat, Chulalongkorn University, Thailand*
- 11.10am Invited Speaker: *Use of microbial technology to improve farm results in shrimp farm in Ecuador. Lachlan Harris, Gerante General de Seaquest SA, South America*

Thursday 28 November 2002

- Session 10: Mollusc Health 1**
Co-chairs: Franck Berthe and Robert Lester
- 8.15am Keynote: *Current Status of Perkinsis Infections in Asian water. Kwang-Sik Choi, Cheju National University, Korea*
- 8.50am Invited Speakers: *Diseases of pearl oysters. Brian Jones, Department of Fisheries, Western Australia, Australia*
- 9.15am *Report on oyster mortality in Wonboyn Lake, Australia. Damian Ogburn, NSW Fisheries, Australia*
- 9.30am *Transmission of *Perkinsus olseni* among wild blacklip abalone in South Australia. Craig Hayward, University of Queensland, Australia*
- 9.45am *Breeding for QX disease *Martelia sydneyi* resistance in Sydney rock oysters *Saccostrea glomerata*. John Nell, NSW Fisheries, Port Stephens Australia*

10.00am Morning Tea **proudly sponsored by University of Queensland**

- Session 11: Mollusc Health 2**
Co-chairs: Kwang-Sik Choi and Mike Hine
- 10.30am Keynote: *Diseases in mollusc hatcheries: a paradox in health management. Franck C.J. Berthe, IFREMER France*

11.05am	Invited Speaker: <i>Survey on the ovarian parasite, Martellioides chungmuensis in the cultured pacific oysters crassostrea gigas in Korea.</i> Myoung-Ae Park , National Fisheries Research & Development Institute, Republic of Korea	2.45pm	<i>Fatal, virus associated peripheral neuropathy and retinopathy (PNR) in farmed penaeus monodon in Eastern Australia.</i> Richard Callinan , NSW Fisheries, Australia
11.30am	<i>Martellioides chungmuensis (paramyxea), an intracellular parasite of the ovocyte of Pacific oyster Crassostrea gigas: Isolation and sequencing of small subunit ribosomal DNA.</i> Naoki Itoh , The University of Tokyo Japan	3.00pm	<i>Zoning for martelliosis in commercial rock oysters in Australia.</i> Robert D Adlard , Queensland Museum, Australia
11.45am	<i>Epizootiology and detection of nocardiosis in oysters.</i> Susan Bower , Fisheries and Oceans Canada Canada	3.15pm	<i>Field investigations on a serious disease outbreak and among common and koi carp in Indonesia.</i> Agus Sunarto , Central Research Institute for Aquaculture, Indonesia
12.00noon	<i>Diseases of cultured paua (Haliotis iris) in New Zealand.</i> Ben Diggles , National Institute Of Water & Atmospheric Research Ltd New Zealand	3.30pm	Afternoon Tea - Proudly sponsored by NSW Fisheries
12.15pm	<i>Discovery of the early infective stages of the protozoan parasite martella sydneyi in oysters and the implications for disease detection and control.</i> Sarah Kleeman , Aquatic Animal Biosecurity, Australia	Session 13: The Future	Co-chairs: Supranee Chinabut and Richard Callinan
12.30pm	Lunch	4.00pm	Keynote: <i>Improving aquatic animal health in Asia.</i> Rohana Subasinghe , Food and Agriculture Organisation of the UN, Italy
Session 12: Trans-boundary and emerging diseases		4.30pm	Invited Speaker: <i>Aquaculture health management: The Australian experience.</i> Iain East , Agriculture Fisheries & Forestry, Australia
Co-chairs: Rohana Subasinghe and Richard Whittington		4.50pm	<i>The role of extension in effecting on-farm practice change for controlling shrimp disease.</i> Derek Foster , Queensland Department of Primary Industries, Australia
1.30pm	Keynote: <i>Limitations to preventing increased international distribution of aquatic animal diseases.</i> Barry Hill , The Centre for Environment, Fisheries & Aquaculture Sciences, United Kingdom	5.10pm	Open Discussion Forum
2.05pm	Invited Speakers: <i>Ornamental disease vectors.</i> Ellen Ariel , EU Community Reference Laboratory For Fish Disease, Denmark	5.40pm	Closing Ceremony – Rohana Subasinghe , Food and Agriculture Organisation of the UN, Italy
2.30pm	<i>Preliminary molecular and biological characterisation of Mourilyan virus (MoV): A new bunya-related virus of penaeid prawns.</i> Jeff Cowley , CSIRO Livestock Industries, Australia	6.00pm	Symposium Concludes
		7.00pm	Symposium Dinner – Sea World proudly sponsored by Queensland Department of Primary Industries

PRESENTING AUTHOR	TITLE	THEME	POSTER BOARD#
Stephen Colquitt, Biosecurity Australia, AUSTRALIA	Quarantine detention of ornamental fish - Practical challenges involved in the implementation of a new policy	T1	1
Abu Tweb Abu Ahmed, Department of Zoology, BANGLADESH	Argulosis in Brood Carp Rearing Ponds of Bangladesh	T2	3
Kanit Chukanhom, Nippon Veterinary and Animal Science University, JAPAN	Freshwater fungi isolated from common carp (<i>Cyprinus carpio</i>) eggs in Thailand	T2	4
Gregoria Erazo-Pagador, Fish Health Section SEAFDEC-AQD, PHILIPPINES	Biology and pathogenicity of the gill monogenean (<i>Pseudorhabdosynochus</i> sp.) in grouper	T2	5
Susan Gibson-Kueh, Agri Food and Veterinary Authority of Singapore, SINGAPORE	Diagnosis of systemic iridoviral disease in fish	T2	6
Tiina Hawkesford, Fisheries and Aquaculture Development Queensland Department of Primary Industries, AUSTRALIA	Tasmanian Isolates of <i>Streptococcus</i> sp. biovar 1 and verified strains of <i>Lactococcus garvieae</i> and <i>Enterococcus seriolicida</i> compared by microbiological, molecular biological and "in vivo" techniques	T2	7
Mangalika Hettiarachchi, University of Kelaniya, Sri Lanka, SRI LANKA	A study on columnaris disease in guppy <i>Poecilia reticulata</i>	T2	8
Michael Kent, Oregon State University, USA	Diseases of Opakapaka held at the Hawaii Institute of Marine Biology	T2	9
Matthew Landos, NSW Fisheries, AUSTRALIA	Winter disease in farmed silver perch (<i>Bidyanus bidyanus</i>) in New South Wales, Australia	T2	10
Thitiporn Laoprasert, Aquatic Animal Health Research Institute Department of Fisheries, THAILAND	First report of systemic amoebosis in oscar, <i>Astronotus ocellatus</i>	T2	11
Thitiporn Laoprasert, Aquatic Animal Health Research Institute Department of Fisheries, THAILAND	Study on <i>Tetrahymena</i> infection in guppy (<i>Poecilia reticulata</i>)	T2	12
Ong-ard Lawhavit, Department Of Veterinary Microbiology And Immunology, Faculty Of Veterinary Medi, THAILAND	Mycobacteriosis in ranchu, goldfish (<i>Carassius auratus</i>) imported from Japan	T2	13
Theerawoot Lerssutthichawat, Rajamangala Institute Of Technology, THAILAND	Diversity of freshwater monogeneans from siluriform fishes of Thailand	T2	14
Allan Mooney, University Of Queensland, AUSTRALIA	Seasonal variation in the ectoparasite assemblage of <i>Pagrus auratus</i> cultured in sea-cages off Eastern Australia	T2	15
Barry Munday, School of Human Life Sciences - University of Tasmania, AUSTRALIA	A <i>Pentacapsula</i> species inhibiting propagation of striped trumpeter - an aquaculture candidate	T2	16
Huu Dung Nguyen, University of Fisheries, VIETNAM	Hemorrhaging septicemia due to <i>Aeromonas hydrophila</i> in the Mekong catfish (<i>Pangasius bocourti</i>) cultured in An Giang province - Vietnam	T2	17
Qiwei Qin, National University of Singapore, SINGAPORE	Production and characterization of monoclonal antibodies to Singapore grouper iridovirus (SGIV)	T2	18
A Rayamajhi, Department Of Microbiology & Parasitology, NEPAL	Aquatic oomycetes from southeast Queensland	T2	19
Dalisay Ribu, The University of Queensland, AUSTRALIA	A new genus of dracunculoid nematode from the gills of the pufferfish <i>Tragulichthys jaculiferus</i>	T2	20

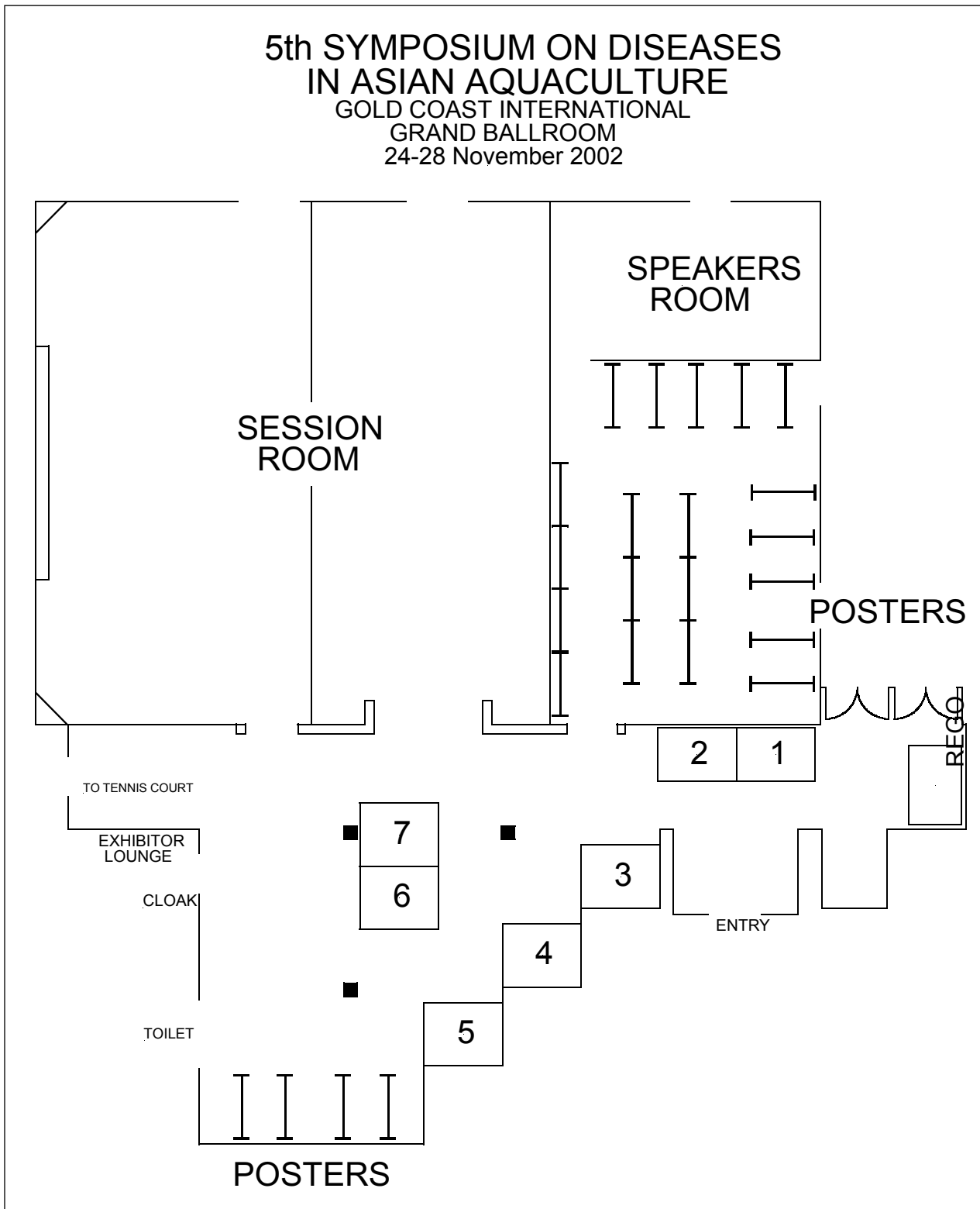
PRESENTING AUTHOR	TITLE	THEME	POSTER BOARD#
Somporn Roongkamnertwongsa, National Institute Of Coastal Aquaculture, THAILAND	Identification of a betanodavirus isolated from viral nervous necrosis-diseased redspotted grouper (<i>Epinephelus coioides</i>) cultured in Southern Thailand using PCR and sequence analysis	T2	21
Frances Stephens, Aquatilia Healthcare, AUSTRALIA	Health problems of captive West Australian dhufish	T2	22
Sivan Subburaju, Temasek Life Sciences Laboratory, SINGAPORE	Induction of caspase-dependent apoptosis by betanodaviruses GGNNV and identification of nucleolus localization signal of protein	T2	23
Kunika Wakita, Nippon Veterinary & Animal Science University, JAPAN	Histopathological study on tetrahymena infection in dwarf gourami (<i>Colisa ialia</i>)	T2	24
Yin-Geng Wang, Yellow Seas Fisheries Research Institute, CHINA	Turbot culture - a newly established industry in China and its disease problems	T2	25
Richard Whittington, The University of Sydney, AUSTRALIA	Pilchard herpesvirus in Australasia 1995-1999	T2	26
Arthur De Vera, Bureau Of Fisheries and Aquatic Resources, PHILIPPINES	Occurrence of hemic neoplasia in slipper oyster, <i>Crassostrea iredalei</i> (Faustino, 1928) in Dagupan City, Philippines	T2	27
Judith Handler, Department Of Primary Industries, Water And Environment, Tasmania, AUSTRALIA	Bacterial infection in Tasmanian farmed abalone: causes, pathology, farm factors and control options	T2	28
Craig Hayward, The University of Queensland, AUSTRALIA	Transmission of perkinsus olsenii among wild blacklip abalone in South Australia	T2	29
Linsheng Song, Institute of Oceanology, Chinese Academy of Sciences, P.R.CHINA	Potential genes involved in immune response identified by expressed sequence tag analysis from scallop <i>Chlamys farreri</i>	T3	30
Serge Corbeil, CSIRO Livestock Industries, AUSTRALIA	Development of a real-time PCR assay for the detection of <i>Piscirickettsia salmonis</i>	T4	31
Mark Crane, ARDL, Australian Animal Health Laboratory, AUSTRALIA	Development of diagnostic antibodies specific for white spot virus	T4	32
Ikuo Hirono, Tokyo University Of Fisheries, JAPAN	Functional microarray analysis of Japanese flounder <i>Paralichthys olivaceus</i> immune related genes for selection of a disease resistance fish	T4	33
Thammanoon Jaturapahu, Aquatic Animal Health Research Institute, THAILAND	Detection and identification of <i>Pseudomonas</i> spp. by polymerase chain reaction-reverse cross blot hybridization (PCR-RCBH) with 16S-23S rRNA intergenic spacer probes.	T4	34
Pani Prasad Kurcheti, Central Institute Of Fisheries Education, INDIA	Development of onfarm diagnostics	T4	35
Rosalind George Mulloorpeedikayil, Fisheries College and Res.Institute, INDIA	Plasmid profile of bacterial isolates from white spot affected shrimps	T4	36
James Munro, James Cook University, AUSTRALIA	Experimental Bacteriophage-mediated virulence in strains of <i>Vibrio harveyi</i>	T4	37
Nakao Nomura, University of Tsukuba, JAPAN	Development and characterization of a monoclonal antibody against white-spot syndrome virus in Penaeid shrimp	T4	38
Myoung-Ae Park, National Fisheries Research And Development Institute, REPUBLIC OF KOREA	Detection and comparison of lymphocystis virus in flounder (<i>Paralichthys olivaceus</i>) and sea bass (<i>Lateolabrax japonicus</i>)	T4	39
Panarat Phadee, Nippon Veterinary and Animal Science University, JAPAN	Identification and diagnosis of <i>Aphanomyces piscicida</i> by PCR	T4	40
Suppalak Puttinaowarat, Aquatic Animal Health Research Institute, THAILAND	Development of a monoclonal antibody to hybrid catfish (<i>Clarias macrocephalus</i> x <i>C. gariepinus</i>) immunoglobulin	T4	41

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Pramoda Sahoo, Institute For Animal Health, UNITED KINGDOM	Cloning and expression of variable region of nucleocapsid gene of aquatic morbilliviruses for serological diagnosis	T4	42
Pattira Taweepreda, Department of Anatomy, Faculty of Science, Mahidol University, THAILAND	Ultrastructural changes in the sperm and eggs of the black tiger shrimp, <i>Penaeus monodon</i> , before and after fertilization	T4	43
Attaporn Taweetungtragoon, Center of Excellence for Shrimp Molecular Biology & Biotechnology (CENTEX SHRIMP, THAILAND	A rapid multiplex real time PCR for the early detection of double targets: white spot syndrome virus (WSSV) and yellow head virus (YHV), in the black tiger shrimp <i>Penaeus monodon</i>	T4	44
Kanokporn Chayaburakul, Mahidol University, THAILAND	Infectious hypodermal and hematopoietic necrosis virus infection in Domesticated <i>P. monodon</i> broodstock	T6	45
Li-Li Chen, National Taiwan University, ROC	Transcriptional analysis of the DNA polymerase gene of shrimp white spot syndrome virus (WSSV)	T6	46
Enrique De La Vega, Australian Institute Of Marine Science, AUSTRALIA	Potential indicators of stress response and their relation to survival in <i>Penaeus monodon</i>	T6	47
Zhenyu Guo, Institute of Oceanology, Chinese Academy of Sciences, P.R.CHINA	A novel antimicrobial peptide isolated from the shrimp <i>Fenneropenaeus chinensis</i> after bacterial challenge	T6	48
Nguyen Van Hao, Research Institute For Aquaculture N2, VIETNAM	White Spot Disease in <i>Penaeus monodon</i> : case definition, accuracy of clinical diagnosis and description of an epidemic	T6	49
Chiu-Jung Huang, National Taiwan University, TAIWAN, ROC	The PK1 protein of the white spot syndrome virus (WSSV) is a nuclear kinase with autophosphorylation activity	T6	50
Celia Lavilla-Pitogo, SEAFDEC Aquaculture Department, PHILIPPINES	Evaluation of pathogenicity of bacterial strains in crustacean larvae by static bath: significance of monitoring bacterial counts.	T6	51
Kok Leong Lee, University Putra Malaysia, MALAYSIA	Growth pattern of <i>Vibrio parahaemolyticus</i> , <i>V. alginolyticus</i> and <i>V. harveyi</i> isolates from Malaysia	T6	52
Masatoshi Matsumura, University Of Tsukuba, JAPAN	Electro-chemical processes for phytoplankton control and shrimp disease disinfection	T6	53
Yasunori Muraosa, Nippoh Veterinary And Animal Science University, JAPAN	<i>Lagenidium thermophilum</i> isolated from zoeae of black tiger (<i>Penaeus monodon</i>) in Thailand	T6	54
Dang Thi Hoang Oanh, Cantho University, VIETNAM	Prevalence of white spot syndrome virus (WSSV) and <i>Monodon baculovirus</i> (MBV) infection in <i>Penaeus monodon</i> postlarvae in Vietnam	T6	55
Songsak Roekring, Center of Excellence for Shrimp Molecular Biology and Biotechnology (Centex Shri, THAILAND	Comparison of penaeid shrimp and insect parvoviruses suggests that viral transfers may occur between two distantly related arthropod groups	T6	56
Sivan Subburaju, Temasek Life Sciences Laboratory, SINGAPORE	The white spot syndrome virus (WSSV) infects specific hemocytes of the shrimp <i>Penaeus merguensis</i>	T6	57
Wasana Sukhumsirichart, Srinakharinwirot University, THAILAND	Prevalence of hepatopancreatic parvovirus (HPV) and <i>monodon baculovirus</i> (MBV) in stunted <i>Penaeus monodon</i> in Thailand	T6	58
Attaporn Taweetungtragoon, Center of Excellence for Shrimp Molecular Biology & Biotechnology (CENTEX SHRIMP, THAILAND	Comparison of various WSSV-PCR detection assays using naturally infected shrimp in Thailand and a fluorogenic WSSV probe as the gold standard	T6	59
Attaporn Taweetungtragoon, Center of Excellence for Shrimp Molecular Biology & Biotechnology (CENTEX SHRIMP, THAILAND	A dual, real time PCR for the simultaneous detection of white spot syndrome virus (WSSV) and yellow head virus (YHV) in the black tiger shrimp <i>Penaeus monodon</i>	T6	60
Eleonor Tendencia, Southeast Asian Fisheries Development Center, Aquaculture Department, PHILIPPINES	Effect of <i>Tilapia Tilapia hornorum</i> on luminous bacteria <i>Vibrio harveyi</i>	T6	61

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Han-Ching Wang, Department Of Zoology, National Taiwan University, TAIWAN, ROC	Does spawning stress trigger the replication of white spot syndrome virus in shrimp	T6	63
Priyanjalie Wijegoonawardane, Csiro, AUSTRALIA	Phylogenetic analysis of replicase (ORF1b) amplicon sequences reveals a fourth genotype of yellow head complex virus in P. monodon from India	T6	64
Lynette Williams, CSIRO Livestock Industries, AUSTRALIA	Attempts to establish continuous cell lines from prawn tissues	T6	65
Li Yu, Temasek Life Sciences Laboratory, SINGAPORE	Cloning and characterization of the White spot syndrome virus (WSSV) p25 gene and its promoter region	T6	66
Asmi Citra Malina A.R. Tassakka, United Graduate School Of Agricultural Sciences, JAPAN	Immunostimulatory effect of CpG oligodeoxynucleotides on the innate immune response of common carp, <i>Cyprinus carpio</i> L.	T7	67
Victoria Alday - Sanz, INVE Technologies, BELGIUM	Industry's need of standard challenge tests for shrimp	T7	68
Pramoda Sahoo, Institute For Animal Health, UNITED KINGDOM	Dietary intake of levamisole enhances the immune response and disease resistance of the Asian catfish, <i>Clarias batrachus</i>	T7	69
Sivan Subburaju, Temasek Life Sciences Laboratory, SINGAPORE	The general protein secretion machinery of <i>Aeromonas hydrophila</i> is involved in fast-killing mechanism of <i>c. elegans</i> and mortality of fish	T7	70
James Torres, University of the Philippines in the Visayas, PHILIPPINES	Effects of B-glucan on non-specific immune response in grouper: hematological analysis	T7	71
Kjersti Gravningen, Alpha Pharma Inc., UNITED STATES OF AMERICA	Bacterial disease prevention in high value marine fish culture by vaccination	T7	72
Clinton Chambers, University of Adelaide, AUSTRALIA	Orally administered praziquantel as a treatment for monogeneans infecting <i>Seriola quinqueradiata</i> : efficacy and practical considerations for a commercial fish farm	T8	73
Piyalai Hemtanon, Walailuk University, THAILAND	Studies on antiviral and antibacterial substances of <i>Spirulina platensis</i> for prevention the infectious diseases in black tiger shrimp (<i>Penaeus monodon</i>) caused by white spot syndrome virus and <i>Vibrio harveyi</i> .	T8	74
Orapin Khongpakdee, Institute Of Agricultural Technology, THAILAND	Studies on the effect of antibacterial and antiviral substance of marine diatom <i>Skeletonema costatum</i> for prevention the infectious diseases in black tiger shrimp (<i>Penaeus monodon</i>) caused by <i>Vibrio harveyi</i> and white spot syndrom virus	T8	75
Seyed Saeed Mirzargar, Aquatic Animal Health, Faculty of Veterinary Medicine, University of Tehran, IRAN	Determination of minimum detectable limit (MDL) of residues of selected antibacterial agents in common carp (<i>Cyprinus carpio</i>) using thin layer chromatography-bioautography (TLC-B)	T8	76
John Stephen Sampath Kumar, Fisheries College & Research Institute, INDIA	Herbal preparations for aquaculture bio-security - Indian experiences	T8	77
Ke Wang, Chinese Academy Of Sciences, PR CHINA	The anti-virus effects of glucosamine and melaleuca oil on Chinese prawn (<i>Penaeus chinensis</i>)	T8	78
Shih-Chu Chen, National Pingtung University of Science and Technology, TAIWAN, ROC	<i>Lactococcus garvieae</i> , a bacterial infection in grey mullet <i>Mugil cephalus</i>	T9	79
CV Mohan, College of Fisheries, INDIA	Health status of cultured shrimp at harvest - epidemiological significance	T9	80

PRESENTING AUTHOR	TITLE	THEME	POSTER BOARD#
Kenton Morgan, University of Liverpool, UNITED KINGDOM	Feeding farmed shrimp with shrimp waste - the lessons for aquaculture from BSE	T9	81
Ana Roque, Ciad,Ac, MEXICO	Effect of methylparathion on the susceptibility of shrimp (<i>Litopenaeus vannamei</i>) on the development of vibriosis	T9	82
Judith Silapan, UP In The Visayas, Cebu College, PHILIPPINES	Acute toxicities of deltamethrin and lambda-cyhalothrin to the fry of green grouper (<i>Epinephelus tauvina</i> Forsskål) and milkfish (<i>Chanos chanos</i> Forsskål)	T9	83
Temdoung Somsiri, Aquatic Animal Health Research Institute, THAILAND	Contamination of mycobacterium spp. in live feed	T9	84
Kishio Hatai, Division of Fish Diseases, JAPAN	Mycobacteriosis in ranchu goldfish (<i>Carassius auratus</i>) imported from Japan	T10	85
Barry Munday, School of Human Life Sciences - University of Tasmania, AUSTRALIA	Fish kill of mullet <i>Liza klunzingeri</i> in Kuwait Bay: The role of <i>Streptococcus agalactiae</i> and the influence of temperature	T10	86
Indunil D.S.I.P Thilakaratne, Veterinary Research Officer, SRI LANKA	Seasonal prevalence of trichodina infection in guppies (<i>Poecilia reticulata</i>) and goldfish (<i>Carassius auratus</i>) cultured for export in Sri Lanka	T10	87
Jiang Yulin, Key Lab of Aquatic Animal Diseases, REPUBLIC OF CHINA	Isolation of IHN virus from imported turbot (<i>Scophthalmus maximus</i>)	T10	88

Trade Exhibition Floorplan



Fish Health Section - Asian Fisheries Society

(Booth 1)

The Fish Health Section of the Asian Fisheries Society (FHS/AFS) is one of four global professional societies involved in aquatic animal health. The FHS has the following objectives: (a) promote effective interaction and cooperation among persons involved in fish health research; (b) encourage and promote investigation and advances in knowledge of fish health; (c) focus attention on fish health problems by disseminating technical and other information on all aspects of fish health; and (d) promote the proper implementation of proper implementation of effective fish health protection practices in the region. To achieve its objectives, the FHS organizes a triennial "Symposium on Diseases in Asian Aquaculture" (DAA), as well as other scientific seminars/workshops on topics of interest to the membership, independently or in conjunction with other organizations and implements projects relevant to aquatic animal health. The Section publishes the proceeding of DAA, a newsletter and other project related reports. The Section is administered by an Executive Committee (ExeCom) consisting of a Past Chairperson, Chairperson, Vice-Chairperson, Secretary-Treasurer, Newsletter Editor, and six elected members; governed by the FHS/AFS ByLaws; and holds a Triennial General Meeting of members in conjunction with DAA. Membership is open to any person interested in the furtherance of Fish Health Section objectives and who is a member in good standing of the Asian Fisheries Society. More detailed information about the FHS activities can be found at <http://afs-fhs.seafdec.org.ph/> <http://afs-fhs.seafdec.org.ph/>

Biosecurity Australia

(Booth 4)

Biosecurity Australia is a one of the operating groups within Agriculture, Fisheries and Forestry- Australia. Our main objective is to ensure that Australian animal and plant industries have increased access to markets and are protected from diseases and pests. The tasks of facilitating the export of live aquatic animals and genetic products and the development of policies that allow the safe importation of aquatic animals and their products is carried out by Aquatic Animal Biosecurity.

Biosecurity Australia - AFFA
Agriculture, Fisheries and Forestry Australia
GPO Box 858 CANBERRA ACT 2601
Work Phone: 02 6272 5330 Fax: 02 6272 3399

CSIRO

(Booths 6 & 7)

CSIRO Livestock Industries conducts research for Australia's livestock and allied industries to facilitate their sustainability and long-term competitiveness. We create, develop and commercialise technologies for novel products, new production options, improved production efficiency, disease control and product quality throughout the livestock industry value chain. We have a significant aquaculture diagnostic capability through our Australian Animal Health Laboratory and are working with a range of research collaborators on prawn, yabby and finfish aquaculture.

CSIRO Livestock Industries
120 Meiers Road INDOOROOPILLY QLD 4068
Work Phone: 07-3214-2700 Fax: 07-3124-2852

Primo/INVE

(Booth 2)

INVE is a multinational group of companies, active in agri and aquaculture. Our aquaculture Business Units provide nutritional and health solutions in fish and shrimp rearing. INVE are leaders in the development, production and commercialisation of a complete range of high quality products, comprising Artemia cysts; a Selco line of rotifer and Artemia enrichment products; an extensive range of compound starter and broodstock diets; a Health line; Standard and Specialty premixes and Concentrates for various stages/species. For more information, please visit us at www.inve.com

Mrs Liz Evans - Primo Aquaculture P/L
PO Box 8007 COFFS HARBOUR NSW 2450
Work Phone: 02 6655 4463 Fax: 02 6655 4988 E-Mail: primaqa@midcoast.com.au

QDPI

(Booth 3)

The Queensland Department of Primary Industries (QDPI) is proud to be a major sponsor of the Symposium. Aquaculture is a growing industry in Queensland and aquatic animal health is an important aspect of the continued expansion of the industry. Prawns, barramundi, silver perch, crayfish and oysters are the peak industries in aquaculture in Queensland are. QDPI plays a major role in research and development to support the sustainable growth of aquaculture in Queensland.

Queensland Department of Primary Industries
Level 2, Primary Industries Building 80 Ann Street, Brisbane QLD 4000
PO Box 46 BRISBANE QLD 4000
Work Phone: 07 3234 0701 Fax: 07 3239 0439

University of Queensland

(Booth 5)

The University of Queensland's Faculty of Biological and Chemical Sciences (BACS) is world-renowned for its strengths in teaching and research in the areas of biological and chemical sciences and since 1981 has been named as Australia's Number 1 Biological Sciences Research University.* Brisbane is fast becoming a centre for excellence in molecular biosciences and a major hub for bioindustries in the Asia-Pacific region, with innovative research institutes based at the University of Queensland. The development of the Institute of Molecular Biosciences (a major joint University / CSIRO / Government initiative) on the UQ campus, confirms this status. As well as underpinning various areas of study in the Faculty, Chemistry links strongly to Biology through new initiatives in Structural Biology. BACS also has the greatest breadth and depth of academic expertise in Ecology, Conservation Biology, Environmental Science, Marine Studies and Aquaculture of any university in Australia and is ranked as one of the best in the world in these fields.

Department of Microbiology and Parasitology
The University of Queensland
Sir Fred Schonell Drive ST LUCIA QLD 4067
Work Phone: 07 3365 3305 Fax: 07 3365 4699

Social Program

Welcome Reception

Sunday 24 November
6.30 – 8.30pm
Gold Coast International Hotel

Take the opportunity to relax with new and old friends at the official Symposium Welcome Reception. Attendance is included for all registrants.

Lunches

Lunches on Monday, Tuesday, Wednesday, and Thursday will be held in the Trade and Poster areas.

Official Symposium Dinner

Thursday 28 November 7.00pm
Sea World, Main Beach

Dinner promises to be the Symposium highlight! Take a Behind the Scenes Tour followed by the opportunity to relax and converse with colleagues over a sumptuous banquet. Included for all registrants. Please note that coaches will depart from the Gold Coast International Hotel at 6.30pm.

General Information

Symposium Venue

Gold Coast International Hotel
Cnr. Gold Coast Highway & Staghorn Avenue
Surfers Paradise QLD 4217

Telephone	Facsimile
(07) 5584 1200	(07) 5584 1280
Int: +61 7 5584 1200	Int: +61 7 5584 1280

Messages

A message board will be located in the Symposium registration area. Please advise potential callers to contact the Gold Coast International Hotel and ask for the 5th Symposium on Diseases in Asian Aquaculture registration desk.

All messages will be placed on the message board. Delegates are asked to check the board regularly throughout the symposium. The Symposium Secretariat takes no responsibility for messages not delivered to the delegate.

Telephones

All mobile phones and pages are to be turned off when delegates are in session. Public telephones are located throughout the hotel.

Prayer Room for Delegates Observing Ramadan

A dedicated room has been set aside as a prayer room for delegates observing Ramadan. This is located on level 2 of the Gold Coast International Hotel. Please enter via the Boardroom - turn left and it is the first door on the right. There will be a sign on the door indicating that this is the Prayer Room.

Symposium Registration Desk

The registration desk located in the foyer of the conference centre will be staffed at the following times:

Sunday 24 November	4.30pm – 6.30pm
Monday 25 November	8.00am – 6.00pm
Tuesday 26 November	7.45am – 6.00pm
Wednesday 27 November	7.45am – 1.00pm
Thursday 28 November	7.45am – 6.00pm

Name Badges

Delegates are requested to wear their name badge at all times during the Symposium.

Dress

Dress for all Symposium sessions and social events is smart casual. Smart casual means open neck shirt and trousers/shorts for gentlemen.

Personal Requirements

If you have any special dietary requirements such as Vegan or Halal and did not indicate this at the time of registration, please let us know so we can make the necessary arrangements.

If you have any medical, wheel chair access, or other special requirements, please let us know so we can assist you.

Personal Mail and Deliveries

Personal deliveries should be sent to your accommodation address.

Tickets

Tickets will be required for entry into the social functions. Delegates who have paid for "additional" reception, lunch and dinner tickets will be given the appropriate ticket(s). All tickets will be inside your registration envelope.

Ticket Refunds

Ticket refunds for social functions will be available if participation is cancelled more than 48 hours prior to the event.

Trade Exhibition

All morning and afternoon teas will be served in the trade area.

Sunday 24 November	6.30pm – 8.30pm
Monday 25 November	8.00am – 6.00pm
Tuesday 26 November	8.15am – 6.00pm
Wednesday 27 November	8.15am – 1.00pm
Thursday 28 November	8.15am – 4.00pm

Poster Exhibition

Posters will be displayed for the duration of the Symposium. Posters should be in place by no later than 5.30pm on Sunday 24 November and must be collected by 6.00pm on Thursday 28 November. Poster boards will be numbered as indicated in the poster program. Please refer to the poster program for further details. Delegates are encouraged to visit all the poster displays during coffee and lunch breaks and the welcome drinks.

Accommodation

Please ensure that the balance of your accommodation (if applicable) is paid directly to your hotel on departure, as well as any incidentals.

Disclaimer

All information disclosed in the Symposium Program is correct at time of printing. The Organising Committee reserves the right to alter the Symposium Program in the event of unforeseen circumstances.

All speakers were invited to provide abstracts for the Symposium Handbook, unfortunately not all were able to provide us with their papers.

Symposium Organisers

OzAccom Conference Services
PO Box 164 FORTITUDE VALLEY QLD 4006
Tel: +61(0)7 3854 1611 Fax: +61(0)7 3854 1507
Email: ozaccom@ozaccom.com.au

Fish Health Section – Asian Fisheries Society Member Directory

#	Name	E-mail/Contact Address
---	------	------------------------

AUSTRALIA

1	Anderson, Ian G.	lan.anderson@dpi.qld.gov.au Oonoonba Veterinary Laboratory P.O. Box 1085 Townsville, Queensland 4810 Australia Tel: (61) 7-47222610 Fax: (61) 7-47784307
2	Callinan, Richard	richard.callinan@agric.nsw.gov.au New South Wales Fisheries Regional Veterinary Laboratory Wollongbar NSW 2477 AUSTRALIA Tel (61) 2 6626 1294 Mob 0427492027 Fax (61) 2 6626 1276
3	Campbell, R. S. F.	Graduate School of Tropical Veterinary Science James Cook University, Townsville, Qld 4811
4	Hanna, Peter John	School of Biological & Chemical Sciences, Deakin University, Geelong, Victoria 3217
5	Lester, Robert J. G.	R.Lester@mailbox.uq.edu.au Department of Microbiology and Parasitology The University of Queensland Brisbane 4072 Australia Tel: 61-7-3365-3305 Fax: 61-7-3365-4620
6	Lilley, James	Jim.Lilley@bigpond.com 18/19 Delamere Ave South Perth, WA 6151 Australia Tel: +61 (08) 93682114
7	Nunn, Michael	Mike.Nunn@affa.gov.au Animal Health Science Office of the Chief Veterinary Officer Agriculture, Fisheries and Forestry – Australia GPO Box 858 Canberra ACT 2601 Tel: + 02 6272 4036 Fax: + 02 6272 4533
8	O'Connor, Paul Francis	New South Wales Aquaculture & Fisheries, (Fisheries Division), P.O. Box K220, Haumarket NSW 2000
9	Owens, Leigh	leigh.owens@jcu.edu.au Dept. Microbiology & Immunology, P.O. James Cook University, Townsville, Qld 4811 Tel:+ 07 4781 4632 Fax:+ 07 4779 1526

10	Perera, Ramesh	Ramesh.Perera@affa.gov.au Biosecurity Australia Department of Agriculture, Fisheries and Forestry - Australia GPO Box 858, Canberra ACT 2601 Tel: +61 2 6272 4675 Fax: +61 2 6272 3399
11	Roper, Katrina	katrina.roper@bigpond.com Dept. Microbiology & Immunology, James Cook University, Townsville, Qld 4811
12	Trott, Lindsay Alexander	Australian Institute of Marine Science, PMB No. 3, MSO, Townsville 4810
13	Walker, Peter	Peter.Walker@csiro.au CSIRO Livestock Industries 120 Meiers Road Indooroopilly Q 4048 AUSTRALIA Tel. 61-7-3214-2758 Fax. 61-7-3214-2718 Mob. 04-1707-2603
14	Warner, Lesley	Biology Department, University College of Central Queensland, Rochhampton, Qld 4702
15	Whittington, Ian	whittington.ian@saugov.sa.gov.au Parasitology Section, The South Australian Museum North Terrace, Adelaide, South Australia 5000 AUSTRALIA Tel: 08-8207-7463 Fax: 08-8207-7222

BANGLADESH

16	Ahmed, Abu Tweb Abu	zooldu@citechco.net Department of Zoology University of Dhaka Dhaka – 1000, Bangladesh Tel: 880-2-9666120 (Office); 880-2-606690 (Res) Fax: 880-2-8615583
17	Chowdhury, Md. Bazlur Rashid	mbrchow@royalnet.net Department of Fisheries, Biology & Limnology, Bangladesh Agricultural University, Mymensingh 2202 Tel: 091-5695-7 ext. 281
18	Kumar, Dilip	dilip@gshakti.com dilip7kumar@hotmail.com Empowerment of Coastal Fishing Communities for Livelihood Security (UNDP/FAO Project: BGD/97/017) ADB Hatchery Campus Cox's Bazaar, Bangladesh Tel/Fax: (88-0341) 63871

19 Hossain, Adul	hossain@atdp.biz Fisheries Consultant ABM Tower, Plot # 8, Room # 113A Dhaka 121 Bangladesh Tel: 880-2-9882009 Fax: 880-2-9825625	28 Hardy-Smith, Paul	paulh-s@mars.ark.com 201-1155 England Avenue Courtenay, British Columbia V9N 2N9 Canada
20 Khan, Masud H.	mhk@bol-online.com Bangladesh Fisheries Research Institute Mymensingh 2201 Bangladesh Fax: 880-915-5259	29 Kabata, Zbigniew (retired)	zkabata@shaw.ca Pacific Biological Station Nanaimo, B.C. Canada V9T 6N7
21 Khondaker, Modabbir Ahmed	Pioneer Hatchery Ltd., Zilla Parishad Bhaban, Cox's Bazar	30 McGladdery, Sharon	McGladderyS@dfo-mpo.gc.ca Department of Fisheries and Oceans Canada Gulf Fisheries Centre Box 5030 Moncton, NB, E1C 9B6 Tel: 1-506-851-2018 Fax: 1-506-851-2079
BELGIUM		CHINA PR	
22 Alday de Graindorge, Victoria	victoria_alday@yahoo.com Avenue de l'Aurore 8 1330 Rixensart, Belgium Tel: 32 2 6539409 Fax: 32 2 6539409 or INVE Technologies Oeverstraat 7, 9200 Baasrode, Belgium Tel: 32 52 331320 Fax: 32 53 334531	31 Jiang, Gui Zhen	Fish Disease Laboratory, NACA-APRRTC(Integrated Farming), Wuxi, Jiangsu Province
23 Ollevier, Frans	K. U. Leuven Zoological Institute, Naamsestraat 59, B-3000 Leuven (Louvain), Belgium	32 Jiang, Yulin	szapqbx@163.net Shenzen Exit and Entry Inspection and Quarantine Bureau 40 Heping Road Shenzhen 518010 China PR Tel: 86-755-5592980 Fax: 86-755-5588630
CANADA		33 Liao, Hsianghua	Department of Biology, Zhongshan University, Guangzhou, Guangdong
24 Arthur, James Richard	rarthur@titanlink.com 6798 Hillside Drive, Sparwood, B.C. Canada V0B 2G3 Tel: (250) 425 2287	34 Pan, Jin-Pei	Institute of Hydrobiology, Academia Sinica, Wuhan, Huhei Province
25 Bower, Susan	BowersS@dfo-mpo.gc.ca Fisheries and Oceans Canada Pacific Biological Station 3190 Hammond Bay Road Nanaimo, British Columbia V9T 6N7 Canada Tel: 250-756-7077 Fax: 250-756-7053 http://www.pac.dfo-mpo.gc.ca/sci/shelldis/title_e.htm	35 Shen, Yalin	Mariculture Research Division, East China Sea Fisheries Research Inst., 300 Jung Gong Rd. 200090 Shanghai
26 Castell, John Danel	castellj@mar.dfo-mpo.gc.ca Department of Fisheries & Oceans, 531 Brandy Cove Road, St. Andrews, New Brunswick E5B 2L9	36 Yang, Xiang-Le	Xlyang@shfu.edu.cn Fish Diseases Division Shanghai Fisheries University 334 Jungong Rd. Shanghai, China PR Tel: +86-21-65710870 Fax: +86-21-65687210
27 Ellana, Eduardo	22 Lucas Avenue, Winnipeg, Manitoba R2R 2H9 Canada	37 Yang, Jifang	jkfwlq@public1.h2.zj.cn Second Institute of Oceanography, SDA P.O. Box 1207, Hangzhou 310012, China PR Fax: 0086-1571-8071539
		38 Zhang, Lan-Tao	Chinese Academy Aquatic Science, Freshwater Fisheries Research Centre, Wuxi, Jiangsu Province

DENMARK

39 McLean, Ewen
Aquaculture Biotechnology
Laboratory
Aalborg University
Sohngardsholmsvej 57
9000 Aalborg Denmark

FRANCE

40 De Kinkelin, Pierre
(retired)
kinkelin@biotec.jouy.inra.fr
INRA, Molecular Virology
& Immunology Unit,
Ichthyopathology, 78352,
Jouy-en, Josas, Cedex

GERMANY

41 Edgerton, Brett
brettedgerton@hotmail.com
Alexander von Humboldt
Research Fellow
Department of Zoology,
Fish Biology and Fish Diseases
University of Munich
Kaulbackstr. 37 Munich 80539
Germany

42 Korting, Wolfgang
wkoert@fisch.tiho-hannover.de
Fish Diseases Research Unit,
School of Veterinary Medicine,
Bunteweg 17, 30559 Hannover
Fed. Rep. of Germany
Tel: 0049 511 953 8501
Fax: 0049 511 953 8587

43 Taraschewski, Horst
dc20@rz.uni-karlsruhe.de
Zoological Institute
Univ. Karlsruhe
Kornblumenstr. 13
D-76128 Karlsruhe Germany

HUNGARY

44 Jeney, Zsigmond
jeneyz@haki.hu
Research Institute for Fisheries
Aquaculture and Irrigation
H-5540 Szarvas Anna liget 8, Hungary
Tel: +36-66-515 314
Fax: +36-66-312 142
Mobile: +36-70-299-0137
Webpage: www.haki.hu

INDIA

45 Chandran, A.
Department of Aquatic Biology
& Fisheries,
University of Kerala, Beach P.O.,
Trivandrum 695 007

46 Damarla, Raghava Rao
Kohinoor Hatcheries Pvt. Ltd.,
Plot No.6,
Journalist Colony 8-2-248-B2,
Rd.3Banjara Hill, Hyderabad 500 034

47 Das, Sanjay Kumar
College of Fisheries,
Assam Agricultural University,
Raha, Nagaon, Assam 782 103

48 Dey, R. K.
FARTC (CIFRI), PO Kausalyagang
via Bhubaneshwar, 751 002 Orissa

49 D'Souza,
Aveline Ronald
St. Aloysius Evening College,
P.B. 720, Mangalore 575 003

50 Goswami, Umesh C.
c/o Dr. Alaka Goswami,
Upendranath Bezbarua Road,
Silpukhuri, Guwahati 781 003, Assam

51 Jeyachandran,
David Jason
Marine Technologies,
43-B, M.G. Road,
Shastri Nagar, Madras 600 041

52 Kalaiselvam,
Morugaiyan
casenivis@yahoo.com
Marine Biology
Parangipettai 608502 India
Tel: 91-4144 483223
Fax: 91-4144 43555
Annamalai University,
Parangipettai 608 502

53 Kandula, Sujatha
Dept. Marine Living Resources,
Andhra University,
Visakhapatnam 530 003

54 Karunasagar, Iddya
College of Fisheries, PB No. 527,
University of Agricultural Sciences,
Mangalore 575 002, Karnataka

55 Karunasagar, Indrani
mircen@sancharnet.in
Department of Fishery Microbiology
University of Agricultural Sciences
College of Fisheries
Mangalore 575 002 Karnataka

56 Kollanoor, Riji John
ttn_rijjohn@sancharnet.in
Virology Unit
Department of Aquaculture
Fisheries College and
Research Institute
Tuticorin 628 008 India
Tel: +91-461-2340554
(Off)/2311792 (Res)
Fax: +9-461-2340574

57 Kumar, Kuldeep
FARTC (CIFRI), PO Kausalyagang via
Bhubaneshwar, 751 002 Orissa

58 Kurcheti, Pani Prasad
Division of Fish Microbiology
& Pathology, Central Institute
of Fisheries Education,
Seven Bungalows, Versova,
Mumbai 400 061

59 Mishra, Safal Kumar
"Yashoda Lok",
Mahatma Gandhi Marg,
Narkatiaganj 845 455,
W. Champaran, Bihar

60 Mitra, Arunabha
Dept. Aquaculture &
Food Engineering,
Indian Institute of Technology,
Kharapur Pin 721 302

61 Mohan, C. V.	cv_mohan@yahoo.com Fish Pathology Laboratory Department of Aquaculture College of Fisheries Mangalore 575 002 India Tel: 91-824-434356 Fax: 91-824-434356/440395	72 Shankar, K. M.	Fish Pathology Laboratory Department of Aquaculture College of Fisheries Mangalore 575 002 India Tel: 91-824-434356 Fax: 91-824-434356/440395
62 Muley, Dipak Vishwanathrao	Dept. of Zoology, Shivaji University, Kolhapur 416 004 M.S.	73 Sharma, Jai Gopal	Department of Zoology, University of Delhi, Delhi 110007
63 Mulloorpeedikayil, Rosalind George	rosalindmg@yahoo.com Department of Aquaculture, Fisheries College, Tamil Nadu Vet. & Animal Science University, Tuticorin 628 008 India Tel: +91-461-2340554 Fax: +91-461-2340574	74 Singh, Akhilesh Kumar	Department of Biology Ranchi Veterinary College Ranchi 834 007
64 Pal, Joydeb	pajoydeb@rediffmail.com Dept. Zoology, Centre for Life Sciences, North Bengal Univ. P.O. North Bengal Univ., Dt. Danjeeling 734 430, West Bengal Tel: 091353 582124 (O), 091353 581854 (R) Fax: 091353 581546 Web:www.nbu.ac.in	75 Tahir, Akbar	Faculty of Marine and Fisheries Science, Hasanuddin University
65 Pal, Radhanath	Central Inland Fisheries Research Institute, Barrackpore, West Bengal 743 101	76 Tamilmami, V.	Estuarine Fish Farm, Portonovo 608 502, Tamil Nadu
66 Padijar, Arun Subhaschandra	arun_padijar@rediffmail.com Padiyar Nivas Main Road Panemangalore - 574231 Karnataka State, India Tel : 91 824 340698 (Residence)	77 Thomas, Shirley	29/2325 Valloor Road, Poonithura P.O., Cochin 682 317
67 Parthasarathy, Chandra Mohan	Andhra University, Waltair, Visakhapatnam 530 003	78 Veerina, Syama Sundar	C/o Dr. U.K. Rao Near Wahab Park, Tenali 522 201, Guntur (DT) AP
68 Patterson Edward, J. K.	Centre of Advanced Study in Marine Biology, Annmalai University, Parangipettai 608 502, Tamil Nadu	INDONESIA	
69 Radhakrishnan, K. V.	College of Fisheries, University of Agricultural Sciences, Mangalore 575 002, Karnataka	79 Angka, Sri Lestari	Faculty of Fisheries Bogor Agricultural University Bogor, Indonesia
70 Radhakrishnan, S.	Department of Aquatic Biology & Fisheries, University of Kerala, Beach P.O., Trivandrum 695 007, Kerala	80 Arifin, Zainal	Cent. Oceanological Research & Development, Balitbang-SDL-LIPI, Guru-Guru, Poka Ambon
71 Seenappa, D.	University of Agricultural Sciences Fisheries Research Station Hessaraghatta Bangalore 560 089	81 Bastiawan, Dayat	Balai Penelitian Perikanan Airtawar, Jalan Sempur No.1, P.O. Box 51, Bogor
		82 Haliman-Rubiyanto, Widodo	Pt. Tirta Mutiara Makmur P.O. Box 14 Besuki-Situbondo 68356 Indonesia Tel: 62-338-891286 Fax: 62-338-891286
		83 Haluan, John	Fakultas Perikanan, Komplek IPB-Dramaga, Bogor
		84 Komarudin, Oman	Jln. Sempur 1, P.O. Box 51, Bogor, Indonesia
		85 Mangunwiryo, Hariyadi (retired)	Res. Inst. Veterinary Science, Jl. R. E. Martadinata 32, Bogor
		86 Rukyani, Akhmad	dfhe@indosat.net.id Directorate of Fish Health and Environment Directorate General of Aquaculture Ministry of Fisheries and Marine Affairs Jl. Harsono R.M. No. 3 B B12-VI Ragunan, Pasar Minggu Jakarta 12550 Indonesia Tel: 62-021-7827844 Fax: 62-021-7827844/78835853

87 Sukandi, Prajitna	wiu1993@rad.net.id PT Minaut Lintas Nusantara Bojonegara Research Station for Coastal Aquaculture 2H, Gunung Sahari Raya Jakarta 10720, Indonesia Fax: +62-21-629-2669	96 Erlambang, Tanza	Fisheries Faculty, Nagasaki University, 1-14 Bungkyo-machi, Nagasaki 852
88 Supriyadi, Hambali	Fish Health Research Laboratory Jl. Ragunan 20, Pasar Minggu Jakarta - Indonesia Tel: 62-21-7805052 Fax: 62-21-7815101	97 Hatai, Kishio	hatai@scan-net.ne.jp Division of Fish Diseases Nippon Veterinary & Animal Science University 1-7-1 Kyonan-cho, Musashino Tokyo 180-8602 Japan Tel: 81-422-31-6796 Fax: 81-422-31-6796
89 Tahir, Akbar	Faculty of Marine and Fisheries Science, Hasanuddin University, Indonesia	98 Itami, Toshiaki	itami@fish-u.ac.jp National Fisheries University 2-7-1, Nagata-honmachi Shimonoseki, Yamaguchi Japan 759-6595 Tel: 81-832-86-5111 ext. 466 Fax: 81-832-86-7435
90 Zafran	Gondol Research Station for Coastal Fisheries P.O. Box 140 Singaraja Bali, Indonesia Tel: 0362-92270 Fax: 0362-92272	99 Kimura, Takahisa (retired)	takak@fish.hokudai.ac.jp takak624@ms3.ncv.ne.jp Faculty of Fisheries, Hokkaido University 6-25 Enomoto-cho, Hakodate, 0420931 Japan Tel./Fax: 81-138-57-6411
IRAN		100 Kusuda, Riichi (retired)	Kochi University, Fish Disease Laboratory, Nankoku, Kochi 783
91 Fakour Motlagh, Hassan	Modje Rajaj Co., No. 4 Laleh St., Kargar Avenue, Tehran	101 Matsusato, Toshihiko	Matusato@fra.affrc.go.jp National Research Institute of Aquaculture (NRIA) Fisheries Research Agency Nakatsuhama-ura, Nansei Mie 516-0193 Japan
92 Sasani, Farhang	University of Tehran, Iran	102 Miharu, Toshio	Fisheries & Aquaculture Intl. Co., Inc., No. 7 Kohji-machi Bldg., Rm B105, 4-5 Kohji-machi, Chiyoda-ku, Tokyo
ITALY		103 Miyazaki, Teruo	miyazaki@sansui.bio.mie-u.ac.jp Faculty Bioresources Mie University 1515 Kamihana, Tsu Mie, Japan Tel: +81-59-231-9532 Fax: +81-59-231-9532
93 Subasinghe, Rohana Padmabandu	Rohana.Subasinghe@fao.org Inland Water Resources and Aquaculture Service Fishery Resources Division, Fisheries Department Food and Agriculture Organization of the UN Viale delle Terme di Caracalla 00100 Rome, ITALY Tel: + 39 06 570 56473 Fax: + 39 06 570 53020	104 Morizane, Tsuneo	Ehime Prefectural Fish. Exptl. Station, 5516 Shitaba, Uwajima City 798-01
ISRAEL		105 Muroga, Kiyokuni	fpath@hiroshima-u.ac.jp Faculty of Applied Biological Science Hiroshima University Higashi-Hiroshima 739-8528 Tel: +81-824-24-7977 Fax: +81-824-22-7059
94 Paperna, Ilan	paperna@agri.huji.ac.il Department of Animal Sciences Faculty of Agricultural, Food & Environmental Quality Sciences, University of Jerusalem Rehovot 67100, Israel	JAPAN	
95 Aoki, Takashi	aoki@tokyo-u-fish.ac.jp Laboratory of Genetics & Biochemistry Department of Aquatic Bioscience Tokyo University of Fisheries Konan 4-5-7, Minato Tokyo 108-8477 Japan Tel: 03-53463-0556 Fax: 03-5463-0690		

106 Ogawa, Kazuo	aogawak@mail.ecc.u-tokyo.ac.jp Laboratory of Fish Diseases Department of Aquatic Bioscience Graduate School of Agricultural & Life Sciences The University of Tokyo Yayoi, Bunkyo, Tokyo 113-8657 Tel: +81-3-5841-5282 Fax: +81-3-5841-5283	115 Kua Beng Chu	bengchu@hotmail.com kuaben01@dof.moa.my Fish Health Research Center Fisheries Research Institute Department of Fisheries 11960 Batu Maung, Penang, Malaysia Fax: 604-6263977
107 Okamoto, Nobuaki	nokamoto@tokyo-u-fish.ac.jp Department of Aquatic Biosciences, Tokyo University of Fisheries, Konan 4, Minato-ku, Tokyo 108	116 Chuah, Toh-Thye	chuahtt@rocketmail.com National Prawn Fry Production & Research Centre Kampong Pulau Sayak, 08500 Kota Kuala Muda Kedah Darul Aman Tel: 04 4374021 Fax: 04-4374470
108 Pakingking, Rolando	roland@hiroshima-u.ac.jp Faculty of Applied Biological Science Hiroshima University Higashi-Hiroshima 739-8528 or 739-0036 Hiroshima ken, Higashi Hiroshima shi, saijo cho, Taguchi 451-7 College Court rm 209 Tel: (81) 090-6404-1129	117 Jintoni, Boniface	Fisheries Department, 8th Flr., Menara Khidmat, 82628 Kota Kinabalu, Sabah
109 Sakai, Masahiro	m.sakai@ikoma.cc.miyazaki-u.ac.jp Faculty of Agriculture, Miyazaki University, Nishi 1-1, Gakuen-Kibanadai, Miyazaki 889-21	118 Leong, Tak-Seng (retired)	mhpg@pc.jaring.my No. 3 Cangkat Minden Lorong 13 11700 Glugor Pulau Pinang Malaysia
KOREA RO		119 Lim, Susan Lee-Hong	susan@umcsd.um.edu.my Institute of Biological Sciences Institute of Postgraduate Studies & Research, University of Malaya 50603 Kuala Lumpur, Malaysia Tel: 603-7594502 Fax: 603-7568940
110 Choi, Kwang Sik	skchoi@cheju.cheju.ac.kr School of Applied Marine Science, Cheju National University, 1 Ara 1 Dong Cheju, Cheju 690756	120 Nagaraj, Gopinath	fani@tm.net.my FanLi Marine & Consultancy, Sdn. Bhd. 27-3 Block F-2, Dataran Prima, Jln. PJU 1/42A, 47301, Petaling Jaya
111 Jung, Sung-Ju	sungju@yosu.ac.kr Department of Fish Pathology, Yosu National University, Yosu 550-749	121 Ong, Bee Lee	ong@jph.gov.my Regional Veterinary Laboratory Services Department of Veterinary Services 8th & 9th Fl, Wisma Chase Perdana Jln Semantan 50630 Kuala Lumpur, Malaysia Tel: 60-3 254 0077 ext. 173 Fax: 60-3 254 0092/253 5804
112 Oh, Myung-Joo	ohmj@yosu.ac.kr Department of Fish Pathology, Yosu National University, San 96-1 Dunduk-dong, Yeosu City Jeollanam-do, Korea 550-749 Tel: 82-61-659-3173 Fax: 82-61-659-3173	122 Palanisamy, Vello	Fish Health Research Center Fisheries Research Institute Department of Fisheries 11960 Batu Maung, Penang, Malaysia Fax: 604-626397
KUWAIT		123 Saidin, Thalathiah Bte	thalathiah@hotmail.com Department of Fisheries, Ministry of Agriculture Malaysia 8th Fl., Wisma Tani, Jalan Sultan Salahuddin, 50628 Kuala Lumpur Tel: 603-2617 5616 D/L 603-2617 5614 (Main) Fax: 603-2698 0227
MALAYSIA			
114 Abdullah, Zahrah	Fish Health Research Center Fisheries Research Institute Department of Fisheries 11960 Batu Maung, Penang, Malaysia Fax: 604-6263977		

124 Sentian, Justin
jsentian@hotmail.com
Department of Environmental Science
School of Science and Technology
University of Sabah Malaysia
Locked Bag 2073
88999 Kota Kinabalu
Sabah, Malaysia
Tel: 088 320000 ext 5757
Fax: 088 435324

125 Shaharom, Faizah
faizah@uct.edu.my
University College of Science &
Technology Malaysia
Mengabang Telipot
21030 Kuala Terengganu, Malaysia
Tel: 609-6683243
Fax: 609-6686441

126 Shamsudin,
Mariana Nor
mariana@medic.upm.edu.my
Universiti Putra Malaysia
Faculty of Medicine & Health Science
UPM 43400 Serdang,
Selangor, Malaysia
Tel: 006039486101 ext. 2537
Fax: 0060394269571/033426246

127 Shariff, Mohamed
shariff@vet.upm.edu.my
Faculty of Veterinary Medicine
Universiti Putra Malaysia
43400 Serdang, Selangor Darul
Ehsan, Malaysia
Tel: 60-3-89488246
Fax: 60-3-89488246

128 Wong, See Yong (retired)

MAURITIUS

129 Jayabalan, Nachiappan ITEC-Expert, Albion Fisheries
Research Centre, Albion Petite Riviere

NEW CALEDONIA

130 Lambeth, Lyn
LynL@spc.int
Secretariat of the Pacific Community,
BP D5 98848, Noumea Cedex

NEW ZEALAND

131 Hine, Mike
hinem@maf.govt.nz
Ministry of Agriculture and Fisheries
New Zealand
Tel: +64-4-526-5600
Fax: +64-4-526-5601

NORWAY

132 Hastein, Tore
Tore.Hastein@vetinst.no
National Veterinary Institute
Ullevalsveien 68
P.O. Box 8156 Dep. 0033 Norway
Tel: 47 22964710
Fax: 47 22463877

PAKISTAN

133 Ataur-Rahim,
Mohammed
National Agriculture Research Center,
Animal Science Institute,
Park Rd., Islamabad

134 Iqbal, Mohammad
C-29 Staff Town,
Karachi University Campus,
Karachi 75270

135 Mian,
Muhammad Javed
Department of Zoology & Fisheries,
University of Agriculture, Faisalabad

136 Khan, Hanif
Khakan Enterpriese,
Fish Hatchery Project,
27 Upper Mall Lahore-15

PAPUA NEW GUINEA

137 Lokani, Paul
Fisheries Research Station,
P.O. Box 337, Kavieng

138 Richards, Andrew Hick
Research & Surveys Branch, Dept. of
Fisheries & Marine Resources,
P.O. Box 165, Konedobu

PHILIPPINES

139 Abello, Isaac
SEAFDEC Aquaculture Department
Tigbauan 5021, Iloilo, Philippines
Tel: (63-33) 335 1009
Fax: (63-33) 335 1008

140 Albaladejo, Juan D.
jalbaladejo99@yahoo.com
Fish Health Section, Bureau of
Fisheries & Aquatic Resources
Arcadia Building, 860 Quezon Avenue
Quezon City, Philippines
Tel: (632) 372 5055
Fax: (632) 372 5055

141 Alegre, Juan
argentph@compass.com.ph
Labcare Research Laboratories,
#6 Ranger St.,
Moonwalk, Las Pinas, Metro Manila

142 Apostol, Maria Abegail
mariaabegail11@yahoo.com
Fish Health Section, Bureau of
Fisheries & Aquatic Resources
Arcadia Building, 860 Quezon Avenue
Quezon City, Philippines
Tel: (632) 372 5055
Fax: (632) 372 5055

143 Bantaya, Mercedita A.
Fish Health Section, Bureau of
Fisheries & Aquatic Resources
Arcadia Building, 860 Quezon Avenue
Quezon City, Philippines
Tel: (632) 372 5055
Fax: (632) 372 5055

144 Bayados, Isaac
PBSP Samar Field Office,
Brgy. New Mahayag,
Catbalalogan, Samar

145 Bigueras-Benetiz, Carmelita	Institute of Fish Processing Technology, University of the Philippines in the Visayas, Miag-ao, Iloilo	155 Del Mundo, Rosario	BFAR Region 4 2/F Infrastructure Computer Center (ICC) Bldg., NIA Complex EDSA, Diliman, Quezon City Tel: 63 2 922 2225 Fax: 63 2 926 8616
146 Borromeo, Emilio	First Asian Universal Industries Corp., No. 1 Fabian Dela Rosa St., Loyola Heights, Quezon City	156 Dionisio, Edna	fkphils@infocom.ph GIFT Foundation Intl. Inc., CLSU Compound, Science City of Munoz, Nueva Ecija
147 Calanoga, Esterlita	Cagayan State University, College of Fisheries, Aparri, Cagayan	157 Duray, Marietta N.	mayet@aqd.seafdec.org.ph SEAFDEC Aquaculture Department, P.O. Box 256, Iloilo City, Philippines Tel: (63-33) 335 1009 Fax: (63-33) 335 1008
148 Casten, Lemuel	l.casten@cgjar.org ICLARM Philippine Outreach Office, Khush Hall, IRRI, College, Laguna, Philippines Tel: (63-32) 845-0563 Ext. 6858 Fax: (63-32)891-1292	158 Gacutan, Rogelio Q.	rgacutan@yahoo.com Fish Health Section SEAFDEC Aquaculture Department Tigbauan 5021, Iloilo, Philippines Tel: (63-33) 3362965 Fax: (63-33) 3351008 www.seafdec.org.ph
149 Catap, Elena	elenacatap@yahoo.com esc@aqd.seafdec.org.ph Fish Health Section SEAFDEC Aquaculture Department Tigbauan 5021, Iloilo, Philippines Tel: (63-33) 3362965 Fax: (63-33) 3351008 www.seafdec.org.ph	159 Gaerlan, Rosario Segundina	BFAR Region 1 Pagdalagan Nort, San Fernando City La Union, Philippines Fax: 242-1559
150 Chua, Wilma	CSU-Aparri College of Fisheries, Maura, Aparri, Cagayan, Philippines	160 Gerundo, Nelson D.	chiqui.g@delifrance.com.ph 425 Sikatuna St. Barangay Village Paranaque City, Philippines 1700 Tel: 632-826-8404 Mobile: 0917-645-7675
151 Cruz-Lacierda, Erlinda R.	eclacier@aqd.seafdec.org.ph Fish Health Section SEAFDEC Aquaculture Department Tigbauan 5021, Iloilo, Philippines Tel: 63-33-335-1009 Fax: 63-33-511-9070 www.seafdec.org.ph	161 Gomez, Dennis K.	Bacolod Oceanic Hardware Libertad Ext., Bacolod City Negros Occidental Current address: Faculty of Applied Biological Science Hiroshima University Higashi-Hiroshima 739-8529 Tel: +81-824-24-7477 Fax: +81-824-22-7059
152 Cuvin-Aralar, Ma. Lourdes	mlcaralar@aqd.seafdec.org.ph SEAFDEC Aquaculture Department Binangonan Freshwater Station Binangonan, Rizal 1940 E-mail??	162 Herrera, Annabelle	aah@nib.upd.edu.ph College of Science, University of the Philippines, Diliman, Quezon City
153 De La Cruz, Rudolph Elmo	Bureau of Fisheries & Aquatic Resources (BFAR) Regional Office No. 6 Molo, Iloilo, Philippines 5000 Tel: 63-33 336 6748 Fax: 63-33 336 9432	163 Jailani, Mohammad	Training & Communications Division, BFAR Region XI, Davao City, Philippines
154 De la Pena, Leobert	leobertd@aqd.seafdec.org.ph Fish Health Section SEAFDEC Aquaculture Department Tigbauan 5021, Iloilo, Philippines Tel: (63-33) 3362965 Fax: (63-33) 3351008 www.seafdec.org.ph	164 Jimenez, Buenafior	benben@biocsm.msuiit.edu.ph Department of Biological Sciences Mindanao State University Iligan Institute of Technology, Tibanga, Iligan City, Philippines Tel: (63-63)2214050 to 55 loc 137 Fax: (63-63)2214068

165 Ladja, Jocelyn	joladja@seafdec.org.ph SEAFDEC Aquaculture Department Tigbauan 5021, Iloilo, Philippines Tel: (63-33) 3362965 Fax: (63-33) 3351008	173 Mateo, Dennis	BFAR, Bonuan-Binloc Dagupan City, Philippines Fax: 63-75-523 0385
166 Landoy, Remus J.	rjlandoy@pacific.net.ph RJ Landoy & Associates Inc., 9574 Sgt. Fabian Yabut Circle Guadalupe, Makati City Tel: (63-32) 882 1291	174 Mercado, Ma. Antonia	yayanmer@yahoo.com # 14 Don Rafael Street, Don Enrique Heights, Commonwealth Avenue, Quezon City Philippines Tel: (63-32) 931 5550
167 Landoy, Romulus J.	rjlandoy@pacific.net.ph RJ Landoy & Associates Inc., 9574 Sgt. Fabian Yabut Circle Guadalupe, Makati City Tel: (63-32) 882 1291	175 Miranda, Rolando	2/F Infrastructure Computer Center (ICC) Bldg., NIA Complex EDSA, Diliman, Quezon City
168 Lavilla-Pitogo, Celia	celiap@aqd.seafdec.org.ph Fish Health Section SEAFDEC Aquaculture Department Tigbauan 5021, Iloilo, Philippines Tel: (63-33) 3362965 Fax: (63-33) 3351008 www.seafdec.org.ph	176 Natividad, Carlo Dante	kdt@biotech.uplb.edu.ph kdt@natividad@hotmail.com BIOTECH, UPLB College, Laguna 4031 Philippines Fax: 63-49 536-2721
169 Leano, Eduardo	eduardo@mail.tfrin.gov.tw Taiwan Fisheries Research Institute 199 Hou-lh Road, Keelung 202 Taiwan Tel: +886-2-2462-2101 loc 3508 Mobile: +886-930-534286 Fax: +886-2-2642-3306	177 Natividad, Jose M.	joemn@info.com.ph
170 Lio-Po, Gilda	liopo@aqd.seafdec.org.ph Fish Health Section SEAFDEC Aquaculture Department Tigbauan 5021, Iloilo, Philippines Tel: (63-33) 3362965 Fax: (63-33) 3351008 www.seafdec.org.ph	178 Nasar, Syed Shams Tabrez	Tabrez.Nasar@iirr.org tabreznasar@yahoo.com Agriculture & Natural Resources Management International Institute for Rural Reconstruction YC James Yen Center 4118 Silang, Cavite, Philippines Tel: (63-46) 4142417 Fax: (63-46) 414 2420
171 Lopez, Nellie	ncl@nib.upd.edu.ph Institute of Biology, College of Science University of the Philippines Diliman, Quezon City 1101, Philippines Tel: 632-9205301 loc. 6452 TeleFax: 632 920-54-71	179 Nieves, Plutomeo	Bicol University, College of Fisheries, Tabaco, Albay
172 Lumanlan-Mayo, Susan	slmayo99@yahoo.com Susan.Mayo@fysiology.uu.se Fish Health Section, Bureau of Fisheries & Aquatic Resources Arcadia Building, 860 Quezon Avenue Quezon City, Philippines Tel: (632) 372 5055 Fax: (632) 372 5055	180 Paclibare, Jose O.	jopac@edsamail.com.ph Fish Health Section Bureau of Fisheries & Aquatic Resources Arcadia Building, 860 Quezon Avenue Quezon City, Philippines Tel: (632) 372 5055 Fax: (632) 372 5055
Current Address:	Division of Comparative Medicine Department of Physiology Uppsala University Biomedical Centre Box 570 751 23 Uppsala, Sweden Tel: +46 18 471 4422 Fax: +46 18 501740	181 Pagador, Gregoria	gop@seafdec.org.ph Fish Health Section SEAFDEC Aquaculture Department Tigbauan 5021, Iloilo, Philippines Tel: 335-1009 Fax: (63-33)335-1008 (63-33) 5119070 www.seafdec.org.ph
		182 Palisoc, Fermin	ferminpjr@hotmail.com Pangasinan State University Binmaley, Pangasinan, Philippines
		183 Pascual, Alberto B.	School of Fisheries, Mariano Marcos State University, Currimao, Ilocos Norte 2903

184 Pilapil-Germano, Bernadita	Visayas State College of Agriculture, Visca, Baybay, Leyte	194 Torres, James L.	jamesitorres@philwebinc.com jltorres@easycom.net Department of Biological Science College of Arts and Sciences University of the Philippines in the Visayas Miag-ao 5023, Iloilo, Philippines Tel: (63) 33 3158160 (office) 3365837 (res) Fax: (63) 33 3381534
185 Receno, Melinda	Seafarming Research & Development Center, Bonuan, Binloc, Dagupan City, Pangasinan	195 Yambot, Apolinario	polyambot@yahoo.com College of Inland Fisheries, Central Luzon State University, Munoz, Nueva Ecija
186 Regidor, Simeona	Fish Health Section Bureau of Fisheries & Aquatic Resources Arcadia Building, 860 Quezon Avenue Quezon City, Philippines Tel: (632) 372 5055 Fax: (632) 372 5055	POLAND	
187 Ricon, Ruther	INTEL Technology Phil., Inc., Gateway Business Park, Cavite	196 Rokicki, Jerzy	Department of Invertebrate Zoology, University of Gdansk, Al. Pilsudskiego 46, 81-378 Gdynia
188 Salvador, Ronelie	University of Eastern Philippines, University Town, Catarman, Northern Samar Current Address: College of Fisheries, University of the Philippines in the Visayas Miag-ao, Iloilo, Philippines	SAUDI ARABIA	
189 Santos, Arsenio	# 1 Ifugao Street, La Vista Subdivision, Quezon City	197 Abuan, Espiritu F.	King Abdulaziz City for Science & Technology., Fish Culture Project, P.O. Box 6086, Riyadh 11442
190 Sim, Alberto	N Royal Circle Townhouse, T. Alonso Street, Baclaran, Paranaque Tel: (63-32) 832 1497	198 Al-Harbi, Ahmed	Research Inst. Natural Resources & Environment, King Abdulaziz City for Science & Technology, P.O. Box 6086, Riyadh 11442
191 Somga, Joselito	jsomga@edsamail.com.ph National Coordinator on Aquatic Animal Health Fish Health Section Bureau of Fisheries & Aquatic Resources Arcadia Building, 860 Quezon Avenue Quezon City, Philippines Tel: (632) 372 5055 Fax: (632) 372 5055	SINGAPORE	
192 Somga, Sonia	sssomga@edsamail.com.ph Fish Health Section Bureau of Fisheries & Aquatic Resources Arcadia Building, 860 Quezon Avenue Quezon City, Philippines Tel: (632) 372 5055 Fax: (632) 372 5055	199 Chua, Sek Chuan	01-02 Lincolnsvale, 22 Surrey Road Singapore 1130
193 Tendencia, Eleonor	gigi@aqd.seafdec.org.ph Fish Health Section SEAFDEC Aquaculture Department Tigbauan 5021, Iloilo, Philippines Tel: (63)(33)335-1009 Fax: (63)(33)336-1008 www.seafdec.org.ph	200 Grisez, Luc	Luc.Grisez@intervet.com Intervet Norbio Singapore Pte Ltd 1 Perahu Road, Singapore 718847 Tel: + 65 6 397 1121 Fax: + 65 6 397 1131
		201 Ho, Ellen	Ellen.Ho@intervet.com Intervet Norbio Singapore Pte Ltd 1 Perahu Road, Singapore 718847 Tel: 65 63971121 Fax: 65 63971131
		202 Lee, Tat-Mong Monty	Rhone Merieux Asia Pacific Pte Ltd., #04-02 The Mendel, 12 Science Park Drive, Singapore Science Park Singapore 0511
		203 Low, Kim-Wah	PhD Student, Dept. of Zoology, National University of Singapore, Lower Kent Ridge Road Singapore 0511
		204 Low, Wai-Ping	Freshwater Fisheries Section, Sembawang Field Experimental Station, 17 Km. Sembawang Rd. Singapore 2776

205 Tan, Zilong
Zilong.Tan@intervet.com
Intervet Norbio Singapore Pte. Ltd.,
1 Perahu Road, Singapore 718847
Tel: +65 6397 1121
Fax: +65 6397 1131

SOUTH AFRICA

206 Crampton, Margaret
JLB Smith Inst. of Ichthyology,
PTE Bag 1015, Grahamstown

207 De Wet, Lourens
University of Stellenbusch,
Aquaculture Research Program,
P.B. XI Matieland 7602

SRI LANKA

208 Ariyaratne, Soma
National Aquatic Resources Research
& Development Agency,
Mattakkuliya, Colombo 15

209 Balasuriya, L. K. S. W.
kamalbalasuriya@yahoo.com
lksw@dialogsl.net
Koshenawatta, Kendalanda
Pallewela, Sri Lanka

210 Cheong, Chin How
Ceylon Grain Elevators Ltd.,
15 Rock House Lane, Colombo 15

211 Mahinda, Kulatilake
hamkulathilaka@yahoo.com
National Aquatic Resources Research
& Development Agency
Ministry of Fisheries &
Ocean Resources
Crow Island, Mattakkuliya,
Colombo -15, Sri Lanka
Tel: 941521000, 941521006,
Fax: 941521932

212 Hettiarachchi,
Mangalika
dchris@sitnet.lk
zoomhr@kln.ac.lk
Department of Zoology
University of Kelaniya
Kelaniya, Sri Lanka

213 Maldeniya,
Rekha Rosanjani Perrera
National Aquatic Resources Agency
(NARA),
Crow Island, Mattakkuliya, Colombo 15

214 Nilakarawasam,
Nayanakanthi
Department of Zoology,
University of Ruhuna, Matara

215 Pathiratne, Asoka
asoka@kln.ac.lk
Department of Zoology,
University of Kelaniya, Kelaniya 11600

216 Vinobaba, P.
baba@eastu.esn.ac.lk
Department of Zoology,
Eastern University,
Vantharumoolai, Chenkaladi, Sri Lanka
Tel: 065 40528

TAIWAN

217 Chen, Hon-Cheng
honcheng@ccms.ntu.edu.tw
Institute of Fisheries Biology,
National Taiwan University,
Taipei, Taiwan
Tel: +886-2-23638554
+886-2-23630231 ext 3324
Fax: +886-2-23636837

218 Chen, Jau-Der
jdchen@ntou66.ntou.edu.tw
Department of Aquaculture,
National Taiwan Ocean University,
2, Pei-Ning Road,
Keelung 20224, Taiwan
Tel: +886-2-24622192 ext 5215
Fax: +886-2-24633150

219 Cheng, Winton
winton@mail.nupst.edu.tw
Department of Aquaculture,
National Pingtung University
1 Hseu Fu Road, Nei Pu, Hsiang,
Ping Tung, Taiwan
Tel: +886-8-7703202 ext 6224
Fax: +886-8-7740401

220 Chi, Shau-Chi
shauchi@ccms.ntu.edu.tw
Department of Zoology,
National Taiwan University,
No. 1 Sect. 4 Roosevelt Rd.,
Taipei 10764, Taiwan
Tel: +886-2-23630231 ext 3819
Fax: 886-2-2367-3852

221 Chien, Maw-Sheng
mschien@dragon.nchu.edu.tw
Department of Veterinary Medicine,
National Chung-Hsing University,
250 Kuo Kuang Road,
Taichung 40227, Taiwan
Tel: +886-4-22840894 ext 409
+886-4-22856419
Fax: +886-4-22862073

222 Chiou, Chwei-Jang
vetchiou@mail.baphiq.gov.tw
Bureau of Animal & Plant Health
Inspection Quarantine,
Council of Agriculture,
Executive Yuan, 3F, No. 51,
Sec. 2, Chung King S. Rd.,
Taipei, Taiwan
Tel: +886-2-23434247
Fax: +886-2-23922494

223 Chou, Hsin-Yiu
hychou@mail.ntou.edu.tw
Department of Aquaculture,
National Taiwan Ocean University,
2, Pei-Ning Road, Keelung 20224
Tel: +886-2-24622192 ext. 5214
Fax: +886-2-24634176

224 Hsu, Jung-Pin	jphsu@ms1.gsn.gov.tw Livestock Disease Diagnostic Lab., 110-1 Shui Yuan Sec., Pingtung 90011, Taiwan Tel: +886-8-7224427 Fax: +886-8-7224432	233 Song, Yen-Ling	song@ccms.ntu.edu.tw Department of Zoology College of Science National Taiwan University 1, Sec. 4, Roosevelt Road, Taipei 10764 Taiwan Tel: +886-2-23630231 ext. 3355 Fax: +886-2-23660243
225 Huang, Hsu-Tien	Livestock Disease Diagnostic Lab., 110-1 Shui Yuan Sec., Pingtung 900, Taiwan Tel: +886-8-7224427 Fax: +886-8-7224432	234 Sung, Hung-Hung	hhsung@mail.scu.edu.tw Department of Microbiology, Soochow University, Shih Lin, Taipei 11120, Taiwan Tel: +886-2-28827405 Fax: +886-2-28831193
226 K'O, Hao-Jan	Taiwan Prov. Research Institute for Animal Health, 376, Chung-Cheng Road, Tansui 251, Taiwan	235 Wang, Way-Shyan	wswang@dragon.nchu.edu.tw Department of Veterinary Medicine, National Chung-Hsing University, 250 Kuo Kuang Road, Taichung 40227, Taiwan Tel: +886-4-22854298 Fax: +886-4-22862073
227 Kou, Guang-Hsiung	ghkou@ccms.ntu.edu.tw Department of Zoology, National Taiwan University, No. 1 Sect. 4 Roosevelt Rd., Taipei 10764, Taiwan Tel: +886-2-23660506 Fax: +886-2-23638179	THAILAND	
228 Lee, Kuo-Kau	kklee@ntou66.ntou.edu.tw Department of Aquaculture, National Taiwan Ocean University, 2, Pei Ning Road, Keelung 20224, Taiwan Tel: +886-2-24622192 ext 5216 Fax: +886-2-24633150	236 Areechon, Nonthawith	ffisnwa@nontri.ku.ac.th Department of Fisheries Faculty of Fisheries Kasetsart University Bangkok 10900 Tel: 662-5792924/5797827 Fax: 662-5613984
229 Lin, Cheng-Chung	cclin1@mailnchu.edu.tw Department of Veterinary Medicine, National Chung-Hsing University, 250 Kuo Kuang Road, Taichung 40227, Taiwan Tel: +886-4-22840369 ext 30 +886-4-22862085 Fax: +886-4-22862085	237 Boonsaeng, Vichai	scvbs@mahidol.ac.th Department of Biochemistry Faculty of Science Mahidol University Rama 6 Road, Bangkok 10400 Thailand Tel: 662 201 5467 Fax: 662 248 0375
230 Lin, Ching-Long	cclin@mail.ncyu.edu.tw National Chiayi Institute of Technology, Department of Aquaculture, 84 Houng Mau Bai, Luh Liao Li, Chiayi, Taiwan Tel: +886-5-2783065 Fax: +886-5-2783065	238 Boonyaratpalin, Sitdhi	Department of Fisheries Kasetsart University Campus Jatujak, Ladyao, Bangkok 10900, Thailand
231 Liu, Michael R. S.	Department of Veterinary Medicine, National Taiwan University, 142 Chou-San Road, Taipei 10764, Taiwan Tel: +886-3-9333743	239 Boonyawiwat, Visanu	fvetvib@nontri.ku.ac.th Department of Veterinary Medicine, Faculty of Veterinary Medicine, Kasetsart University, Bangkok, Thailand
232 Lo, Chu-Fang	gracelow@ccms.ntu.edu.tw Department of Zoology, National Taiwan University, No. 1 Sect. 4 Roosevelt Rd., Taipei 10764, Taiwan Tel: +886-2-23630231 ext 3840 Fax: +886-2-23638179	240 Bunnajirakul, Sumrarn	sbunna@mut.ac.th sbunna@hotmail.com Faculty of Veterinary Medicine, Mahanakorn University of Technology, 51 Cheum-Sampan Rd., Nong Chok, Bangkok 10530 Tel: 66 2 988 3655 ext. 147, 148 Fax: 66 2 988 4040

241 Chanratchakool, Pornlerd	pornlerc@fisheries.go.th Aquatic Animal Health Research Institute (AAHRI) Department of Fisheries Kasetsart University Campus Jatujak, Ladyao, Bangkok 10900, Thailand Tel: 662-5796803 Fax: 662-5613993	248 Griffiths, Don	pondg@loxinfo.co.th DDS Aqua, 55/5, Village 4 Tambon Koh Yor Amphur Muang 90100 Thailand Tel: (66-74) 450065
242 Chansue, Nantarika	Veterinary Aquatic Animal Research Center, Chulalongkorn University, Bangkok 10330	249 Kanchanakhan, Somkiat	somkiatkc@fisheries.go.th Aquatic Animal Health Research Institute (AAHRI) Department of Fisheries Kasetsart University Campus Jatujak, Ladyao, Bangkok 10900, Thailand Tel: 662-5796803 Fax: 662-5613993
243 Chen, Ming-Dang	12th Flr. C.P. Tower, 313 Silom Road, Bangrak, Bangkok 10500	250 Kasornchandra, Jiraporn	kasornj@hadyai.loxinfo.co.th Marine Shrimp Research & Development Center Pawong Sub-district, Muang District Songkhla 90100 Thailand Tel: 667-4334516-8 ext. 111 Fax: 667-4334515
244 Chinabut, Supranee	supranee@fisheries.go.th Aquatic Animal Health Research Institute (AAHRI) Department of Fisheries Kasetsart University Campus Jatujak, Ladyao, Bangkok 10900, Thailand Tel: 662-5796803 Fax: 662-5613993	251 Laoprasert, Thitiporn	thitiporl@fisheries.go.th Aquatic Animal Health Research Institute (AAHRI) Department of Fisheries Kasetsart University Campus Jatujak, Ladyao, Bangkok 10900, Thailand Tel: 662-5796803 Fax: 662-5613993
245 Direkbusarakom, Sataporn	sataporn1@hotmail.com dsatapor@wu.ac.th Institute of Agricultural Technology, Walailak University, 222 Thaiburi, Tasala, Nakornsri Thammarat 80160 Thailand	252 Lawhavinit, Ong-ard	fvetonl@ku.ac.th Department of Veterinary Microbiology & Immunology Faculty of Veterinary Medicine Kasetsart University Bangkok, Thailand 10900 Tel: 662 9428436 Fax: 662-9428436
246 Fegan, Daniel	Dfegan@usa.net National Center for Genetic Engineering and Biotechnology (BIOTEC), Shrimp Biotechnology Business Unit, 4th Fl. Chalem Prakiat Bldg., Faculty of Science, Mahidol University, Rama 6 Road, Bangkok 10400, Thailand Tel: 662-201-5870-2 Fax: 662-247-7051	253 Lerssutthichawal, Theerawoot	1316/1-5 Buranaram Rd. Tawang, Muang Nakomsithammarat 80000
247 Flegel, Timothy William	sctwf@mahidol.ac.th Centex Shrimp, Chalem Prakiat Building Faculty of Science, Mahidol University Rama 6 Road, Bangkok 10400, Thailand Personal Tel: (66-2) 201-5876 Office Tel: (66-2) 201-5870 201-5871 or 201-5872 Fax: (66-2) 247-7051 Mobile: (66-1) 403-5833	254 Limsuwan, Chalor	aahri@yahoo.com Faculty of Fisheries, Kasetsart University, Bangkok 10900 Tel: 0-2579-5955 Fax: 0-2579-5955
		255 Nash, Gary L.	nashgar@hotmail.com Centex Shrimp, Chalem Prakiat Building Faculty of Science, Mahidol University Rama 6 Road, Bangkok 10400, Thailand Office Tel: (66-2) 201-5870-2 Fax: (66-2) 201-5873

256 Nitithamyong, Charoen	Department of Marine Science, Faculty of Science, Chulalongkorn University, Bangkok 10330	265 Soowannayan, Chumporn	National Center for Genetic Engineering and Biotechnology c/o Center of Excellence for Shrimp Molecular Biology and Biotechnology Faculty of Science, Mahidol University 4th Floor Chalermprakiat Bldg., 272 Rama 6 Road Rajdhevee, Bangkok 10400 Thailand
257 Pasharawipas, Tirasak	Department of Microbiology, Faculty of Medical Technology, Rangsit University, Paholyathin Road, Pathumthani 12000	266 Sritunyalucksana, Kallaya	Biotechnology Department Faculty of Science, Mahidol University, Rama VI Road, Bangkok 10400
258 Phillips, Michael John	michael.phillips@enaca.org Network of Aquaculture Centres in Asia-Pacific (NACA) Suraswadi Bldg., DOF Complex Kasetsart University Campus Ladyao, Jatujak, Bangkok 10900 THAILAND Tel: (662) 561-1728 to 9 ext. 115 Fax: (662) 561-1727	267 Supamattaya, Kidchakan	Department of Aquatic Science Faculty of Natural Resources, Prince of Songkhla University, Hat Yai, Songkhla 90110
259 Promwikorn, Waraporn	pwaraporn@ratree.psu.ac.th Department of Anatomy, Faculty of Science, Prince of Songkla University, Had-Yai, Songkhla, 90112 Thailand Tel: +66 (0) 74 446663 Fax: +66 (0) 74 446663	268 Tangtrongpiros, Jirasak	Jirasak.T@chula.ac.th Veterinary Aquatic Animal Research Center Faculty of Veterinary Medicine Chulalongkorn University Patumwan, Bangkok 10330 Thailand
260 Puttinaowarat, Suppalak	suppalap@fisheries.go.th Aquatic Animal Health Research Institute (AAHRI) Department of Fisheries Kasetsart University Campus Jatujak, Ladyao, Bangkok 10900, Thailand Tel: 662-5796803 Fax: 662-5613993	269 Taveekijakarn, Paveena	Faculty of Agricultural Technology, King Mongkut's Institute of Technology, Ladkrabang, Bangkok 10520
261 Ruangpan, Lila	National Institute of Coastal Aquaculture, Songkhla 90000	270 Tonguthai, Kamonporn	kamonpot@fisheries.go.th Aquatic Animal Health Research Institute (AAHRI) Department of Fisheries Kasetsart University Campus Jatujak, Ladyao, Bangkok 10900, Thailand Tel: 662-5796803 Fax: 662-5613993
262 Sirikanchana, Prapaisiri	College of Fisheries, Kasetsart University, Bangkok 10900	271 Tweetungtragoon, Attaporn	Attaporn2002@yahoo.com National Center for Genetic Engineering and Biotechnology c/o Center of Excellence for Shrimp Molecular Biology and Biotechnology Faculty of Science, Mahidol University 4th Floor Chalermprakiat Bldg., 272 Rama 6 Road Rajdhevee, Bangkok 10400 Thailand
263 Somsiri, Temdoug	temdouns@fisheries.go.th Aquatic Animal Health Research Institute (AAHRI) Department of Fisheries Kasetsart University Campus Jatujak, Ladyao, Bangkok 10900, Thailand Tel: 66 0-2579-4122 Fax: 66 0-2561-3993	272 Wisessang, Suchana (retired)	Marine Science Department, Faculty of Science, Chulalongkorn University, Bangkok 10500
264 Soontornkit, Sommas	Charoean Pokphand Feedmill Co. Ltd. 36 Soi Yencht Chand Road, Yannawa, Bangkok 10120	273 Wongtavatchai, Janenuj	Janenuj.W@chula.ac.th Department of Medicine Faculty of Veterinary Medicine Chulalongkorn University Patumwan, Bangkok 10330 Thailand

UNITED KINGDOM

274 Adams, Alexandra	Alexandra.adams@stir.ac.uk Institute of Aquaculture Stirling University Stirling FK 9 4LA Scotland, United Kingdom
275 Alderman, David	d.j.alderman@cefes.co.uk CEFAS Weymouth Laboratory, Weymouth, Dorset, DT4 8UB, United Kingdom + (44) 1305 206641 + (44) 1305 206638
276 Crumlish, Margaret	margaret.crumlish@stir.ac.uk mc3@stir.ac.uk Institute of Aquaculture Stirling University Stirling FK 9 4LA Scotland, United Kingdom Tel: + 44 01786 467911 Fax: + 44 01786 472133
277 Hamley, Ernest Basil	Hamley Consultancies International, 59 Eylewood Road, London SE27 9LZ
278 Maclean, Marlie	Marlie.maclean@headoffice.mrc.ac.uk
279 Prayitno, Slamet Budi	Department of Marine Biology, University of North Wales, Menai Bridge, Gwynedd LL59 5Eyb
280 Roberts, Ronald John	HeronPisces@compuserve.com
281 Macrae, Ian Hamilton	ianmacrae@hotmail.com imacrae@napier.ac.uk
282 Millar, Stuart	s.d.millar@stir.ac.uk Institute of Aquaculture Stirling University Stirling FK 9 4LA Scotland, United Kingdom Tel: +44 (0)1786 467992 Fax: +44 (0)1786 472133
283 Thompson, Kim	k.d.thompson@stir.ac.uk kdt1@stir.ac.uk Institute of Aquaculture Stirling University Stirling FK 9 4LA Scotland, United Kingdom Tel: +44 (0)1786 467912; Fax: +44 (0)1786 472133
284 Turnbull, James	j.f.turnbull@stir.ac.uk Institute of Aquaculture Stirling University Stirling FK 9 4LA Scotland, United Kingdom Tel: +1786 467913 Fax : +1786472133

UNITED STATES OF AMERICA

285 Athappilly, Jim Chacko	Aquaculture Department, Unity College, Unity, Maine 04988
286 Baxa, Dolores	dvbaxa@ucdavis.edu School of Veterinary Medicine Department of Medicine & Epidemiology University of California Davis CA 95616 USA Tel: 530-752-9318 Fax: 530-752-0414
287 Browdy, Craig Lawrence	browdycl@musc.edu Marine Resources Research Institute South Carolina Department of Natural Resources 217 Ft. Johnson Road PO Box 12559 Charleston, SC 29422 USA Tel: 843-953-9840 Fax: 425-944-2449
288 Cardella, Matteo	National Veterinary Services Lab., APHIS, US Dept.Agriculture, AMES, Iowa 50010
289 Dunlap, Paul	pvdunlap@umich.edu University of Michigan Department of Ecology & Evolutionary Biology Kraus Natural Science Building 830 North University Avenue University of Michigan Ann Arbor, MI 48109-1048 Tel: 734 615-9099 (office) 734-764-3580 Tel: 734 615-9804 (lab) Fax: 734 763-0544
290 Furtado, Jose I. dos R.	Economic Development Institute [EDI/ARI], The World Bank, 1818 H Street, NW, Washington DC 20433
291 Hetrick, Frank M.	Maryland Department of Agriculture, University of Maryland, College Park, MD 20742, USA
292 Lee, Kendrick K. F.	Lee Resources, 1071 Kalikimaka St., Honolulu, HI 96817-1224
293 Moore, James D.	jimmoores@ucdavis.edu Marine Region, California Department of Fish and Game Bodega Marine Laboratory 2099 Westside Road, Bodega Bay CA 94923 USA Tel: 07 875-2067 Fax: 707 875-2089

294 Overstreet, Robin Robin.Overstreet@usm.edu
 Gulf Coast Research Laboratory
 The University of Southern Mississippi
 P.O. Box 7000 (Courier: 703 East
 Beach Drive, 39564)
 Ocean Springs, MS 39566-7000
 Tel: 228-872-4243
 Fax: 228-872-4204
<http://www.coms.esm.edu>

295 Quines, Oscar D. 9538 van Ruiten Street,
 Bellflower, CA 90706

296 Reantaso, Melba B. Melbar99@yahoo.com
 mreantaso@dnr.state.md.us
 Cooperative Oxford Laboratory
 Maryland Department
 of Natural Resources
 904 S. Morris Street
 Oxford, Maryland 21654
 Tel: 410-226-5193
 Fax: 410 226-5925

297 Shelton, William L. wshelton@ou.edu
 Department of Zoology
 University of Oklahoma,
 Norman, Oklahoma 73019
 Tel: 405-325-1058

298 Vinitnantharat, Somsak somsakv@msn.com
 20619 Filbert Drive
 Bothell, WA, 98012
 Tel/Fax: +1 425 489 2045

VIETNAM

300 Phan Thi, Van phanvan@hn.vnn.vn
 Research Institute for
 Aquaculture No. 1
 Dinh Bang, Tien Son
 Bac Ninh, Vietnam
 Fax: 84-48273070

INSTITUTIONAL MEMBERS

301 San Miguel Corporation (Aquaculture Operations)
 San Miguel Foods Inc.,
 2nd Floor, PDCP Building,
 Nichols Interchange,
 South Super Highway, Makati City
 Philippines

302 CSIRO Australian Fish Health Laboratory (Geelong)
 CSIRO Australian Animal Health Lab.,
 P.O. Bag 24, Geelong, Victoria 3220
 Australia

303 Fisheries & Aquaculture
 International, Ltd. No. 7, Kohji-machi Bldg.,
 Room B105, 4-5 Kohji-machi,
 Chiyoda-ku, Tokyo 102-0083 Japan

304 Network of Aquaculture
 Centres in Asia-Pacific (NACA) naca@enaca.org
 pedro.bueno@enaca.org
 simon.wilkinson@enaca.org
 Surawadi Bldg., DOF Complex
 Kasetsart University Campus
 Ladyao, Jatujak,
 Bangkok 10900 THAILAND
 Tel: (662) 561-1728 to 9
 Fax: (662) 561-1727
<http://www.enaca.org>

305 Ocean Star International (Simon Goe)
 P. O. Box 643, Snowville,
 UT 84336 USA

FHS/AFS Secretariat Offices

FHS Secretariat C/o Celia Lavilla-Pitogo
 celiap@aqd.seafdec.org.ph
 Fish Health Section
 SEAFDEC Aquaculture Department
 Tigbauan 5021, Iloilo, Philippines
 Tel: (63-33) 3362965
 Fax: (63-33) 3351008
 Website: <http://afs-fhs.seafdec.org.ph>

AFS Secretariat C/o Elsie Tech
 Executive Officer
 Asian Fisheries Society
 25-A Mayaman Street
 UP Village, Quezon City
 Tel: 921-1914
 Fax: 920-2757
 Email: afs@compass.com.ph
<http://www.compass.com.ph/~afs/>
<http://www.cgjar.org/iclarm/afs/>



S Y M P O S I U M A B S T R A C T S

5th Symposium On Diseases in Asian Aquaculture

Session 1 - Biosecurity and Risk Assessment

Biosecurity: A new word for an old concept

Peter Beers^{1}, Vanessa Findlay¹ and Ramesh Perera¹*

1. Aquatic Animal Biosecurity, Biosecurity Australia, PO Box 858, Canberra

Biosecurity is a fashionable word that is being used in a number of different circumstances. Its meaning in these situations is explored. The concept of biosecurity is used to cover the management of risks arising from biological organisms and agents that may cause harm to living organisms and other aspects of the environment. Following the spread of diseases such as whitespot and Taura syndrome in prawns, Akoya disease in pearl oysters and epizootic ulcerative syndrome in fish, the need for improvement to biosecurity for aquatic animals has been recognised. The principles underpinning the development of a biosecurity program are identified. Biosecurity programs should have a strong scientific basis and use risk assessment to evaluate risk management approaches to ensure that measures provide appropriate protection without unduly hindering business opportunities.

NOTES

Viral DNA sequences of genes encoding the ATPase and the major capsid protein of tropical iridovirus isolates which are pathogenic to fishes in Japan, South China Sea and Southeast Asian countries

Miyazaki Teruo ^{1*}, Sudthongkong Chaiwud ¹, Miyata Masato ¹

1. Mie University

Tropical iridovirus infection causes severe epizootic resulting in mass mortalities and large economic losses in freshwater ornamental fishes cultured in Southeast Asian countries, in wild fish seedlings captured in South China Sea, and in marine fishes farmed in Japan, Singapore, and Thailand. All of tropical iridovirus-infected fishes histopathologically showed the systemic formation of inclusion body-bearing cells and necrosis of virus-infected splenocytes and hematopoietic cells. We designed primer sets for the ATPase gene and the major capsid protein (MCP) gene and sequenced the PCR products derived from 5 iridovirus isolates from sea bass in South China Sea (SBIV), red sea bream in Japan, brown-spotted grouper with a grouper sleepy disease in Thailand, dwarf gourami from Malaysia and African lampeye from Sumatra Island, Indonesia. The ATPase gene and the MCP gene of these 5 viral isolates were highly homologous (>95.8%, >94.9% identity, respectively) and the deduced amino acid sequences of the ATPase and the MCP were also highly identical (>98.1%, >97.2% identity, respectively). Based on the high homology, these 5 isolates of tropical iridovirus from various fishes in geographically different regions were determined to have a single origin and to be native to Southeast Asian regions. However, these sequences were far different from those of members of the genera Ranavirus (FV3, ECV, ESV), Lymphocystivirus (LCDV-1) and Iridovirus (CIV) in the Family Iridoviridae. The SBIV MCP amino acid sequence had 47% identity and 67% similarity to that of LCDV-1, 45% identity and 63% similarity to FV3, 45% identity and 68% similarity to ECV, 45% identity and 63% similarity to ESV, and 45% identity and 63% similarity to CIV. We propose a new genus 'Tropivirus' for tropical iridovirus in the Family Iridoviridae.

NOTES

**Session 3 - Diseases of aquatic vertebrates****Ranavirus of fish, amphibians and reptiles***Hyatt Alex D* ^{1*}

1. CSIRO Livestock Industries

Ranaviruses are a group of viruses belonging to the genus *Ranavirus* within the family *Iridoviridae*. The genus contains viruses known to infect and cause disease in a range of ectothermic animals namely fish, amphibians and reptiles. The piscine viruses belong to three, possibly four sub-groups within this genus whilst those isolated from amphibians and reptiles belong to the same group. Another group of associated iridoviruses, namely the erythrocytic iridoviruses, are yet to be classified by the ICTV and are known to cause anemia in three classes of animals. Each group of viruses has their own characteristics and can be identified via an array of diagnostic tests. The various groups will be discussed within the presentation together with accepted (OIE) testing procedures. So what is the significance of ranaviruses? Ranaviruses have been described as the 'cold-blooded killers'. Viruses have been associated with fish kills in Asia, Australia, North America and Europe. Within North America, endangered tiger salamanders have undergone major population fluctuations and in the United Kingdom the common frog has undergone annual population crashes because of infection with ranaviruses. These viruses have also been isolated from healthy and diseased reptiles. The potential for ranaviruses to cross animal classes and species make them potentially lethal to the finfish industry and to piscine and herpetofauna biodiversity. In this presentation these viruses will also be discussed in terms of 'ecosystem health' i.e. do ranaviruses represent emerging viruses, are they a threat to aquaculture and the herpetofauna, and are they the cause of declines in amphibian populations?

NOTES

Emerging diseases of soft-shell turtles

Yulin Jiang ^{1*}

1. Shenzhen Exit-entry Inspection and Quarantine Bureau

The soft-shelled turtle has been widely cultured in whole China in the recent 10 years because of its nutritional and medicinal values. Unfortunately, epizootic outbreaks of disease usually occurred in densely stocked young turtle ponds. Some symptoms, such as shell perforation (furuncle), red neck, red dots, ulcerated skin, blisters, white spots and white dots has been described and called as names of diseases by farmers. The major problem is that the pathogen causing the clinical signs has not been identified clearly. Frequently, same bacteria were isolated from sick turtle with different symptoms, or different kinds of bacteria were isolated from sick turtle with same symptoms. Besides, there exists secondary infection and mixed infection, It could cover up primary infection. This review describes in detail some diseases of soft shelled turtle in China according to pathogens, such as Trionyx sinensis virus (TSV), Soft shelled turtle iridovirus(STIV), *Aeromonas hydrophila*, Edwardsielliasis. Aspects of their etiology, epidemiology, immunology and control are summarized.

NOTES



New and emerging diseases in reef and Estuarine fishes from North Queensland, Australia

Bowater Rachel ^{1*}, *Cox Elizabeth* ², *Thomas Annette* ¹, *Humphrey John* ³, *Deveney Marty* ⁴

- 1. Queensland Department of Primary Industries
- 2. Queensland Department of Primary Industries. Northern Fisheries Centre, Cairns
- 3. Department of Business, Industry and Resource Development
- 4. Department of Primary Industries & Resources South Australia

The Department of Primary Industries Northern Fisheries Centre (NFC), Cairns, Queensland is undertaking research into reproduction and larval rearing technologies for a range of finfish including grouper species (barramundi cod, *Cromileptes altivelis*, flowery cod *Epinephalus fuscoguttatus* and estuary cod *Epinephalus cuoides*) and estuarine species (mangrove jack, *Lutjanus argentimaculatus* and barramundi, *Lates calcarifer*). Potential broodstock have been captured from wild stocks, as sources of captive-bred stocks are unavailable in Australia. This paper presents a synopsis of newly emerging diseases seen in these species. Disease investigations included a range of diagnostic aids and tests done at Oonoonba Veterinary Laboratory, Townsville, including gross pathology, histopathology, electron microscopy, bacterial and fungal culture, isolation and identification and viral isolation in cell culture. The clinical features and diagnostic findings for each disease are presented here. Emerging diseases include a new syndrome of hemangioma-like neoplasia of the gills with unknown aetiology observed in *C. altivelis*, a fungal infection in the swim bladder of *C. altivelis* with *Cladosporium cladosporioides* and *Scopulariopsis brumptii*, and a gill fluke infestation of *E. fuscoguttatus* with *Allobenedenia* n. sp. Wild-harvested, captive barramundi cod *C. altivelis* and coral trout *Plectropomus leopardus* have also shown infection with larval *Terranova* sp. Type II associated with visceral fibrosis, peritonitis and death. Other diseases encountered infrequently, are briefly mentioned and include White spot (*Cryptocaryon irritans*), Amyloodiniasis (*Amyloodinium ocellatum*), Vibriosis (*Vibrio harveyi*, *Vibrio alginolyticus*) and gill and skin fluke infestation.

NOTES

A virological survey in diseased grouper in Thailand using virus isolation and polymerase chain reaction (PCR) technique

Kanchanakhan Somkiat^{1}, Roongkamnertwongsa Somporn², Dolnayadol Yaowanit²*

1. Aquatic Animal Health Research Institute, Department Of Fisheries

2. National Institute Of Coastal Aquaculture, Department Of Fisheries

A virological survey had been conducted in brown spotted grouper, *Epinephelus malabaricus*, cultured in cages, earthen ponds and pens during April 2001 to January 2002 in East Coast and South Coast of Thailand. The adult grouper samples exhibited a wide range of clinical signs beginning from darkening body color with focal distension of the skin to red boil skin and to red spot ulcers on the body and head, while diseased fry to juvenile grouper showed darkening body color and swirling movement. Thirty-five isolates of viruses were discovered from using freshwater fish cell lines, SSN-1 and EPC. These isolates could be grouped into two different groups of fish viruses. One viral group was identified as a betanodavirus, which belonged to genotype RGNNV. As RT-PCR tests using specific primers to SJNNV revealed negative, while specific primers to RGNNV showed positive RT-PCR product. The second viral group could be identified as an Iridovirus. This grouper iridovirus had some level of similarities to red sea bream iridovirus. As one set of specific primers for RSIV could give positive PCR product while the second set of specific primers gave negative PCR result. The grouper iridovirus was found different to Ranavirus. Betanodavirus could infect both adult and fry to juvenile stage of the grouper and seemed to have more significant during November to January or during the dry and cold season in Thailand. Grouper iridovirus seems to cause disease in adult size of the fish with low level of mortality and seems to infect the fish predominantly in August and September or during rainy season in Thailand. Findings indicate that two different viruses are associated with diseased grouper cultured in Thailand that are identified as Betanodavirus and grouper iridovirus. There is a need to do more molecular studies of these new viral isolates as well as their pathogenesis.

NOTES



SYMPOSIUM ABSTRACTS

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Isolation and identification of *Edwardsiella ictaluri* from diseased *Pangasius hypophthalmus* (Sauvage) cultured in Vietnam

Crumlish Margaret ^{1*}, *Dung Tu Thanh* ², *Turnbull James* ¹, *Ngoc NTN* ², *Ferguson Hugh* ¹

1. Institute of Aquaculture, Stirling University

2. Aquaculture and Fisheries Science Institute, CanTho University

Farming of the indigenous freshwater catfish *P.hypophthalmus* in Vietnam has contributed to the livelihoods of rural families for decades. Many features of this fish make it an attractive option for culture. It has a relatively short production cycle of 8 months from stocking to harvest, there is a ready domestic and international market for the product and captive reproduction ensures that fish seed is available all year. Many families raise this omnivorous species extensively in polyculture ponds for family consumption or to sell locally on the domestic markets. Since 1995 there has been an increase in the number of families producing *Pangasius* spp. intensively in river-based cages, from where 80% of the total annual production is exported as frozen processed products. Fish farmers in Vietnam recently reported lower yields and high level mortalities in their systems due to disease outbreaks. These outbreaks occurred in fingerling and grow-out fish where diseased animals had few if any external signs of disease but in which white spots were observed on the internal organs. The disease was called bacillary necrosis of *Pangasius* (BNP) due to the presence of microcolonies of large rod-shaped bacteria within necrotic areas of tissue, as observed by light microscopy in tissue sections. Pure bacterial growth was recovered from *P. hypophthalmus* with clinical signs of BNP from various provinces of the Mekong Delta. The fastidious colonies required 48 hours at 28°C before punctate, small, light coloured colonies were visible on solid agar. These colonies were identified as *E. ictaluri* by primary and biochemical tests, by immunohistochemistry and 16S rRNA analysis. The optimal growth temperature on solid media was 28°C and colonies grew on agar selective for *E. ictaluri*. This presentation describes the findings in naturally occurring infections in farmed *P.hypophthalmus* due to *E.ictaluri*. To the authors' knowledge this is the first time that *E. ictaluri* has been identified from farmed *Pangasius* spp.

NOTES

Session 4 - Epidemiology

International trade and risk analysis

Baldock Chris ^{1*}

1. AusVet Animal Health Services

The importation of animals and animal products involves a degree of disease risk to the importing country. This has been highlighted by the pandemic of white spot disease of shrimp and very recently by the outbreak of disease in koi carp and common carp in Indonesia suspected to have resulted from imports of live animals. Because of the serious impacts of infectious diseases, particularly in farmed aquatic animals the process of import risk analyses (IRA) to prevent the entry and spread of unwanted pathogens is assuming increasing importance. The principal aim of IRA is to provide importing countries with an objective and defensible method to assess the disease risks associated with the importation of animals, animal products, animal genetic material, feedstuffs, biological products and pathological material. It forces a thorough and logical approach to be adopted in considering the likelihood of undesirable events, and identifies gaps in our current knowledge. In undertaking an import risk analysis, a country must be guided by the International Aquatic Animal Health Code (Code) of the Office International des Epizooties (OIE). The OIE Code provides guidelines for national authorities to assist them in addressing the principles laid out in the WTO's Agreement on the Application of Sanitary and Phytosanitary Measures. Facilitating trade while at the same time managing the associated disease risks is a challenge for all those associated with aquatic animal health. The coming years are likely to see an increasing need for skills and experience in this very important area.

NOTES

Farm level risk factors for white spot disease outbreaks

Corsin Flavio^{1*}

1. University of Liverpool

White spot disease (WSD) is a pandemic disease of crustaceans caused by White Spot Syndrome Virus (WSSV). The scientific community has acted promptly in an attempt to limit its devastating impact on the shrimp farming industry. In the earlier stages of the epidemic, a number of potential risk factors for WSD outbreaks were hypothesised from information on other diseases and circumstantial evidence; as a result recommendations were distributed to farmers. In addition, highly sensitive diagnostic methods were developed that allowed the detection of WSSV in shrimp broodstock, post-larvae and wild animals. This led to further interventions aimed at reducing the risk of introducing the virus into populations. The effect of some of the hypothetical risk factors for WSD was tested through experimental trials. These, however, used WSD at the animal level as an outcome. Field investigations were also carried out and provided further information on risk factors for outbreaks at the pond level and the information generated has been used to investigate WSSV transmission using mathematical modelling. In this presentation, the farm level risk factors for WSD are discussed paying particular attention to the effects of the presence or introduction of WSSV into the farming system; shrimp stocking density and other management practices that might be associated with WSD outbreaks. The information presented is derived from manuscripts published in peer-reviewed journals, reports and the experience gathered by the authors during longitudinal epidemiological studies of WSD carried out in Vietnam and India. In addition to the information from field studies, results from experimental trials and mathematical modelling are also examined for their potential value at the farm level.

NOTES

Application of epidemiology to support better health management in black tiger shrimp *Penaeus monodon* aquaculture: an experience from India

Padiyar Arun^{1*}, *Primphon Mongkhon*², *Chanratchakool Pornlerd*³, *Bhat Vishnu*⁴, *Cameron Angus*⁵, *Phillips Michael*¹

1. Network of Aquaculture Centres in Asia-Pacific, Bangkok, Thailand
2. Siam Natural Resources Ltd., Bangkok, Thailand
3. Aquatic Animal Health Research Institute, Bangkok, Thailand
4. The Marine Products Export Development Authority, Ministry of Commerce & Industry, Govt. of India

In 2001, a longitudinal epidemiological study of 365 randomly selected shrimp ponds was carried out in two districts of Andhra Pradesh, India to identify risk factors for shrimp disease outbreaks (emphasising the economically devastating white spot disease) and low pond production. Risk factors significantly associated with disease outbreaks were then used to identify locally relevant risk management practices. During 2002, demonstration sites were established in the two districts to support and evaluate the practical implementation of these better management practices (BMPs) on private farms. In this paper, experiences from the demonstrations in two villages in West Godavari district (8 ponds on 4 farms) are presented. Risk and protective factors identified from 184 epidemiological study ponds in this district were used to design and implement BMPs relevant to the local modified extensive farming system. Although farmers experienced disease outbreaks, demonstration ponds had better performance, in terms of days of culture (DOC), mean body weight (MBW), production compared to 2001 in the same ponds and with nearby non-demonstration ponds during 2002. The average DOC, MBW, production was 93, 18.5 grams and 417 kg/ha respectively. The demonstrations provided further understanding of risk factors, BMPs, shrimp disease occurrence and farm performance. Timely implementation of BMPs by farmers was a major factor in crop performance, which was dependent on farmers’ understanding and willingness to adapt BMPs and their financial status. Experiences from this study show the epidemiological approach provided understanding of shrimp disease risks, but constraints to implementation of BMPs at farm level need to be understood and addressed in research and extension if such findings are to contribute to adoption of better health management practices at farm level.

NOTES

Patterns of occurrence of the helminth parasites of the bullseye pufferfish (*Sphoeroides annulatus*) from Sinaloa, Mexico

Roque Ana ^{1*}, Aguilar G ¹, Duncan N ¹, Fajer-Avila E ¹

1. CIAD/Mazatlan Unit for Aquaculture and Environment Management

The Mexican shrimp culture industry is starting to diversify into culturing other species. The bullseye puffer (*Sphoeroides annulatus*) has good aquaculture potential and is under pilot culture. However, parasitic infestations are a common problem for the bullseye puffer. The aim of this study was to determine patterns of occurrence of the helminth parasites from wild pufferfish populations used as potential broodstock. A total of 226 specimens were collected from two locations (Teacapan and Mazatlan) in Sinaloa state and were examined for the presence of helminth parasites. Two monogenea (*Neobenedenia melleni* and *Heterobothrium ecuadori*) and four trematodes (*Bianium plicatum*, *Lintonium vibex*, *Homalometron longisinsum* and *Phyllodistomum mirandai*) were identified. The most prevalent species were *H. ecuadori* (44%), *B. plicatum* (28%) and *N. melleni* (19%). Generally fish from Mazatlan had 2 helminthic species and fish from Teacapan 5 species. A relative risk analysis showed fish from Teacapan were at higher risk of presenting parasites (RR=1.85) than fish from Mazatlan. *B. plicatum* had higher probabilities (RR= 1.5) of being present in temperatures of 21-25 C than 26-30 C. *H. ecuadori* had higher probabilities (RR=1.72) of being present in fish from Teacapan than Mazatlan, in temperatures of 23-24.5 C (prevalence=67%, mean abundance =3 parasites/fish); and when other parasites were also present on the gill (RR=1.34). *N. melleni* had higher probabilities of being present when other parasites were present, such as *Lepeophtheirus simplex* (RR=1.31), trichodinids (RR=1.47), *Udonella* sp.(RR=1.34). This study is one of the first contributions to the knowledge of the parasitic biology of this fish species.

NOTES



Epizootic haematopoietic necrosis virus: Epidemiology and uncertainty

Whittington Richard ^{1}, Dixon Robert ¹, Li Lun ¹, Marsh Ian ², Hyatt Alex ³*

- 1. The University of Sydney
- 2. NSW Agriculture
- 3. CSIRO Australian Animal Health Laboratory

EHNV is one of only a handful of finfish viral agents requiring notification to OIE. Although closely related to other ranaviruses found in north America and Europe, it is distinguishable from these agents at the molecular level and is biogeographically isolated within south east Australia. Responsibility for control and field research lies with Australia. Two species are known to become infected naturally, free-living redfin perch *Perca fluviatilis* and farmed rainbow trout *Oncorhynchus mykiss*. Epidemiological evidence will be presented to support a view that neither of these is the natural host; the origin of the virus and its reservoir in Australia remains a mystery. However, slow continued expansion of the geographic range, and experimental evidence of a broader range of susceptible hosts is cause for concern related to future trade in finfish and effects on the aquatic ecosystem.

NOTES

Session 5 - Shrimp Health 1

Key farm management issues to reduce loss from diseases

Chanratchakool Pornlerd ^{1*}

1. Aquatic Animal Health Research Institute

Among the diseases in cultured shrimp, bacterial and viral infection has proved to be the most serious cause of loss. Many attempts have been tried in order to reduce the loss. Currently closed system with liner pond and more intensive farming has been tried in Latin America whereas less intensive, low input system has been used in Asian country. Screening of post larvae for low prevalence of various viral infections is commonly used with various results. The post larvae sampling techniques still requires further study in order to ensure more accurate result. Formalin treatment for post larvae prior to stock does seem to be effective in eliminating infected/weak animals from the population and therefore minimizing the risk of a serious outbreak later. However, basic farm management including pond preparation, water and pond soil management as well as the stocking schedule is seemed to play an important role in trigger an outbreak. In order to reduce the loss from serious disease outbreak, some steps on basic pond management with emphasized on the water quality and pond soil monitoring were discussed, in particular on oxygen requirement in the system, relationships between pH, ammonia and hydrogensulfide.

NOTES

Experimental transmission of Hepatopancreatic parvovirus (HPV) infection in *Penaeus monodon* postlarvae

Catap Elena ^{1*}, *Travina Remia* ¹

1. Southeast Asian Fisheries Development Centre

Hepatopancreatic parvovirus (HPV) infection in penaeid shrimps was first reported in various countries of the Asia-Pacific region in mid-1980's. The virus affects the hepatopancreas of postlarvae and juveniles, usually leading to slow growth and mortality during the early stage of culture. At present, there is no established experimental model of infection since there has not been any report of successful HPV transmission under laboratory condition in any known susceptible penaeid shrimp. Therefore, experiments were undertaken to induce HPV infection by feeding *Penaeus monodon* postlarvae (PL's) with virus-infected PL's. *P. monodon* PL's (PL-15), initially examined to be free from HPV, were found HPV-positive 24 hours after they were fed with the infected material. Percentage of infection was from 30% (day 1) to 100% (day 7) based on the examination of wet mounts of hepatopancreas (squashed tissue) stained with malachite green and through histopathology. This is the first report of a successful horizontal transmission of HPV in *P. monodon* PL's. This infection model could be used to study the pathogen further and would permit controlled experiments to be undertaken in order to identify prevention and control against the pathogen.

NOTES



Does spawning stress trigger the replication of white spot syndrome virus in shrimp?

Kou Guang-Hsiung^{1}, Hsia Hui-Lan¹, Chen Li-Li¹, Lo Chu-Fang¹*

1. Department of Zoology, National Taiwan University

In shrimp, a persistent viral infection can be converted to productive viral infection by stressors such as poor water quality and overcrowding. Here we investigate whether the stress associated with spawning can also trigger virus replication in shrimp. Quantitative real-time PCR was used for a temporal analysis of WSSV loads in a batch of 14 wild-caught black tiger shrimp (*Penaeus monodon*) brooders. For these specimens, four basic patterns emerge: Group 1 was comprised of 4 specimens which had a relatively high initial virus load (approximately 800 ~ 8000 viral DNA copies/per µg total DNA) and in these specimens, the virus replicated rapidly up to the time of spawning. After spawning the virus levels remained high and all of these shrimp died within a few hours. Two other groups (II and III) both had similar initial virus loads (approximately 10~ 600 viral DNA copies/per µg total DNA), but in group II (5 specimens), as in group I, the virus replicated rapidly up to and beyond spawning and all the shrimp died soon after spawning, whereas in Group III (3 specimens), the virus load increased only relatively slightly (approximately a 10-fold increase) and the shrimp survived well beyond spawning. The Group IV (2 specimens) shrimps did not spawn at all during the observation period, and no viral replication was triggered. At this time, it seems clear that spawning stress can trigger WSSV replication, but it is not known why or how this does not occur in some (25%) specimens.

NOTES



Predicting outbreaks of White Spot Disease in a semi-intensive *Penaeus monodon* culture system in Karnataka, India

Turnball JF^{1*}, *Corsin F*², *Mohan CV*³, *Padiyar PA*³, *Thakur PC*³, *Madhusudan M*³, *Hao NV*⁴, *Morgan KL*²

- 1. Institute Of Aquaculture, University Of Stirling
- 2. Department Of Veterinary Clinical Science And Animal Husbandry, The University Of Liverpool
- 3. Fish Pathology Laboratory, Department Of Aquaculture, College Of Fisheries
- 4. Research Institute For Aquaculture

A longitudinal study of 70 semi-intensive shrimp farms was undertaken in Karnataka, southwest India. White Spot Disease (WSD) was defined as the observation of mortality by farmers and detection of White Spot Syndrome Virus (WSSV) by 1-step PCR in harvested shrimp. Samples were collected from 62 ponds at harvest and of these 37 (59%) fulfilled the case definition. In this system WSD had a significant effect on both the length of production cycle and productivity. Farmers have tried to reduce losses from WSD through avoiding risks and harvesting in the face of an outbreak. However, the information available on which to base such strategies can be misleading. Typical white spots are not highly predictive of a WSD outbreak and in this study neither was the presence of the virus (nested 2 step PCR). Stocking PCR positive post-larvae was not significantly associated with the length of the production cycle, yield, the presence of WSSV at harvest or the risk of WSD. Nor was the presence of WSSV in shrimp from the pond (1- or 2-step PCR) associated with length of production cycle, yield or the risk of WSD. The findings indicate that WSSV and shrimp with WSD histopathology can be present in the pond without progressing to a full outbreak of WSD. However, there were significant associations between WSD and both observation of typical histopathology in dead shrimp from the pond and the pattern of mortalities. The implications of these findings for informing decisions on harvest strategy in various farming systems will be discussed.

NOTES

Session 6 - Shrimp Health 2

White spot syndrome: What we have learned about the virus and the disease

Lo Chu-Fang^{1*}, *Kou Guang-Hsiung*¹

1. Department of Zoology, National Taiwan University

One of the lessons ultimately learned from the first dramatic outbreaks of white spot syndrome virus (and other shrimp viral diseases) in the early 1990's was that aquaculture management practices needed to be improved. Subsequent research revealed that critical factors included: water quality and sourcing, culture density, broodstock sourcing, postlarva sourcing, screening techniques and strategies, diverse transmission pathways, critical infection levels, disease susceptibility, and stressors. This paper reviews the work done in the last decade by several research groups, and shows how studies on the key aspects of the biology of WSSV infection have led to improved disease management solutions that are now widely used by the shrimp aquaculture industry.

NOTES



SYMPOSIUM ABSTRACTS

5th Symposium On Diseases in Asian Aquaculture

Molecular genetics of white spot syndrome virus

Viak Just M^{1*}, Marks Hendrik¹, Ren Xinjing^{1 & 2}, Wittevelde Jeroen¹, Sandbrink Hans², Van Hulten Marielle C W¹

1. Wageningen University

2. Plant Research International

White spot syndrome virus (WSSV) is a major disease agent of penaeid shrimp and other crustaceans. The virus particle has an ovoid-to-bacilliform shape with a tail-like appendage. Its rod-shaped striated nucleocapsid contains a large circular double-stranded DNA molecule with a size of about 300 kbp and is wrapped in an envelope. With this DNA size WSSV is among the largest animal viruses. The virus replicates and assembles in the nucleus of infected cells. Its DNA is further characterized by the presence of multiple regions with homologous repeats dispersed around the genome. Isolates differ primarily in a single region where extensive insertions / deletions have occurred. The viral DNA encodes about 180 open reading frames, most of which have not been described elsewhere in data bases. The largest ORF encodes a putative protein of over 660 kDa. Five major and many minor structural virion proteins have been analyzed and their genes identified. Genes involved in nucleotide metabolism, DNA replication and protein modification have also been found. The WSSV sequence was further analyzed in silico using the computer program MEME and a set of computer scripts for the presence of conserved promoter motifs. This analysis was complemented by 5' RACE analyses of a multitude of ORFs. Transcription initiation sites within the nucleotide sequence TCAYTC for early and TMMTRACM for late transcripts were identified and suggest a unique transcriptional regulation machinery. On the basis of its unique morphological and genetic features WSSV has been accommodated in a new virus family (Nimaviridae).

NOTES

Breeding shrimp for disease resistance: A panacea or pariah

Moss Shaun M ^{1}, Doyle Roger W ², Lightner Donald V ³*

- 1. The Oceanic Institute
- 2. Genetic Computation Ltd.
- 3. University of Arizona, Department of Veterinary Sciences

After the 1992 Taura Syndrome Virus (TSV) epizootic in Ecuador, shrimp farmers in the Western Hemisphere began implementing a variety of strategies to mitigate crop loss from this pathogen, including selective breeding programs to develop TSV-resistant shrimp using between-family selection. Although TSV-resistant shrimp initially improved production and profitability for those farmers who were experiencing a TSV outbreak, genetically improved stocks were not a panacea to the broader disease problems plaguing the industry. In fact, breeding shrimp for resistance to a single viral pathogen, using current selective breeding strategies, may not be the most prudent course of action for the long-term viability of the shrimp farming industry. There are a number of concerns associated with current breeding strategies. Similar to other organisms, there appears to be a trade-off between disease resistance and shrimp growth. Disease-challenge tests typically used to estimate shrimp breeding values are inefficient, and there are concerns that results from laboratory challenge tests may not be predictive of survival in commercial ponds. There are growing concerns about viral mutations, whereby previously resistant shrimp strains may become susceptible to evolving viruses. Effective strategies for selecting disease resistant shrimp do exist, as do biosecurity protocols which exclude specifically listed pathogens from the production environment. However, the decrease in revenue caused by 'living with' shrimp diseases and the increased operating costs due to genetic improvement and/or pathogen exclusion have qualitatively similar effects on profit, and the proper choice between these two strategies should be based on quantitative estimates of their relative cost-effectiveness.

NOTES

B-type virus of *Carcinus mediterraneus* and WSSV of Penaeid shrimp: Similarities and possible relationships

Bonami Jean-Robert^{1, 2, 3*}, *Durand Stéphanie*⁴

- 1. CNRS
- 2. IFREMER
- 3. UM2
- 4. UnimaFrance

Nuclear envelopped rod-shaped viruses were first reported in marine crabs. The first one was found in *Carcinus maenas*, in 1974, and named B virus. Later, two similar viruses were described in *Callinectes sapidus* in 1977, named Baculo-B, and a third virus named B2 in 1987 from *Carcinus mediterraneus*. All the data available on these viruses are based on morphology and morphogenesis. Moreover, they are all located in connective cells and in non circulating hemocytes. We regrouped them under the name of B-type viruses. Since, numerous morphological similarities were noted between these B-type viruses from crabs and the White Spot Syndrome virus from shrimp. Morphologically, in TEM sections, the B-type viruses exhibit similar size and structure as the WSSV. After purification, the two possess a tail-like structure at one extremity of the envelope. Their nucleocapsids are asymmetric and segmented in appearance. Spacing of the WSSV nucleocapsid segments is the same as that of B2 (20 to 22 nm) but greater than that of the B virus from *C. maenas*. The number of nucleocapsid segments reported varies from 14 in B2 to 16 in B virus, compared to 16 in WSSV. Many morphological data are available on the ultrastructure, morphogenesis and first steps of cell infection (viral penetration) of the B-type viruses. At the opposite, no data was reported about WSSV early stages of infection. Therefore, we can hypothesize some steps in WSSV development using known data observed in the B-type viruses.

NOTES

Use of WSSV cDNA Microarray for gene profiling during WSSV infection in shrimps.

Khadijah Siti ¹, Miller Lance ², Ying Neo Soek ², Mathavan S ², Kwang Jimmy ^{1}*

1. Institute Of Molecular Agrobiolgy, THe National University Of Singapore

2. Genome Institute Of Singapore

White spot syndrome virus (WSSV) is currently a major shrimp pathogen causing high mortality resulting in huge economic losses. Although the entire genome of WSSV, which environs approximately 180 open reading frames (orfs), has been determined, the mode of infectivity is still unknown. We have approached this subject by employing microarray technology which allows one to profile the gene expression patterns of thousands of genes simultaneously under different experimental conditions. We have created clones derived from AluI restricted WSSV genome which theoretically generates about 744 fragments with an average size of 400bp to 500bp. More than 3000 of such fragments have been cloned into pbluescript plasmid and spotted on the microarray thus statistically covering the entire genome. Using fluorescent labelled probes, we compared the gene expression of commercially obtained specific pathogen free and WSSV infected shrimps. We identified genes which displayed expression ratios greater than 2.0 fold. Interestingly, 1% of the analysed data showed over-expression in SPF shrimps which indicates that these shrimps were carrier of the virus. A BLAST search revealed that these differentially regulated genes share similarity to proteins involved in signal transduction and ubiquitination. These exciting findings may shed light on the molecular mechanisms of WSSV-induced mortality.

NOTES



Variations in tandem repeat DNA segments in the ribonucleotide reductase gene of white spot syndrome virus (WSSV) isolates from Vietnam

Walker Peter J ¹, Hodgson Richard AJ ¹, Preston Nigel J ², Nguyen T Phuong ³, Dang TH Oanh ³, Tran TT Hoa ^{1}*

1. CSIRO Livestock Industries, Indooroopilly
2. CSIRO Marine Research, Cleveland
3. Laboratory of Fish Pathology, College of Aquaculture and Fisheries, Cantho University

White spot syndrome is a viral disease that affects most commercially cultivated marine shrimp species. The disease first emerged in East Asia in 1991 and has since spread throughout most shrimp farming regions of Asia and the Americas. Disease outbreaks usually result in high mortalities in affected ponds. However, shrimp may also be infected chronically with no signs of disease and often obtain the infection in hatcheries from infected broodstock. A wide range of other crustaceans can also act as asymptomatic carriers of infection. In this report, we describe the application of a 54 nucleotide tandem repeat sequence in the WSSV ribonucleotide reductase gene as a strain-specific genetic marker. We use the marker to examine the extent of variation among WSSV isolates from hatcheries different areas of Vietnam and to trace the progression of infection in ponds during grow-out. The analysis of approximately 200 WSSV isolates has shown common variations in number of repeats among these strains, with some broodstock and postlarval batches harbouring more than one genotype. In healthy ponds and in healthy broodstock or postlarval samples collected from hatcheries, viral strains containing 4-, 5-, 6- 7-, 8- and 9-repeat sequence genotypes were detected with no evidence of any predominant genotype. However, amongst ponds sampled during disease outbreaks, the 7-repeat sequence was dominant. Of ten diseased ponds sampled from three provinces in the Mekong Delta, eight were infected with the 7-repeat genotype, one with the 4-repeat genotype and one with the 9-repeat genotype. In one pond in Bac Lieu Province, several genotypes were detected in shrimp sampled at 30 days after stocking but only the 7-repeat genotype was detected in samples collected at 60 days during a disease outbreak. In another pond in Soc Trang Province, the 7-repeat genotype was detected in postlarvae with disease only 4 days after stocking. Studies in progress are investigating the possibility that the 7-repeat sequence is a reliable marker of viral virulence.

NOTES



Construction of recombinant *Vibrio harveyi* to study its adherence and pathogenicity in shrimp larvae

Suwanto Antonius ^{1*}, Widanarni ²

1. South East Asian Regional Centre for Tropical Biology (SEAMEO-Biotrop)
2. Bogor Agricultural University

Gene for green fluorescent protein (gfp) was inserted into a broad-host range plasmid, pBBR1MCS2 to generate recombinant plasmid pWG01. This constructed recombinant plasmid was introduced into a pathogenic *Vibrio harveyi* M employing triparental mating. *V. harveyi* M (pWG01) resulted in green-fluorescent colonies due to the expression of gfp. Growth curve analysis showed that the recombinant *V. harveyi* exhibited almost identical profiles to that of the wild type parental strain. However, after 48 h of incubation in antibiotic-free medium, approximately 50% of the cells had lost their recombinant plasmid carrying gfp. Expression of gfp in *V. harveyi* and its application for adherence or pathogenicity assays in shrimp larvae will be presented.

NOTES

**A hypothetical model for VHML bacteriophage conversion of vibrio harveyi***Oakey Jane* ^{1*}, *Owens Leigh* ²

1. Queensland Department Of Primary Industries, Animal And Plant Health Services
2. Department Of Microbiological And Immunology, James Cook University

The bacteriophage VHML (Vibrio harveyi Myovirus Like) originates from a toxin-producing strain of Vibrio harveyi. It has been demonstrated previously that the presence of the VHML prophage can induce virulence to V. harveyi in the laboratory. Here, a hypothetical model for the mode of action of the phage virulence conversion of V. harveyi will be presented. Through the nucleotide sequence determination of the entire VHML genome (43,193 bp), we have identified the putative structural genes of the phage virion and these were consistent with the physical characteristics of the virions as observed by TEM. We have also identified putative genes consistent with integration of the genome, supporting the theory that VHML integrates the host V. harveyi genome as a prophage. In addition, based upon nucleotide sequence of the phage DNA, we have identified a potential toxin gene on the VHML genome. This gene includes DNA sequence that is similar to the reported active site of the ADP-ribosylating group of toxins. These ADP-RT's include also toxins from other bacteria that have been previously reported to be a result of bacteriophage conversion. This presentation will illustrate how the genes upon the phage genome could cause infection of the Vibrio harveyi host cells, integration of the phage genome into the hosts' chromosome and subsequent production of the potential toxin, thereby conferring virulence to Vibrio harveyi.

NOTES

Co-detection and differentiation of yellow head complex viruses using monoclonal antibodies

Soowannayan Chumporn ^{1}, Slater Joanne ³, Cowley Jeff ⁴, McCulloch Russell ⁴, Hyatt Alexander ³ Cramerri Sandy ³, Wise Terry ³, Sithigorngul Paisarn ²*

1. Centex Shrimp, Mahidol University
2. Department of Biology, Srinakarinwirot University
3. Australian Animal Health Laboratory
4. CSIRO

Three monoclonal antibodies (MAb) raised against yellow head virus from Thailand (YHV) were tested against tissues of shrimp from Thailand, Australia, Ecuador and India purported to be infected with viruses in the yellow head virus complex. As a preliminary step, the site of reactivity of the 3 MAb in YHV nucleocapsids and mature virions was also determined by immuno-electron microscopy using ultra thin sections of YHV-infected shrimp tissue and negatively stained, semi-purified YHV particles. With MAb Y-19, gold particles deposited on YHV nucleocapsids and the inner core of mature, enveloped virions in ultra-thin sections but not on negatively-stained YHV particles, confirming its capsid protein specificity. With MAb V-3-2B, gold particles deposited on the surface of negatively stained YHV particles, confirming its specificity to the 135 kDa YHV structural glycoprotein. MAb Y-18 did not react with YHV particles either in sections or in semi-purified preparations. Immuno-histochemistry using *Penaeus monodon* infected with either YHV or GAV showed that MAb Y-19 and Y-18 reacted with both while MAb V-3-2B reacted with YHV only. All these tissue samples also gave positive in situ hybridization reactions with a cDNA probe specific to the ORF1b gene of YHV. They also gave the expected differential RT-PCR results for YHV and GAV. By contrast, 2 natural Thai shrimp specimens with no gross signs of disease gave positive immuno-histochemical reactions with MAb Y-19 and Y-18 but not with MAb V-3-2B. Although they also gave positive in situ hybridization reactions, their RT-PCR products were suggestive of GAV. Indeed, sequencing of the RT-PCR products showed that they shared 90% identity to GAV but only 80% identity to YHV. Specimens from Ecuador and India that displayed histopathology suggestive of YHV infection gave negative immunohistochemical reactions with all 3 MAb and negative in situ hybridization results. Additional work is required to determine whether a virus from the yellow head complex was responsible for the observed histopathology. These preliminary data suggest that the 3 YHV MAb could be used in diagnostic situations to identify and differentiate some viruses in the yellow head virus complex.

NOTES

Session 8 - Immunology

Innate immunity in vertebrates and invertebrates

Van Muiswinkel Willem B ^{1*}, Van De Braak Karin BT ², Rombout Jan H W M ¹

- 1. Cell Biology & Immunology group Wageningen University
- 2. Fish Culture & Fisheries Group, Wageningen University

Two systems providing internal defence against infectious agents have been selected during evolution: the innate (natural) and the acquired (adaptive) immune system. The innate immune system can be found in all multicellular animals (both invertebrates and vertebrates) and consists of cellular and humoral elements. The most prominent cellular defence reactions against invading micro-organisms are phagocytosis, encapsulation, non-specific cytotoxicity and clotting. The humoral defence factors, such as clotting proteins, agglutinins, proteases, anti-proteases and anti-microbial peptides are often produced by the defence cells. The acquired immune system is phylogenetically younger and is found exclusively in vertebrates, including fish. The acquired response is characterised by specificity and memory formation, but is usually slower than the innate response. Both responses must start with the recognition of invading micro-organisms. The innate defence system presumably recognises molecular patterns shared by large groups of pathogens, rather than specific structures. The important role of the subsequent innate response will be illustrated by data derived from functional studies using haemocytes from black tiger shrimp (*Penaeus monodon*) and macrophages or granulocytes from common carp (*Cyprinus carpio*).

NOTES

Vaccine development for Asian aquaculture

Grisez Luc ^{1*}

1. Intervet Norbio Singapore

Although more than 90% of the world's aquaculture takes place in Asia, less than 10% of the vaccine sales in aquaculture are accounted for in Asia. The reason for this marked discrepancy is not that Asian aquaculture is a recent development; in fact it could be considered as one of the pioneers with in aquaculture. It is also not because the species cultured are free from diseases. In general, the development of effective vaccines and vaccination strategies follow the development and intensification of the industry. Prerequisites for vaccine development are: 1) intensified production of a fish species with optimized management practices; 2) identification of the disease causing agents, 3) an understanding of the epidemiology of the disease, the available windows for vaccination and duration of protection needed, 4) knowledge of the immune development of the fish species and 5) a vaccine that ultimately will profit both the fish farmer and the vaccine company. Asian aquaculture is still thought of as a hit and run operation with regard to the fish species produced. Farms are mostly small, low-cost, family-owned operations. Some disease causing agents are described but comparative studies between isolates from different countries and different fish species are lacking. Epidemiology data are generally missing as are basic data on the immune system of Asian fish species. However, several universities and private companies are recently engaging in these fields and it is clear that vaccines will soon become increasingly available in Asia. The present status of 'vaccines for Asia' will be discussed with reference to specific examples.

NOTES

Immunostimulants Induced Immunity and its Quantification in Tiger Shrimp, *Penaeus monodon* (Fabricius) through ' Challenge ' and ' Pro-po Assay '

Felix Sugantham ^{1*}

1. Fisheries College and Research Institute

While immunostimulants play an important role in shrimps, their relative efficiency in terms of enhancing immunity in shrimps is still not understood. This study aims to test the relative efficiency of immunostimulants viz., β -1,3-Glucan and Lipopolysaccharide (LPS) on the giant black tiger shrimp, *Penaeus monodon* by incorporating immunostimulants in the basal shrimp diet. Glucan at 10, 20, 40 and 60 mg/kg levels and Lipopolysaccharide at 10, 20, 30 and 40 mg/kg levels for 10 days treatment were attempted. The challenge study and 'pro-PO activity' (in haemocytes and plasma) were carried out using *Vibrio parahaemolyticus* and Microplate Reader (ELISA Reader) respectively. The maximum immune enhancement (in challenge and pro-PO) was recorded for Glucan - 10 and LPS - 30 mg/kg followed by LPS - 10, LPS - 20, Glucan - 20 and Glucan - 40 mg/kg of feed. Further, all other doses except Glucan - 60 showed higher immune enhancement (in challenge and pro-PO) than that of control.

NOTES



Session 9 - Probiotics and therapeutics

Probiotic bacteria: Are they beneficial?

Rengpipat Sirirat ^{1*}

1. Chulalongkorn University

Probiotic bacteria are now widely used as prophylactic treatments with poultry, swine, and other land animals for protection against pathogenic microorganism. More recently, probiotics have been developed for aquaculture crops, but are still not widely used in aquaculture. There are, however, substantial advantages for probiotic use with shrimp and fish culture. These benefits include protection against bacterial pathogens for which there are no medications. Probiotics also do not create chemical residues that can taint animals intended for human consumption. We have demonstrated the live weight gain and more survival on *Penaeus monodon* after fed a probiont as a feed supplement and their resistance to luminescent disease infection by challenge tests including the greater immune response compared with control shrimp not fed probiotic bacterium in laboratory tanks. Possible transferring a probiont via artemia by bioencapsulation for nursing *P. monodon* larvae in hatchery was confirmed and more effective benefits on post larvae shrimp were obtained. The assessment of our probiont to earthen pond settings more similar to commercial growthout of *P. monodon* was extensively studied. Survival and growth of shrimp fed probiotic feeds were significantly greater. Disease challenge tests confirmed that probiotic feed could delay disease onset and reduce disease severity.

NOTES

Use of microbial technology to improve farm results in shrimp farms in Ecuador

Harris Lachlan^{1*}, *Alex de Wind*², *Walter Serrano*³, *David Moriarty*⁴, *Daniel Villamar*⁵

1. Seaquest SA, Guayaquil, Ecuador
2. Bravito SA, Machala, Ecuador
3. Marecuador SA, Machala, Ecuador
4. Acuabiotec LLC, Brisbane, Australia
5. Acuabiotec LLC, Milwaukee, USA

It has been a difficult struggle for Ecuadorian shrimp farmers since 2000, attempting to recover production levels since the devastating effects resulting from the spread of WSSV. However WSSV has been only one of Ecuador's disease problems. Bacterial diseases have always been present and important in Ecuadorian shrimp culture. Overcoming bacterial disease problems, using sustainable technology, is one of the major keys to producing in the presence of serious viral problems.

Coupled with improved pond management, the administration of species of *Bacillus*, used in culture water for juveniles, and applied to feed throughout the culture cycle, has been effective in controlling bacterial diseases without requiring the application of antibacterials. *Bacillus* isolates, resembling the administered strains in colonial morphology, have been re-isolated from shrimp mid-gut and hepatopancreas in concentrations sufficient to competitively inhibit pathogenic bacteria such as *Vibrio species*.

The experiences of several Ecuadorian producers will be presented, comprising more than 1000 hectares of extensive ponds. These producers stopped using antibiotics between February to July 2001 and have now produced four cycles using solely microbial technology for bacterial control. Their production has increased more than 200% since July 2001, reaching pre-WSSV levels. Key factors involved in applying microbial technology have been the control of feeding and method of stocking of juveniles, the bacterial concentration (not less than 10⁸ CFU/g) of administered *Bacillus* in feed pellets, and continual monitoring of the population of shrimp in order to augment bacterial doses during times of environmental stress or viral challenge.

NOTES



Biocontrol of Bacterial Pathogens In Aquaculture With Emphasis on Phage Therapy

Karunasagar Indrani ^{1*}, *Karunasagar Iddya* ¹

1. University of Agricultural Sciences, College of Fisheries

One of the major constraints for the development of aquaculture is the mortality due to diseases which affect both hatcheries and farms. Presently to control diseases a number of antibiotics, sanitisers and other chemicals are being used by the aquaculture industry. However, these have adverse environmental effects and emergence of antibiotic resistance, persistence of chemical residues in animal tissue are the major problems. In this context, biological control of pathogens would be a very useful strategy to prevent diseases. In this paper, our experience in controlling the major shrimp pathogen, *Vibrio harveyi* which causes luminous bacterial diseases in hatcheries and farms is described. Some aquatic bacteria such as *Pseudomonas* and *Bacillus* have been found to produce anti-Vibrio compounds. In the case of *Pseudomonas*, the compound has been found to be chloroform soluble and present in the culture supernatant. In seawater, both these bacteria were observed to reduce the levels of *Vibrio harveyi*. Bacteriophages capable of lysing several *V. harveyi* strains have been isolated from shrimp culture environments. These bacteriophages have been found to be very efficient in reducing levels of *V. harveyi* in water. Ability of bacteriophages to control luminous bacterial diseases in shrimp hatcheries were tried. Bacteriophage treatment was observed to reduce luminous bacterial counts in water and in larvae and tremendously improved larval survival. Bacteriophages acting on other vibrio pathogens in aquaculture such as *V. alginolyticus*, *V. parahaemolyticus* and *V. vulnificus* have also been isolated and found effective in reducing the respective vibrio numbers in microcosms. The results show promise. These results suggest that biocontrol of pathogens would be very environment friendly approach to manage disease problems in aquaculture.

NOTES



SYMPOSIUM ABSTRACTS

5th Symposium On Diseases in Asian Aquaculture

The probiotic potential of *Vibrio alginolyticus* (Val 1) in the oyster hatchery

Tan Cheok Keong ^{1*}

1. University Of Technology, Sydney

The present study investigated the probiotic potential of a strain of *Vibrio alginolyticus* (Val 1), for use in oyster hatcheries. The results found that rearing Sydney Rock Oyster (*Saccostrea glomerata*) larvae in autoclaved seawater and in the presence of *V. alginolyticus* improved larval growth and survival by 15µm and 40% respectively, than when compared to larvae reared in 1µm filtered seawater with the absence of *V. alginolyticus*. Larval survival and growth were also compared for different seawater treatment, with and without *V. alginolyticus*. The treatments were 1µm filtered; 0.2µm filtered; activated carbon filtration; pasteurisation at 65oC and 85oC; chlorination; U.V.; and chlorination followed by activated carbon filtration. The results showed larval growth and survival in 1µm-filtered seawater without *V. alginolyticus* were higher than the larvae reared in the other seawater treatments, irrespective of the presence or absence of *V. alginolyticus*. This suggests that in the absence of an outbreak, treatment of seawater and the presence of *V. alginolyticus* provided no beneficial effect on the growth and survival of *S. glomerata* larvae. When *S. glomerata* larvae reared in 1µm-filtered seawater were subjected to a challenge with the pathogen *Vibrio tubiashii*, larval survival decreased from 62% in the unchallenged control to 8% in the challenged control. In contrast, larval reared in the presence of *V. alginolyticus* challenged with *V. tubiashii* demonstrated 35% improvement in survival to that of the challenged control. Thus, indicating a substantial beneficial effect. These findings indicate the potential of *V. alginolyticus* as a probiotic for the prevention of bacterial epidemics in oyster hatcheries.

NOTES

Antagonistic activity of *Aeromonas media* strain A199 against *Saprolegnia* sp. in two species of finfish, the eel *Anguilla australis* and silver perch *Bidyanus bidyanus*

Lategan Josie ^{1*}

1. University Of Technology Sydney

Antagonistic activity of *Aeromonas media* strain A199 against *Saprolegnia* sp. in two species of finfish, the eel *Anguilla australis* Richardson and silver perch *Bidyanus bidyanus* Mitchell. *Aeromonas media* strain UTS A199, an organism isolated from fresh water is a potential probiotic for use in the aquaculture industry against *Saprolegnia* sp, the oomycete associated with "winter kill", environmental stresses and farming practices that may cause the loss of fish stock. *Aeromonas media* produces a bacteriocin-like inhibitory substance (BLIS) that has shown a wide spectrum of in vitro antagonistic activity against various fish pathogens. In this study, BLIS inhibited in vitro the growth of *Saprolegnia* sp. isolated from affected fish. Inhibition was observed against both the vegetative state of the aquatic mould and cyst germination. A number of separate in vivo tank observations involving affected eels revealed that the daily addition of A199 to tank water contributed to the subsequent swift recovery of affected hosts from invasion by this opportunistic pathogen. While, in a small scale, pilot trial, to test the effectiveness of A199 during an outbreak of saprolegniosis, silver perch displaying early lesions of the disease were maintained in tank water containing A199 for a period of 3 weeks. Eleven percent (11%) accumulated mortality was observed for fish exposed to A199 for the three-week treatment, increasing to thirty-three percent (33%) in the week post treatment. In contrast, 77%-accumulated mortality was obtained for fish not exposed to A199, with the majority of saprolegniosis related deaths occurring during the first two weeks of the study. The discovery of both in vitro and in vivo antifungal activity by *A. media* appears to be a promising novel approach in the use of probiotics for the management of the opportunistic oomycetes that lead to the rapid death of affected fish.

NOTES

**Session 10 - Mollusc Health 1****Perkinsus disease in Korean waters; taxonomy, distribution, diagnostics and their effects on clam ecology***Choi Kwang-Sik*^{1*}, *Kyung-II Park*¹

1. Cheju National University

Perkinsosis is a shellfish disease caused by parasitic protozoan in some commercially important mollusks including oysters, clams, abalones and scallops. Heavy infection with Perkinsus often results in mass mortalities and commercial loss. Currently Perkinsus is classified as a molluscan disease in the OIE. In Korean waters, Perkinsus sp. has been found in the Manila clams, *Ruditapes philippinarum*. Perkinsus is also believed to be responsible for the decline in clam landings for the past decade in Korea. Perkinsus trophozoites were distributed commonly in gills, digestive glands, mantle and gonadal connective tissues. They were relatively scarce in foot and siphons. Heavy infection with Perkinsus often caused white nodule formation on gills and mantle, as well as massive concentration of haemocytes around the infected tissues. Microscopic features of different life stages and 5.8S rRNA nucleotide sequences read from the non-transcribed spacer and internal transcribed spacer indicated that Perkinsus sp. discovered in the clam in Korea could be *P. atlanticus* in European waters. Ray's fluid thioglycollate medium (RFTM) with Choi's NaOH lysis method was used in identification of Perkinsus in the clams. The prevalence was mostly 80 to 100% in commercial clam beds on the west and south coast of Korea. The infection intensity was found to be highest in October when most clams completed spawning and mass mortalities were observed in the fields. Quantification of clam eggs using enzyme-linked immunosorbent assay with rabbit anti-clam eggs protein IgG demonstrated that the amount of eggs produced during spawning was negatively correlated with infection intensity of Perkinsus. In conclusion, high level of Perkinsus infection in the clam could precipitate reduced growth and reproduction, as well as mass mortalities in the field, resulting in decreases in clam harvesting in Korean and possibly in other Asian waters.

NOTES

Diseases of pearl oysters

Jones Brian ^{1*}

1. Department of Fisheries Western Australia

The aquaculture of pearl oysters is an expanding multi-million dollar industry in the tropical marine environment of many countries, including Australia, French Polynesia, the Middle East, SouthEast Asia and Japan. Despite the size and extent of the industry there is remarkably little known about the diseases and parasites of the genus *Pinctada*. There is a growing awareness among the industry that, as with other molluscs under cultivation, disease can be an important constraint and that translocation of shellfish poses a serious risk. This paper will review known diseases caused by pathogenic agents as well as those with a non-infectious aetiology. Management techniques, which can be used to minimise the impact of disease, will also be discussed.

NOTES



SYMPOSIUM ABSTRACTS

5th Symposium On Diseases in Asian Aquaculture

Report on oyster mortality in Wonboyn Lake, Australia

Ogburn Damien^{1*}, *Callinan Richard*¹, *Hallegraeff Gustaaf*², *Landos Matthew*¹

1. New South Wales Fisheries
2. University Of Tasmania

In March 2002, a major mortality of Sydney rock oyster (*Saccostrea glomerata*) was reported on aquaculture leases in Wonboyn Lake (latitude 37° south) in southern NSW. This paper describes the investigations surrounding the mortality event and the subsequent conclusions. Gross lesions in the stomach epithelium of oysters was observed. The dinoflagellate *Prorocentrum minimum* has been implicated, together with a changing ecology in the lake resulting from a restricted sea entrance due to sand accretion. The strategy for ongoing management and monitoring for the oyster industry in Wonboyn Lake and potentially elsewhere is discussed.

NOTES

Grouper hatchery health in the APEC region: surveys of hatcheries and nurseries and regional experts and institutions involved in grouper health

Hayward Craig ^{1*}

1. Department of Microbiology and Parasitology, The University of Queensland

An APEC Fisheries Working Group workshop, entitled "Development of a Regional Research Program on Grouper Virus Transmission and Vaccine Development", was held in Bangkok in October 2000. To assist in implementing some of the workshop's recommendations, four surveys were prepared at the Network of Aquaculture Centres of Asia-Pacific (NACA) and the Aquatic Animal Health Research Institute (AAHRI) of Thailand. Questionnaires were designed to assess the impacts of diseases among grouper hatcheries and nurseries, and to inventory the scientists and laboratories working on grouper diseases in the region, to help pinpoint expertise and capacity for contribution to the proposed Collaborative Resource Centre. Surveys were distributed to representatives of 13 APEC economies involved in the production of groupers. Four respondents to the hatchery and nursery health questionnaire listed 7 cases of disease in the last 12 months in 4 countries. These included one diagnosed and two suspected cases of infection with a nodavirus (VNN), one occurrence of vibriosis, one case of leech infestation, and a disease of unknown aetiology characterized by deformed skeletons and floating larvae. A further two respondents reported no diseases among larvae in their hatcheries. These responses will be accessible in Epilnfo databases.

NOTES



Breeding for QX disease *Marteilia sydneyi* resistance in Sydney rock oysters *Saccostrea glomerata*

Nell John^{1*}, *Hand Rosalind*¹

1. NSW Fisheries

The Sydney rock oyster, *Saccostrea glomerata*, breeding program was established by NSW Fisheries in 1990 to provide the industry with a faster growing oyster. The Georges River component of the breeding program was severely interrupted by the appearance of QX disease in this estuary in 1994. Selection for QX disease resistance *Marteilia sydneyi* in Sydney rock oysters, in Georges River commenced in 1997. Wherever QX disease has appeared, the local oyster industry has severely declined, slowly in three northern NSW rivers, taking 30 years for a 56% decline in production, but quickly in Georges River, Sydney, taking only 7 years after the parasite was detected for a 97% decline in production. Selection for QX disease resistance is important in case yet another estuary should suffer this fate. The progeny of second generation Sydney rock oyster breeding lines were tested for resistance to QX disease *Marteilia sydneyi* against a non-selected control. Mortality was reduced from 86% for the controls to 64% for the most improved breeding line. This is a reduction in mortality of 22% after only two generations of selection. These partially QX disease resistant oysters in which *M. sydneyi* was found were also 21% heavier than controls. Selection for resistance to *M. sydneyi* is feasible and may be improved through further selection.

NOTES

Session 11 - Mollusc Health 2

Diseases in mollusc hatcheries: A paradox in health management

Berthe Franck CJ^{1}*

1. IFREMER

The world aquaculture of molluscs is still increasing, reaching 10.73 millions of tonnes in 2000. Among the 42 cultivated mollusc species, 5 species are dominating the global production, which the Pacific cupped oyster, *Crassostrea gigas*, is leading. In many countries, there is a traditional mollusc aquaculture, usually based on wild stocks. However, these natural populations often do not fulfil the market demand - because of their poor value, over-fishing of the resource or impact of diseases - and an answer to this has frequently been the introduction of new stocks or new species. Furthermore, genetic improvements, availability of juveniles, as well as species diversification for aquaculture do increase the demand for international movements and transfers of live molluscs. Diseases have become a primary constraint to mollusc aquaculture growth and sustainability, given their severe impact on socio-economic development in many countries. Diseases are a major threat to aquaculture, natural resources alike. Pathogen transfers via transfers of live molluscs are currently recognised as a major cause of disease outbreaks and epizootic. In this prospect, hatchery production may be seen as a way to provide disease-free batches of juveniles and therefore a pivotal tool in effective programs to prevent the transfer of infected stocks in disease free areas. On the other hand, several diseases are known to occur in hatcheries that could be disseminated with release of hatchery products in grow-out areas. After reviewing the importance of hatcheries of molluscs and molluscs diseases in hatcheries, their paradox in health management approach will be discussed.

NOTES



Survey on the ovarian parasite, *martellioides chungmuensis* in the cultured Pacific oysters *Crassostrea gigas* in Korea

Park Mi Seon ^{1*}

1. National Fisheries Research and Development

In Korea, oyster culture began in early 1900's and commercial oyster culture using hanging culture system started in 1960's in the southern coast. After 1960's, production amount of cultured oyster has been increased every year until late 1980's and the highest production amount as shellstock was recorded 288,078 metric ton in 1987. But increment of the production amount in 1990's has shown a stagnant status. Production amount recorded 174,117 metric ton as shellstock in 2001. From 1990's, oyster culture industry in Korea has faced hard circumstances because of insufficient seed collection and mass mortality of the cultured oyster. National Fisheries Research and Development Institute (NFRDI) presumed contaminants from inland side, self contamination in the growing area and reactivation of broodstock induced the results. The ovarian parasite of oyster has been supposed a cause of the bad seed collection and mass mortality. In Korea, the parasite was reported first time in 1970 and then presumed belonged to *Acanthamoeba* (Chun, 1979). Comps et al. (1986) performed morphological identification of the parasite isolated from the Pacific oyster collected at Chungmu area in Korea and named the parasite *Martellioides chungmuensis*. The infection rate of *M. chungmuensis* in Korea has been increased every year and appearance period of the parasite also also has extended from spawning season to all the year round. The regulatory authorities such as Minister of Maritime Affairs & Fisheries (MOMAF) and provincial government have performed growing area cleaning project for a better condition keeping. And also NFRDI recommend oyster industry triploid oyster that is not infected by the parasite.

NOTES

Marteilioides chungmuensis (paramyxea), an intracellular parasite of the ovocyte of Pacific oyster *Crassostrea gigas*: isolation and sequencing of small subunit ribosomal DNA

Itoh Naoki ^{1*}, *Oda Tadashi* ², *Ogawa Kazuo* ¹

1. University Of Tokyo

2. Fisheries Experiment Station Of Okayama Prefecture

Marteilioides chungmuensis is the intracellular parasite of the ovocyte of Pacific oyster *Crassostrea gigas* in Japan, causing irregular enlargement of the infected ovary. Infected oysters lose their marketability due to the unaesthetic appearance. Although the parasite causes a serious economical impact on oyster fisheries, biological aspects of the parasite including the infection route, multiplication stages and early infective stage remain unknown. Recently, molecular biological techniques have become powerful tools to detect microorganisms or discover unknown stages of parasites. We developed a molecular detection method for *M. chungmuensis* using parasite DNA as follows. Infected gonads were frozen at -20oC to rupture parasitised ovocytes and homogenized with glass homogenizer. Sporonts were collected with nylon meshes to remove host cells and debris, and purified with discontinuous sucrose and Percoll gradients. Further, we extracted parasite DNA from the sporonts and sequenced partial 18s small subunit ribosomal DNA (ca. 1200 bp). Two specific probes were designed based on the sequence, and in situ hybridization was applied on histological sections. Positive signals were recognized only on parasite cells, confirming the sequenced DNA derived from *M. chungmuensis*. These results provide basic molecular tools which would be helpful to elucidate the life cycle and phylogenetical position of this economically important parasite.

NOTES



Epizootiology and detection of nocardiosis in oysters

Bower Susan M^{1}, Carnegie Ryan B¹, Meyer Gary R¹*

1. Fisheries and Oceans Canada

Nocardiosis is a bacterial disease of oysters caused by *Nocardia crassostreae*. This disease has many common names, some of which describe its pathology, including: fatal inflammatory bacteraemia, focal necrosis, multiple abscesses and Pacific oyster nocardiosis (PON). Infection with *N. crassostreae* induces a massive accumulation of haemocytes resulting in the formation of green coloured lesions or pustules up to 1 cm in diameter in the mantle, gill, adductor muscle, and heart of oysters. However, histological evidence suggests that some oysters are capable of ridding themselves of infection by the process of diapedesis (haemocyte migration through intact epithelium). First described as a fatal disease among Pacific oysters (*Crassostrea gigas*) in Matsushima Bay, Japan in the 1960s, it has since been reported from various locations along the west coast of North America but the true geographic distribution is not known. Recent investigations in British Columbia clearly indicate that this bacterium is also pathogenic and lethal to flat oysters (*Ostrea edulis*). Infection and mortalities seem to be highest among beach cultured oysters, especially those on a muddy substrate. However, other environmental factors such as reduced water circulation in shallow embayments and warm temperatures also increase the prevalence of infection and severity of nocardiosis. Although eradication of nocardiosis is not feasible, off-bottom culture seems to mitigate the disease. The potential negative impact of this disease on oyster culture around the world warrants further investigations and precautions against the transplantation of infected oysters. A polymerase chain reaction (PCR) assay for management and study of nocardiosis is currently undergoing validation.

NOTES

Diseases of cultured paua (*Haliotis iris*) in New Zealand

Diggles Benjamin^{1*}, *Oliver M*¹

1. National Institute of Water and Atmospheric Research

Mortalities of cultured paua (*Haliotis iris*) in New Zealand have been associated with a variety of disease agents. During the summers of 1999/2000 and 2000/2001, mortalities of between 82.5 and 90% of juvenile paua (*Haliotis iris*) were reported in a commercial culture facility. Affected paua exhibited behavioural abnormalities including lethargy, loss of righting reflex, and easy detachment from surfaces. Histology of moribund paua showed heavy infections of a novel haplosporidian. Laboratory experiments failed to transmit the haplosporidian horizontally by cohabitation or by injection of healthy paua with hemolymph containing haplosporidian plasmodia. Spore formation was not observed in juveniles, but sporocyst-like bodies containing acid-fast putative spores were observed in the right kidney of poorly performing adult paua collected from the wild. The epidemiology of the haplosporidian disease in affected culture facilities remains poorly understood. A survey of 1094 paua collected from 5 spat producing farms and 3 grow out farms during the summer of 2001/2002 failed to detect the haplosporidian. A number of other potential disease agents and syndromes were detected, however, including rickettsial inclusions in the gut, haemocytic neoplasia, granuloma-like lesions in internal organs, and erosion of external epithelia associated with bacterial infection and ectocommensal ciliates. Other disease agents which affect the shell of cultured paua include a mycosis and mudworm infestation. The possible effects of these disease agents on paua farming in New Zealand are discussed.

NOTES



Discovery of the early infective stages of the protozoan parasite *Marteilia sydneyi* in oysters and the implications for disease detection and control

Kleeman Sarah^{1*}, *Adlard Rob*², *Lester Bob*³

- 1. Aquatic Animal Biosecurity Australia, Agriculture Fisheries Forestry Australia
- 2. Protozoa Section, Queensland Museum
- 3. Department Of Microbiology And Parasitology, University Of Queensland

Marteilia sydneyi, the aetiological agent of QX disease, causes persistent mortalities in commercial rock oysters on the east coast of Australia. In order to diagnose QX in oyster tissue, the OIE recommends various examination procedures involving surveillance, presumptive and confirmatory techniques. Specifically: histological examination is considered the most suitable technique for surveillance of disease; tissue imprints allow rapid presumptive analysis of disease following mass mortality outbreaks, and; electron microscopy and in situ hybridisation are often required for species identification following detection by the aforementioned means. However, most of these techniques have relied on the presence of sporulating stages residing in the digestive gland of the oyster host, which, until recently, were the only lifecycle stages known. The discovery and characterisation of the initial infective stages of *M. sydneyi* in the gills and palps of oysters, as well as presporulating stages in the digestive gland and connective tissue, has not only presented new opportunities for disease diagnosis and detection in *Marteilia* species but has also identified limitations in some existing methods. This paper discusses the possibility of improved protocols, as well as the value of existing protocols, in the surveillance, monitoring and species confirmation of *M. sydneyi* in light of these new data.

NOTES

Session 12 - Trans-boundary and emerging diseases

Limitations to preventing increased international distribution of aquatic animal diseases

Hill Barry ^{1*}

1. Centre for Environment, Fisheries and Aquaculture Science

The belief that serious diseases of aquatic animals have been, or could be, introduced into their territory from abroad has led some countries to impose strict controls on imports of live and dead aquatic species. Although national quarantine and health certification requirements for imports are certainly a valid part of first line defence against introduction of exotic diseases, they must be developed within the context of international standards (WTO SPS Agreement and the OIE Code and Manual) and should not be used as an unjustified barrier to competitive trade from other countries. Even so, it is important also to recognise that such legal import safeguards alone may not necessarily prevent the sudden appearance of a serious disease in a country from which it was previously believed to be absent. Reasons include failure of an import risk analysis, ineffective surveillance for the disease/pathogen in the exporting country and inadequate enforcement of the import restrictions (e.g. illegal imports). Furthermore, there are other possible pathways for pathogen introduction than just the importation of live or dead aquatic animals (or their products for human consumption) e.g. dead wild fish as fresh feed for farmed fish or as fishing bait, live fish transport vehicles that have been used in other countries and possibly ships' ballast water. Finally, rather than recent importation of the pathogen being the cause of a first-time occurrence of an 'exotic' disease, there is the possibility of the emergence of a more virulent strain of the agent that has existed benignly in a local reservoir without previously being detected. This is particularly relevant to marine diseases and the author will present some possible examples.

NOTES

Ornamental disease vectors

Ariel Ellen ^{1*}

1. EU Community Reference Laboratory for Fish Disease

The ornamental fish industry transfers large quantities of fish between many countries around the world thereby presenting significant potential for trans-boundary transfer of disease agents. Although tropical ornamental fish are usually kept in indoor aquaria and are unlikely to survive for long if released into the natural environment in temperate regions, they are capable of carrying viruses that pose a threat to temperate fish species. Of particular concern are the ranaviruses which have been reported in several tropical ornamental species in Australia, the USA and Israel. Some of these ranaviruses are highly pathogenic to certain temperate fish species, both farmed and wild. The so-called 'cold water ornamentals' are fancy varieties of common species, mostly cyprinids, and do survive and breed in temperate regions as well as in tropical countries. The most widely traded of these species internationally are goldfish (*Carassius auratus*) and koi carp (*Cyprinus carpio koi*) and both species have been implicated in trans-boundary transfer of viral diseases to farmed fish species such as carps and also to wild fish populations. A major example is koi herpes virus disease, which has been spread to several countries via this trade and is causing considerable economic damage to the industry. More so, in Indonesia this disease is currently suspected to be responsible for recent largescale mortalities not only in koi carp but also in common carp varieties farmed as food, and imports of koi carp are suspected to be the route of entry. Such events are likely to lead to increased demands for health certification for ornamental fish, which may in the short term cause restrictions on trade, however, in the long term it is likely to aid the sustained development of the industry.

NOTES



Fatal, virus associated peripheral neuropathy and retinopathy (PNR) in farmed *Penaeus monodon* in Eastern Australia

Callinan Richard ^{1*}, *Jiang L* ¹, *Smith P T* ², *Soowannayan C* ³

- 1. NSW Fisheries, Regional Veterinary Laboratory
- 2. Centre For Sustainable Aquaculture, University Of Western Sydney
- 3. Centex Shrimp, Faculty Of Science, Mahidol University

Outbreaks in farmed *Penaeus monodon* of a previously unreported, fatal disease, 'peripheral neuropathy and retinopathy' (PNR) are described. These outbreaks, associated with minor to heavy mortalities, occurred in most ponds on an eastern Australian farm during two consecutive growout periods. Moribund prawns, 5-26 g mean body weight, were typically reddish in colour, lethargic, with partially amputated appendages. Histologically, mild to severe degeneration and necrosis of axons and their sheaths, together with associated glial cell apoptosis, were consistently present in peripheral nerve fibres. Mild to severe, acute to chronic retinitis, associated with degeneration and necrosis of reticular cells and their axons, also occurred in most clinically affected prawns. Intracytoplasmic nucleocapsids and enveloped virions, morphologically consistent with a yellow head related virus, were present in peripheral nerve and eye lesions. Immunohistochemical examinations were conducted using monoclonal antibodies reacting with both yellow head virus, considered exotic to Australia, and with the closely related gill-associated virus (GAV), widely endemic in *P. monodon* in eastern Australia. Positive reactions were consistently observed in lesions, but not histologically normal tissues, in peripheral nerves, eyes, lymphoid organ and vas deferens. Findings strongly suggest that a yellow head related virus, most probably GAV, is the causal infectious agent of PNR, an emerging disease of *P. monodon*. It is also likely that PNR is a component disease within the ill-defined 'mid crop mortality syndrome'. Although confirmatory transmission trials remain to be done, measures to prevent further spread of GAV, both within Australia and beyond, should be seriously considered.

NOTES



Field investigations on serious disease outbreak among Koi and Common Carp (Cyprinus carpio) in Indonesia

Sunarto Agus ^{1}, Taukhid ¹, Rukyani Akhmad ², Supriyadi Hambali ¹, Koesharyani Isti ¹, Huminto Hernomoadi ³, Agungpriyono Dewi Ratih ³, Pasaribu Fachriyan H ³, Widodo ³, Hardiawan Dwika ², Rukmono Puguh ², Nilawati ², et al*

- 1. Fish Health Research Laboratory, Central Research Institute for Aquaculture
- 2. Directorate of Fish Health and Environment, Directorate General of Aquaculture
- 3. Faculty of Veterinary Medicine, Bogor Agricultural University

Since April 2002, disease outbreaks have occurred in cultured *Cyprinus carpio* (koi and common carp) in Java Island, Indonesia. The outbreak caused huge economic losses and significant social impact. For instance, in Blitar regency, East Java, it has wiped out koi carp belong to 5,000 fish farmers with economic losses more than Rp. 5 billions (US\$ 5 millions) within 3 months periods. To prevent the spread of the outbreak to other island, the government of Indonesia has closed Java Island from any movement of koi and common carp. This paper described the clinical history, gross signs, histopathology, PCR detection and experimental infection of the disease. The outbreak occurred after heavy rain, movement of adult fish to other pond or transportation of fry to other area. This phenomenon leads to the hypothesis that the virus was latent and becoming active under particular circumstances, such as stress of transportation and handling and environmental changes particularly temperature fluctuation. Another scenario was that the disease occurred through trans-boundary movement of infected koi carp from Hongkong. The disease occurred in on-growing fish of all ages and in all culture system including stagnant, running water and cage culture system. A variety of symptoms have been reported from infected fish. Infected fish may be lethargic, show loss of balance and gasp for breath. Sloughing off the epithelium with loss of mucus, and rough, hemorrhage of operculum, fins, tail and abdomen are common symptoms. However, the only consistent clinical sign of the disease is gill necrosis. In the early stage of infection, the gill filaments showed typical focal necrosis. In the late stage, the necrotic gill filaments fused and badly damaged. However, the symptoms of the disease may be complicated by secondary infection of debilitated fish by opportunistic organisms such as bacteria, fungi and parasites. *Flexibacter columnaris* and *Aeromonas hydrophila* have been isolated from necrotic gill filaments and skin ulcer, swollen kidney and liver, respectively. *Dactylogyrus* sp., *Trichodina* sp, *Ichthyophthyrus multifiliis* and *Argulus* sp has also been found in some infected fish. Attempts have been made to treat the infected fish with Kalium Permanganat and antibiotics (Enrofloxacin, Erythromycin, Amoxicilin and Oxytetracyclin), but it has no prevail. Histopathological study revealed necrotic changes in the gill, fin, skin, kidney, spleen, liver, heart and intestine. Prominent basophilic intranuclear inclusion bodies were observed in the gill and kidney of infected fish. Experimental infection by cohabitation test and injection with 0.45 μ m-filtered homogenate resulted 100% and 70% mortality, respectively. Based on the clinical history, gross signs and histopathological changes, experimental infection and polymerase chain reaction (PCR) detection of naturally and experimentally diseased fish, it is strongly suspect that Koi Herpesvirus (KHV) is involve on the serious outbreak on koi and common carp in Indonesia. This is the first KHV outbreak reported in Asian region.

NOTES

Session 13 - The Future**Improving aquatic animal health in Asia**

Subasinghe Rohana P^{1}*

1. Food and Agriculture Organization of the United Nations (FAO)

Asian aquaculture has expanded, diversified, intensified and advanced technologically. Its growth and contribution to national economic and societal goals are expected to continue, provided enabling environments for sustainable development are established. Bitter experiences and substantial economic losses have demonstrated that good health management is a key to success in aquaculture. Prevention and control of disease, avoidance of introductions and transfers of pathogens, development and adoption of better farm management practices, capacity building, and targeted research, are a few vital components of good health management. Although, progress has been made in Asia, much remains to be done to achieve adequate capacity in aquatic health management.. Rapid sector growth increases the risk of unsustainable development, thus the urgency for achieving necessary management capacity. Globalization and liberalization of international trade will also require adherence to international health guidelines and standards if Asian aquaculture is to maintain its place in the international market place. Increased consumer awareness of food quality and safety, animal welfare and the environment will also exert pressure in local and regional markets. These are some of the issues that need to be addressed. The development and implementation of integrated, practical health management strategies to this end, which include appropriate regulatory frameworks and enforceable laws, will only be possible with relevant national policy and appropriate institutional arrangements. While researchers and scientists must continue to provide the necessary scientific base, through targeted research and dissemination of information, ensuring institutional, financial and human capital will continue to depend on the political will.

NOTES



Aquaculture health management: The Australian experience

East Iain ^{1*}

1. Aquatic Animal Health, Agriculture, Fisheries & Forestry - Australia

Aquaculture is of increasing economic importance in Australia. The gross value of production (A\$750 million in 2000-01) now exceeds 30% of the total value of fisheries, with major contributing sectors being pearl farming, tuna cage culture, salmon culture, edible oysters and prawn culture . Recognising the critical importance of aquatic animal health for overall profitability, competitive trade advantages and public health, Australia in 1998-99 signed on to 'AQUAPLAN', the National Five-Year Strategic Plan for Aquatic Animal Health. AQUAPLAN is a world-first as a joint industry-government strategic initiative in aquatic animal health. AQUAPLAN provides a framework for health management that is underpinned by both legislation and policy. It is a coordinated approach that emphasises stakeholder consultation and industry participation. The major aims of AQUAPLAN are to support industry profitability and maintain the quality of the aquatic environment. Because Australian waters are contiguous with Asia, international linkages and regional cooperation are a key component of AQUAPLAN. Recent key achievements include the implementation of the Building a National Approach to Animal and Plant Health initiative, the establishment of the Fisheries Research and Development Corporation's Aquatic Animal Health Subprogram, and the steps taken to establish the Aquatic Animal Health Consultative Committee as the primary government/industry interface committee for policy, communication and awareness related to aquatic animal health issues. A critical outlook towards the future will also be given.

NOTES

The role of extension in effecting on-farm practice change for controlling shrimp disease

Foster Derek ^{1*}, *Demaine Harvey* ²

1. Queensland Department of Primary Industries
2. Aquaculture and Aquatic Resources Management

The history of participatory extension methods throughout Asia and the Pacific illustrates the effectiveness of these methodologies in creating sustainable changes to farming practices. By involving farmers in research on farms it is possible to construct appropriate technologies to improve farming practice. The ACIAR project 'Development and Delivery of Disease Control Programs to Small Scale Shrimp Farmers in Indonesia, Thailand and Australia (FIS/2000/061)' is designed to create effective processes for controlling shrimp diseases such as white spot and yellow head. The project builds on technologies developed in a previous project and focuses on developing more sustainable aquaculture systems for each of the participating countries. This contextual relevance for specific extension approaches and resulting technologies means that there is a fusion between scientific and social systems in play. The paper will describe extension processes used and the role of extension in creating on farm adoption and adaptation of technologies. In outlining the main methodologies used in each country, the paper will define the theoretical underpinnings of these methods and explain the reason why different methods suit different countries. The paper contends that without effective social research and extension program design there will be little adoption of appropriate technologies. No matter how good the scientific research, there will be little change if people participating in the aquaculture industry do not contextually apply technologies. Extension is one key factor for successful control of shrimp disease in the Asia/Pacific region.

NOTES



Theme 1

Quarantine detention of ornamental fish - Practical challenges involved in the implementation of a new policy

Colquitt Stephen^{1}, Stevenson Robyn²*

1. Biosecurity Australia
2. Australian Quarantine & Inspection Service, Agriculture, Fisheries And Forestry - Australia

Based on the findings of an import risk analysis (IRA), Australia introduced new biosecurity requirements for the importation of ornamental finfish in November 1999. All ornamental finfish imported into Australia must meet pre-export and post-arrival biosecurity requirements before release into the aquarium trade, including the satisfactory completion of post-arrival detention in quarantine facilities approved by the Australian Quarantine and Inspection Service. These import controls address risk factors identified in the IRA and are designed to reduce the likelihood of exotic fish diseases and pests establishing in Australia, to an acceptable level. Although these controls are amongst the most stringent in the world, their practical implementation has not been without problems. This presentation will describe some of the challenges that Australia has faced in the 3 years following the introduction of new import controls in 1999, and based on these experiences, explore ways to better manage biosecurity risks associated with imported ornamental fish in the future.

NOTES

Theme 2**Argulosis in Brood Carp Rearing Ponds of Bangladesh**

Ahmed Abu Tweb Abu ^{1}, Chowdhury Mamun ¹*

1. Department of Zoology, University of Dhaka

Recent investigation on the infection of a new species of Argulus Muller shows that Argulosis is a common problem in almost all carp hatcheries of Bangladesh, and the farmers are using the pesticides- Cymbush and Sumithion at regular intervals. As a result, the pond ecology as well as the abundance of zooplankton were found to be adversely affected. All the 3 major carps viz. Catla catla, Labeo rohita and Cirrhinus mrigala are infected and shows the clinical signs of agitation, lethargy, sporadic movement, cessation of feeding, shortage of ovulation, opaqueness and darkness of skin, shrunken eyes and skin erosion. The infected brood fish becomes unfit for breeding as well as table fish thus causing economic loss. Out of 1440 fishes studied from two infected ponds of BRAC hatchery at Rajendrapur near Dhaka, the overall prevalence of Argulus infestation on L. rohita, C. mrigala and C. catla was 50%, 36.66% and 12.91% respectively in pond no.3 and 26.66%, 20% and 10.41% respectively in pond no.1. Whereas the overall intensity of infestation was 13.78, 11.37 and 9.6 in pond 1 and 10.6, 9.97 and 9.22 respectively in pond 3. The prevalence and intensity of infestation in all the 3 species of fish of both the ponds were highest in the largest length group of 60-74.9cm in the month of July, and female fishes were more susceptible to infestation. To overcome the hazards of using pesticides, a mechanical means of using pyrex sheets to collect and destroy Argulus eggs and biological control measures of culturing predatory organisms like Macrobrachium with the brood carps are being tried

NOTES



Freshwater fungi isolated from common carp (*Cyprinus carpio*) eggs in Thailand

Chukanhom Kanit ^{1, 2*}, Hatai Kishio ¹

- 1. Division Of Fish Diseases, Nippon Veterinary And Animal Services University
- 2. Faculty Of Veterinary Medicine, Khon Kaen University

Fungal infection in eggs of freshwater fishes is well known as problematic disease. We had a chance to examine the fungal infection in common carp (*Cyprinus carpio*) eggs at fish farms in Khon Kaen Province, Northeast of Thailand in February 2002. We attempted to isolate fungi from the eggs with fungal infection at three fish farms. Each egg with a fungal infection was placed directly on a GY agar plate, and then a small amount of streptomycin sulphate and ampicillin was scattered on the medium to retard bacterial contamination. All agar plates were incubated at 25°. Nineteen fungi from the farm A, two fungi from the farm B and two fungi from the farm C were isolated. The fungi from the farms A, B and C were identified as *Saprolegnia diclina*, *Achlya* sp. and *Allomyces arbuscula*, *Achlya* sp., and *Saprolegnia diclina*, respectively. *S. diclina* and *Achlya* sp., and *A. arbuscula* grew well at 25-30° and pH 5-7, and at 37° and pH 6-8, respectively. Artificial infection to platy (*Xiphophoras maculatus*) was made using the selected fungi. For isolates *S. diclina* and *Achlya* sp., the mortality in injured fish challenged with 10⁴ zoospores/ml was 100%. But the mortality was 0% in the other experiments

NOTES

Tasmanian Isolates of *Streptococcus* sp. biovar 1 and verified strains of *Lactococcus garvieae* and *Enterococcus seriolicida* compared by microbiological , molecular biological and "in vivo" techniques

Hawkesford Tiina^{1*}, *Carson Jeremy*², *Burke Chris*³, *Munday Barry*³, *Oakey Jane*⁴

- 1. Queensland Fisheries Service, Department of Primary Industries, Brisbane, Queensland, Australia
- 2. Department of Primary Industries Water and Environment, Launceston, Tasmania, Australia
- 3. University of Tasmania, Launceston, Tasmania, Australia
- 4. Animal & Plant Health Service, Department of Primary Industries, Townsville, Queensland, Australia

A gram-positive coccus was isolated from diseased, farmed rainbow trout and ocean-caged trout (*Oncorhynchus mykiss*) in Tasmania in 1988. This disease caused significant commercial loss to the trout farming industry at that time. Based on morphological, physiological and biochemical characteristics of the Tasmanian isolates, the initial identification placed them in the genus *Streptococcus* and they were hence given the epithet *Streptococcus* sp. biovar 1. Subsequent studies indicated that these organisms were more closely related to the genus *Enterococcus*, especially the fish-pathogenic bacterium known as *Enterococcus seriolicida*, than the genus *Streptococcus*. Further investigation has indicated that these fish pathogenic isolates could be identical to the species *Lactococcus garvieae* isolated from cattle. In order to clarify the situation, studies were undertaken using molecular biology to investigate the possibility of some genetic similarity. The technique used for this investigation was the random amplified polymorphic DNA polymerase chain reaction (RAPD-PCR). Pathogenicity of the isolates was also performed using rainbow trout (*Oncorhynchus mykiss*). The results of all these studies showed that the Tasmanian type strains (*Streptococcus* sp. biovar 1) were closely related to, if not identical with, both *Lactococcus garvieae* and organisms previously named *Enterococcus seriolicida*. Therefore *Streptococcus* sp. biovar 1 should be reclassified as *Lactococcus garvieae*, which must now be considered a major fish pathogen worldwide.

NOTES

A study on columnaris disease in guppy *Poecilia reticulata*

Hettiarachchi M ^{1*}, Hettiarachchi DC ²

1. Department Of Zoology, University Of Kelaniya
2. Confifi Aquaculture Ventures (Pvt) Ltd

About 65% of ornamental fish exported from Sri Lanka is guppy and guppy breeders in the country have been trying to produce fish with wider caudal fin to suit the requirement in the export market. This attempt has been hampered by the presence of larger proportion of male guppies with eroded margins of caudal fins and the occurrence of subsequent mortalities. Therefore the present study was undertaken to investigate the cause of fin erosion and mortality in guppy. Monthly samples of guppy were obtained from a guppy farm in the Western Province of Sri Lanka that had the experience of having fish with eroded caudal fins. These fish were maintained and observed in the laboratory under simulating conditions to those of the farm. Within 48 hours, erosion of the margin of caudal fin progressed gradually leaving the fin rays bare and lesions developed on the caudal peduncle, body surface and on other fins which rapidly progressed in to yellow to orange coloured ulcers. Tips of gill lamellae of some fish showed necrosis which progressed rapidly reaching the base of the gill arch. Histopathology revealed that the ulceration could spread in to deeper tissues. In wet mounts of lesion, long thin rods (0.60 to 1.0 μm width; 5.0 to 10.0 μm length) with characteristic gliding motion of *Flexibacter columnaris* were present. The bacteria produced yellow-green flat colonies with uneven margins on cytophaga agar. Percentage prevalence of the disease was increased during the months with water temperature above 28.5 oC causing 85 - 91% mortality within a period of 8 days. Water quality management together with prophylactic treatment is suggested to control *F.columnaris* in culture facilities of guppy.

NOTES

Nodavirus infections as an emerging disease in aquaculture

Kapo Nime ^{1*}

1. National Agriculture Quarantine and Inspection Authority

[Please note: The summary below is done as a requirement for necessary funding to attend the symposium. It was necessary for me to do it in order for my funding agency to allocate finance. It is hoped that I can learn a lot more than what I know now, from attending the symposium. It is based on an incomprehensive review of recent literature on the subject] **ABSTRACT/SUMMARY** Several RNA and DNA viruses of lower vertebrates have induced diseases resulting in important economic losses, especially in fish aquaculture. One of the emerging RNA virus is the nodavirus, also called nervous necrosis virus (NNV). The disease caused is called viral nervous necrosis (VNN), sometimes also called viral encephalitis and retinopathy (VER). It is one of the OIE's notifiable fish pathogenic RNA viruses. The same viral diseases have been observed in barramundi, sea bass, turbot, groupers and other fish in the world. It mainly affects larvae and juveniles via vertical or horizontal transmission and causes a vacuolating encephalopathy and retinopathy. The virus strains have adapted to different geographic locations and environments and could possibly extend its host range easily. Diagnostic procedures recommended are at present electronic microscopy, immuno histochemistry, fluorescent antibody techniques, ELISA, PCR and viral isolation. Control measures include prevention of vertical spread by selection of non-infected spawners, sterilisation of eggs and larvae, and possibly vaccination of juveniles at the end of the nursery period. There is still need for continued epidemiological surveillance of fish nodavirus and the development of effective control measures.

NOTES

Winter disease in farmed silver perch (*Bidyanus bidyanus*) in New South Wales, Australia

Landos Mathew ^{1*}, *Callinan Richard* ¹, *Rowland Stuart* ², *Read Phil* ², *Boyd Peter* ², *Nixon Mark* ², *Mifsud Charlie* ², *Beakes Gordon* ³

1. NSW Fisheries, Regional Veterinary Laboratory
2. NSW Fisheries, Grafton Aquaculture Centre
3. Department Of Biological And Nutritional Sciences, The University, Newcastle Upon Tyne

Health related losses of silver perch cultured in earthen ponds are the subject of a three year research project which began in 2001. Winter disease, characterised by severe dermatitis and branchitis associated with *Saprolegnia parasitica* infection, is responsible for a significant proportion of these losses. Outbreaks of winter disease cause mortalities between 1% and 100% in fish >80g. Typically affected fish have large, irregular, pale areas on skin and/or gills, with attached fungal hyphae. *S. parasitica* has been isolated consistently from these lesions. The fungus does not provoke a significant inflammatory response in the skin and does not penetrate the stratum compactum. Where gill is infected, there is extensive destruction of gill lamellae by invasive fungi. Moderate to severe proliferative branchitis, unrelated to fungal infection, is always present in fish prior to outbreaks. A temporal association between infestation by the monogenean gill parasite *Lepidotrema bidyana* and this gill lesion is being investigated. No significant lesions have been identified in internal organs of affected fish. Timing and severity of outbreaks varies within and between farms. Outbreaks occur below 16 oC water temperature and often coincide with rapid drops in water temperature. Physical damage to skin from handling has also preceded some outbreaks. Occurrence does not appear to be correlated with fluctuations or absolute levels of pH, hardness, alkalinity, DO, or TAN. Improved understanding of causal factors and therapeutic agents are required for cost-effective control.

NOTES



First report of systemic amoebosis in oscar, *Astronotus ocellatus*

Laoprasert Thitiporn^{1*}, *Kanchanakhan Somkiat*¹, *Chinabut Supranee*¹, *Hatai Kishio*²

1. Aquatic Animal Health Research Institute, Department of Fisheries, Thailand
2. Division of Fish Diseases, Nippon Veterinary and Animal Science University, Japan

Systemic amoebosis was found in oscar (*Astronotus ocellatus*). Diseased fish showed loss of appetite, emaciation, skin darkening, small lesion on the body surface, aggregation on the bottom of the tanks. Mortality rate at 80-100 percent was found within 3-5 days after disease onset. Causative pathogen was isolated and maintained in pure culture. Morphological study by light microscope was conducted and found two distinctive forms, trophozoites which were spiny-like pseudopodium and cysts which showed in polygonal shape. Application of Polymerase Chain Reaction technique with specific primer and nucleotide analysis of PCR product indicated that this organism is an amoebic protozoa belonging to the genus *Acanthamoeba*. Keyword: Amoebosis, oscar (*Astronotus ocellatus*), *Acanthamoeba*

NOTES

Study on Tetrahymena infection in guppy (*Poecilia reticulata*)

Laoprasert Thitiporn ^{1*}, Chinabut Supranee ¹, Hatai Kishio ²

1. Aquatic Animal Health Research Institute, Department of Fisheries, Thailand
2. Division of Fish Diseases, Nippon Veterinary and Animal Science University, Japan

Recently Tetrahymena infection is serious problem in guppy (*Poecilia reticulata*). Infected fish were collected from guppy farms in Bangkok and the other areas nearby. The clinical sign of infected fish is whitish patch associated with lesion on the body. Fish scales are bristled, in serious infection, outer skin and muscular tissues was lost. The infection results in mortality within short time. Diseased fish were observed under light microscope and histopathologically studied. Large number of ciliated protozoas are presented on skin and in scale pocket. It was identified as Tetrahymena sp. Histopathological studies showed severe extensive necrosis occur in subdermal and muscular tissues. These ciliated protozoa could invade through skin into muscle and also internal organs. Inflammatory reaction was rarely observed. It was possible to conclude that these ciliated protozoa were the main causative agent of the heavy mortality in guppy. The endemic of this ciliated protozoa may relate to the importation of ornamental fish into the country without the proper quarantine system. The suitable health management and quarantine of new stock before introducing into the farms is recommended for preventing of this disease.

NOTES



Mycobacteriosis in ranchu, goldfish (*Carassius auratus*) imported from Japan

Lawhavint Ong-Ard^{1*}, *Wada Shinpei*², *Hatai Kishio*²

- 1. Faculty of Veterinary Medicine, Kasetsart University
- 2. Division Of Fish Diseases, Nippon Veterinary And Animal Science University

Ranchu, Japanese goldfish, is very famous for culturing among Thai people. Each year, many company have imported ranchu from the famous farm in Japan. In the past winter, January 2002, 30 percent of imported juvenile fish showed inactive swimming, anorexia and lethargy. Therefore, the six diseased fish were collected to investigate the cause of diseases. The clinical signs of the fish showed asymmetrical swelling of abdomen, pale discoloration of gills, swelling and discoloration of head and trunk kidneys. Abdominal organs were attached to peritoneal wall, indicating peritonitis, associated with numerous white nodules of various sizes. These nodules were also found on the abdominal organs, head and trunk kidneys and heart. The isolation was done with Ogawa egg medium and identified as *Mycobacterium chelonae* by the PCR method at Aquatic Animal Health Research Institute, Department of Fisheries, Thailand. The histopathological features showed that these white nodules on the affected organs were chronic proliferative lesions composed of multiple caseous epithelioid cell granulomata with surrounding granulation tissue. In the central caseous area, a number of colonies of slender, long rod, Gram positive were found to be acid-fast by Ziehl-Neelsen method. These acid-fast bacteria were also observed in the epithelioid cells and in the macrophages of the granulation tissue. From these findings, this case was diagnosed as serious systemic mycobacteriosis.

NOTES



Hemorrhaging septicemia due to *Aeromonas hydrophila* in the Mekong catfish (*Pangasius bocourti*) cultured in An Giang province - Vietnam

Nguyen Huu Dung ^{1*}, *Hua Thi Phuong Lien* ²

1. Faculty of Aquaculture - University of Fisheries - Vietnam
2. Faculty of Agriculture and Natural Resources - An Giang University - Vietnam

Hemorrhaging septicemia disease has been observed in the Mekong catfish *Pangasius bocourti* cultured in earth ponds and floating net cages in An Giang province since the early 1990's. The infection caused serious losses to the industry due to high mortalities and/or reduced value of chronically infected fish. In order to identify the causative agent, 497 infected fish were collected from various fish farms in the province in four years from 1997 to 2000. The samples were used for histopathological, parasitological and bacteriological examinations. Various parasitic and bacterial species were detectable from the collected samples. However, the motile Gram-negative strains of *Aeromonas hydrophila* showed to be the causative agent as it was isolated at highest frequency from the liver, spleen and kidney of all of the infected fish. The infection was also successfully reproduced in experimental fish of 15g in body weight being injected with 0.1ml of the isolated *A. hydrophila* strains with LD50 = 106,41CFU/fish. The pathogenic isolates showed resistance to common antibiotics i.e. penicillin, ampicillin, oxytetracycline, streptomycin, sulfamethoxazol, and chloramphenicol. Drug resistance of the isolates has resulted in difficulty for controlling the disease. However, initial study on vaccination development using the isolated strains has showed that water-based heat- and formalin-killed vaccines did not produce effective protection in the experimentally vaccinated fish.

NOTES



Production and characterization of monoclonal antibodies to Singapore grouper iridovirus (SGIV)

Qin Qiwei^{1, 2}, Shi Chengyin¹, Gin Karina Yew Hoong³, Lam Toong Jin^{1, 2}*

- 1. Tropical Marine Science Institute, National University of Singapore
- 2. Department of Biological Science, National University of Singapore
- 3. School of Civil and Environmental Engineering, Nanyang Technological University

A panel of 6 monoclonal antibodies (mAbs) against a grouper iridovirus (SGIV) were produced by immunization of Balb/c mice with purified virus preparations. Isotyping test revealed all the mAbs were IgG1. None of the mAbs possessed ability to neutralize SGIV in cell cultures but all reacted with the cytoplasm of SGIV-infected grouper cells (GP) as determined by an indirect immunofluorescence test (IIF). Western blot assay showed that 4 mAbs reacted with 2 SGIV proteins at molecular mass of approximately 100 and 117 kDa in gradient-purified virus. Fractionations of the iridovirus in a 20-60% sucrose gradient were successfully detected by all the six mAbs using immunodot blot. These mAbs will facilitate the development of more specific and standardized diagnostic techniques for marine fish iridovirus.

NOTES

Aquatic oomycetes from southeast Queensland

Rayamajhi A^{1*}, *Lester RJG*², *Hayward CJ*²

1. Department Of Microbiologoy & Parasitology
2. University Of Queensland

The occurrence of two oomycetes, *Aphanomyces* sp. and *Achlya* sp. in freshwater fish, *Scortum barcoo*, from a local fish farm led to an investigation of the moulds that were present in local waterways. Oomycetes were initially cultured on Czapek Dox Agar and later on split autoclaved chickpeas in autoclaved pond water with antibiotic (Penicillin G, 100 units/ml and oxolinic acid 100 mg/ml). Four *Achlya* species were identified, *A. americana*, *A. bisexualis*, *A. caroliniana* and *A. proliferoides*, and two *Saprolegnia* species, *S. luxurians* and *S. terrestris*, each by the morphology of their sporangia, oogonia and antheridia. *Aphanomyces* sp. was frequently isolated but cultures failed to develop sexual stages and therefore isolates were not identifiable to species. Goldfish from which 3 scales had been removed were exposed to an isolate of *Aphanomyces* sp. or an isolate of *Achlya* sp. for 2 hr. then kept at 18.5, 24.5, or 27.5 C. No infection developed over the next 3 weeks.

NOTES

**A new genus of dracunculoid nematode from the gills of the pufferfish *Tragulichthys jaculiferus****Ribu DL*^{1*}, *Lester RJG*²

1. Department Of Microbiology & Parasitology

2. University Of Queensland

A new dracunculoid species belonging to the Guyanemidae is described from gill filaments of the pufferfish *Tragulichthys jaculiferus* Cuvier (Tetraodontiformes: Diodontidae, 'green porcupine fish') from Moreton Bay, Queensland. The species has a reduced buccal capsule and divided oesophagus. Females have a functional vulva and single ovary; males have caudal alae and spicules. These characters are typical of Guyanemidae. Of the three known genera, *Guyanema* and *Travassosnema* are exclusively parasites of freshwater fishes in South America while *Pseudodelphis* is a parasite of the tidepool sculpin in Canada. All existing species are parasitic in the body cavity except for *T. travassosi* which occurs in the eye. The new species differs from members of these genera in that it has fine cuticular transverse striations, two forwardly protruding cephalic elevations, a circumoral elevation, a small oval mouth surrounded by a peribuccal membrane, 6 internal cephalic papillae arranged in two clusters of three papillae each and a pair of large oval external cephalic papillae. The males have only two pairs of pedunculated caudal papillae supporting the caudal alae. Gills of all 69 green porcupine fish contained mobile larvae in the gill filament between the epithelial basement membrane and the efferent artery. Abundant larvae resulted in mild oedema in the filament. Eleven fish harboured adult nematodes in the same location with little apparent tissue response.

NOTES

Identification of a betanodavirus isolated from viral nervous necrosis-diseased redspotted grouper (*Epinephelus coioides*) cultured in Southern Thailand using PCR and sequence analysis

Roongkamnertwongsa Somporn ¹ *, Danayadol Yaowanit ¹, Kanchanakan Somkiat ³,
Direkbusarakom Sataporn ⁴

- 1. National Institute of coastal Aquaculture
- 3. Aquatic Animal Health Research Institute
- 4. Walailuk University

A viral agent was successfully isolated from redspotted grouper seeds (*Epinephelus coioides*) exhibiting viral nervous necrosis (VNN) clinical signs in Southern Thailand in March 2001. Grouper seeds, ~1.5-2 cm in total length, had been collected from the wild and maintained in private nursing farms. The VNN disease occurred within a few days after stocking. Eyes and brains tissue of diseased fish were pooled, extracted and filtered through 0.45 µm disposable filters. The filtrates were inoculated on to striped snakehead whole fry tissue or SSN-1 cells and incubated at 28°C. Cytopathic effect (CPE) was first observed on day 3 post-inoculation which recognized as shrunken and rounded shape of cells. Cells continued to aggregate and the CPE completed on day 7. Viral particles in virus-infected SSN-1 cells were seen using a transmission electron microscope. The isolated virus possessed icosahedral nucleocapsid with ~25 nm in diameter. Viral propagations were not sensitive to chloroform or IUDR, which indicated that they were naked virions and contained RNA genome. These viral characteristics were primarily classified as a viral member of Nodaviridae family. Viral identification had been conducted using polymerase chain reaction (PCR) and sequence analysis of PCR product. PCR amplification using specific primers designed from Betanodavirus genotype RGNNV gave a better PCR product intensity in agarose gel than an amplification using specific primers designed from genotype SJNNV. One 730 bp PCR product was sequenced in both directions. Sequence analysis of this product using Blast program showed 720/730 or 98% nucleotide homology to dragon nervous necrosis virus and redspotted grouper nervous necrosis virus. Only a 400 bp part of the sequence was most similar to genotype SJNNV with 357/400 bp or 89% homology. Findings indicate that this first isolated Betanodavirus in Thailand can be identified and grouped as nodaviral genotype RGNNV.

NOTES



Health problems of captive West Australian dhufish

Stephens F J ¹ *, Raidal S R ¹, Jones B ¹, Thomas T ¹, Jenkins G ², Cleary J ¹

1. Aquatilia Healthcare

The dhufish *Glaucosoma hebraicum* is a potential aquaculture species but captive dhufish experience several health problems, including exophthalmos in otherwise apparently normal fish and infestation of gills with a monogenean parasite, *Haliotrema abaddon*. Gas and haemorrhage was present in the choroid of exophthalmic eyes, with haemorrhage in retrobulbar tissues resulting from perforation of the sclera in some eyes. Oxygen content of gas in eyes with recently developed exophthalmos was high (up to 73%). In some eyes with retrobulbar haemorrhage, oxygen tension approached zero, indicating severe disruption of blood supply to the eye. Oxygen tension at the retinal-vitreous junction of normal dhufish eyes was high (344 ± 26 mm Hg), with oxygenated blood supplied to the choroid body from the gills via the pseudobranch. The finding of a single haemoglobin with pronounced Root and Bohr effects in dhufish was significant and may contribute to the susceptibility of the species to exophthalmos. Investigations suggest that exophthalmos is physiological in origin and is related to the environmental differences between the natural habitat of the fish and the conditions that are experienced in aquaculture. Rapid changes of temperature or blood acid-base characteristics may precipitate the development of exophthalmos. The monogenean parasite, *Haliotrema abaddon*, was described and stages of its life-cycle identified. Potential treatments were investigated using *in vitro* and *in vivo* studies. Praziquantel was identified as the most effective 'in water' treatment of fish infested with *H. abaddon*. Other useful but less effective and safe treatments were low salinity baths (<1.5 ppt for ninety minutes) and 0.5 mg L⁻¹ trichlorphon for 36 hours.

NOTES



Histopathological study on tetrahymena infection in dwarf gourami (Colisa lalia)

Wakita Kunika ^{1*}, Imai Soichi ², Hatai Kishio ¹

- 1. Division Of Fish Diseases, Nippon Veterinary And Animal Science University
- 2. Division Of Veterinary Parasitology, Nippon Veterinary And Animal Science University

Tetrahymena infection in guppy (*Poecilia reticulata*) caused by *Tetrahymena corlissi* is well known as the most problematic disease in Southeast Asian countries. Recently, the similar parasitic ciliate was found in dwarf gourami (*Colisa lalia*), which was imported from Southeast Asian countries to Japan. The fish sometimes had ulcer lesions on the body surface. The fish usually died of the infection within several days after their arrival. We had a chance to examine some dwarf gourami with ulcer lesions on the body surface, which were imported from Singapore to Japan, and attempted to isolate the parasite from the lesion and fixed in 10% phosphate buffered formalin solution for histopathological examination. The cultured ciliates were stained with Protargol method for morphological characteristics. As a result, the ciliate was identified as *T. corlissi*, which has been reported as a pathogen of guppy. In histopathological examination, the parasites were mainly found in the connective tissues between muscle fibers, and a few ciliates invaded into abdominal cavity. The inflammatory reaction in dwarf gourami mainly consisted of macrophages and neutrophils, and was intensive than that in guppy.

NOTES

Turbot culture - a newly established industry in China and its disease problems

Wang Yin-Geng ^{1*}, Li Qiu-Fen ¹, Cheng-Yin Shi ¹

1. Chinese Academy of Fishery Sciences

Turbot (*Scophthalmus maximus*) is one of the most important culture fish species in Europe, from where it has been introduced into China in 1992. With the consequential successes in artificial breeding from 1998, the commercial culture has rapidly spread along the coast of North China. In 2001, about 3 to 4 million juveniles were produced, and yielded more than 3000 MT marketable fish with the culture area of 600,000M², which totally valued about 140 million US dollars. The turbot was mostly cultured in indoor cement tank, which is an intensive culture system equipped with oxygen generator and flowing water. At such conditions, 60-90% survival rate and biomass of 15-20kg/ M² (less than 18-month old) were normally achieved. In addition, general husbandry for broodstock, hatchery and on-growing aspects were described in the present paper. The rapid expansion of turbot culture led to the occurrence of diseases. There were several diseased conditions have been found, some of the syndromes were firstly recognized in turbot culture. In which, the non-infectious disease included depigmentation and deformities, and infectious disease were displayed with viral, bacterial and parasitic infections. The epidemiology and prevention skills were provided for each particular condition, as well as the limiting factors in the sustainable development of turbot culture in China were also discussed. **Key Words:** turbot (*Scophthalmus maximus*) fish aquaculture disease

NOTES



SYMPOSIUM POSTER ABSTRACTS

5th Symposium On Diseases in Asian Aquaculture

Pilchard herpesvirus in Australasia 1995-1999

Whittington Richard ^{1*}, Jones Brian ²

1. The University of Sydney
2. Fisheries Department of Western Australia

Two epizootics have occurred in populations of the Australasian pilchard *Sardinops sagax neopilchardus* in waters of southern Australia. The first occurred between March and September 1995, and the second in 1998-1999. In 1995 mortalities occurred along more than 5000 km of the Australian coastline and also affected pilchards in New Zealand. It was thought to be the largest fish kill ever recorded. The disease front spread from its origin in South Australia at about 30 km/day, often against prevailing currents and was not impeded by storm events. Thus it was not caused by planktonic toxins/pathogens. Affected fish died within minutes of signs of respiratory distress. Fish were sampled before, during and after the advancing mortality front. Relevant lesions were confined to gills and were unlike lesions associated with known gill pathogens or toxins in other species of fish. Lesions were initially focal but became locally extensive then generalised, with inflammation then being replaced by epithelial hypertrophy and hyperplasia over about 4 days. The pathology in affected fish across the distribution of the disease was similar, suggesting a common aetiology. A herpesvirus was the only factor consistently associated with lesions, including those in early stages of the disease. The herpesvirus was not isolated in fish cell lines but has been detected by PCR. Approximately 60% of the total pilchard biomass in Southern and Western Australian waters was lost in the 1998-1999 epizootic, but spread was slower than in 1995. The source of the virus remains controversial. Studies of the virus and its relationship to lesions are on-going.

NOTES



Bacterial infection in Tasmanian farmed abalone: causes, pathology, farm factors and control options

Handlinger Judith ^{1}, Carson Jeremy ¹, Donachie Linda ¹, Gabor Les ¹, Taylor David ¹*

1. Department of Primary Industries, Tasmania

Abalone aquaculture based on artificial feeds and intensive generally land-based systems is increasing in Australia. Intensive systems are vulnerable to diseases of poor environment control. This paper reports several types of bacterial diseases investigated in grow-out stock of *Haliotis rubra*, *H. laevigata* and their hybrids over approximately 10 years, their pathology, farm factors precipitating disease outbreaks, and the potential for disease control using antibiotics. *Vibrio* species infections have been the most common cause of infection, with a variety of species isolated from moribund animals, generally irrespective of the primary insult. Specific disease outbreaks, with differing pathology, have been associated with two species, *Vibrio harveyi* and *V. splendidus* L. Specific pathology has also been associated with *Flavobacterium*-like species. In most cases stress factors precipitating disease has been identified, and control has been largely directed to eliminating this stress. Antibiotics have also been used, with equivocal results, precipitating preliminary studies of antibiotic absorption and efficacy in these species. The limited potential for antibiotic use, and the requirement for understanding and control of on-farm stress, is discussed.

NOTES

Transmission of perkinsus olseni among wild blacklip abalone in South Australia

Hayward Craig ^{1*}, Lester Robert ¹, Barker Steven ¹, McCallum Hamish ², Murrell Anna ¹, Kleeman Sarah ¹

- 1. Department of Microbiology & Parasitology, The University of Queensland
- 2. Department of Zoology and Entomology, The University of Queensland

The protozoan *Perkinsus olseni* was recently confirmed to be widespread among molluscs in the Pacific Ocean by rRNA NTS sequence data; the parasite also occurs in the North Atlantic. In temperate waters off southern Australia, *P. olseni* infects aggregations of abalone in the wild. Individuals that develop lesions in response to the parasite are unmarketable; at other localities, infections may be acute, leading to dieback of abalone populations. To develop appropriate management strategies, we aim to model the infection dynamics of the parasite. Samples of healthy blacklip abalone were tagged and translocated to a site east of Taylor Island, a known 'hotspot' for the disease in South Australia, in two consecutive summers. In 2002, the proportion of abalone acquiring *Perkinsus* three months after translocation was markedly lower than in 2001 (11% cf. 85%); infections were also lighter. Similarly, resident abalone were infected at a lower rate this year, both at the beginning and end of the experiment (19% cf. 50% in late January; 30% cf. 57% in early May). This fall in prevalence is consistent with a cooling of the maximum sea surface temperature this summer by almost 3 C, to below 20 C. Other than blacklip abalone and one *Roe's* abalone, no molluscs in the vicinity of the Taylor Island site were hosts of *P. olseni*, and this will simplify construction of the epidemiological model.

NOTES



Investigations on Microbial Pathogens Associated with Diseased Oysters

Natarajan P¹, *Rajan AN¹, *Ginu Baby¹, *Reshmi¹****

1. Department of Aquatic Biology & Fisheries, University of Kerala

The pearl and edible oysters, collected from Tuticorin of Tamil Nadu and Astamudi Lake of Kerala, India were subjected to pathological investigations. A total of 94 specimens of pearl oysters (*Pinctada fucata*) and 130 specimens of edible oysters (*Crassostrea madrasensis*) were examined. A few pearl oysters were found to have mantle discolouration and damaged tissues on the mantle and some edible oysters were characterized with abnormal yellowish ulceration on the mantle. Bacterial samples were taken from the diseased sites of both pearl and edible oysters. A total of 3 bacterial samples from pearl oysters and 2 from edible oysters were isolated. The isolated bacterial strains were tested for their pathogenicity (Koch's postulate). Among the three bacterial isolates, only one bacterium was found to be pathogenic against pearl oyster and one among the two of edible oysters was pathogenic. These two pathogenic bacterial isolates were subjected to several biochemical tests. Based on the biochemical properties, the bacteria, pathogenic for pearl oysters were characterized to be *Vibrio sp.*, and that of edible oyster was *Bacillus sp.* These two bacterial pathogens were tested against ten antibiotics for their susceptibility. The most effective antibiotic against *Vibrio sp.*, was Chloramphenicol and that of *Bacillus sp.*, was Kanamycin. Invitro experiments were also carried out to test extracts of certain natural products for their bioactivity.

NOTES



THEME 4

Development of a real-time PCR assay for the detection of *Piscirickettsia salmonis*

Corbeil Serge¹, McColl Ken¹, Crane Mark¹

1. CSIRO Livestock Industries

Rickettsia are a group of gram-negative bacteria which are obligate, intracellular parasites that can infect a range of vertebrate and invertebrate hosts, including finfish, and aquatic invertebrates. *Piscirickettsia salmonis* epizootics in several countries have been responsible for significant economic loss to the salmonid aquaculture industry. Recently, there have been a number of reports of Rickettsia-like organisms (RLOs) associated with disease in other farmed fish (e.g. tilapia, white sea bass) and marine organisms (e.g. abalone, crayfish). The threat of the possible introduction of *P. salmonis* to Australia has prompted the development of an RLO-specific, real-time PCR assay that provides a rapid means of identifying RLO-infected animals. In this report we describe the use of real-time PCR as a diagnostic tool for the specific detection of laboratory grown exotic strains of *P. salmonis*. This study was partly funded by FRDC project number: 2001/624

NOTES

Development of diagnostic antibodies specific for white spot virus

Crane Mark ^{1}, Slater Joanne ¹, White John ¹, Hyatt Alex ¹*

1. CSIRO Livestock Industries

White spot disease (WSD) of penaeid prawns, first reported from farmed *Penaeus japonicus* in Japan in 1993, has since spread throughout the prawn farming regions of Southeast Asia and beyond. Over this period, there has been a huge effort throughout the region to develop diagnostic tools for the detection and identification of white spot virus (WSV) infected prawns, including a number of PCR assays as well as immunoassays based on WSV-specific polyclonal and monoclonal antibodies. At the Australian Animal Health Laboratory (AAHL), polyclonal and monoclonal antibodies have been raised against white spot virus and it is anticipated that such antibodies will underpin development of a number of diagnostic tests. Progress on this project is reported here. White spot virus was imported from Thailand and expanded using experimental infections of penaeid prawns. Following infection, prawns were bled, and virus was purified from the prawn haemolymph using sucrose gradient centrifugation. Purity of viral fractions was monitored using SDS-PAGE and electron microscopy. Following purification, viral fractions were used to immunise rabbits and mice for production of specific antibodies. Out of a total of 1500 hybridoma supernatants, screened by ELISA, 20 cultures were demonstrated to contain antibodies with high activity. Preliminary characterisation of the polyclonal and monoclonal antibodies specific for white spot virus is presented. This study was partly funded by ACIAR project number FIS9698

NOTES



Functional microarray analysis of Japanese flounder *Paralichthys olivaceus* immune related genes for selection of a disease resistance fish

Hirono Ikuo^{1*}, *Aoki Takashi*¹

1. Tokyo University of Fisheries

The use of DNA markers to define the genotype and predict the performance of an animal is a powerful aid to animal breeding. One strategy is known as marker-assisted selection (MAS). Recently, a new technology Microarrays has developed in parallel with the genome projects of humans and other model organisms. Microarrays allow the gene expression profiles of tens of thousands of genes to be compared in a single experiment. The use of high throughput DNA microarray technology is thought to be revolutionize animal breeding in near future. In view of the above reasons, we constructed cDNA microarray and to assess a possibility of using fish cDNA microarray for selection breeding. We used about 850 different cDNA clones from more than 3,000 cDNA clones which were identified by the expressed sequence tag (EST) analysis of Japanese flounder, *Paralichthys olivaceus* liver, spleen, skin, hirame rhabdo virus infected Japanese flounder leukocytes and Ig positive cells, and ConA/PMA or LPS treated leukocytes and kidney cells cDNA libraries for making cDNA microarray. The target mRNAs were prepared from LPS or ConA/PMA treated peripheral blood leukocytes (PBLs) and kidney and rhabdovirus infected PBLs of Japanese flounder. The target mRNAs were labeled with either Cy3 or Cy5 using the commercial labeling kit. After hybridization, the slide was scanned using the Genepix400B Scanner. The expression patterns and the amount of expressed mRNAs of these genes were different in the various genes examined, stimulations and time points. These gene-expression profiling studies of Japanese flounder have suggested some key molecular players in a large number of immune-regulated genes with unknown immunological function. All of our results suggest that the using microarray technology will be enable for selection breeding in aquaculture.

NOTES



Development of onfarm diagnostics

Kurcheti Pani Prasad^{1*}

1. Central Institute of Fisheries Education

As industrialization and human activity has expanded and intensified, the microbes and diseases have also increased exponentially, putting tremendous pressure to find ways and means for early detection and control of these pathogens. The first step in the successful control is the early detection of microbes and immediate control measures. The different methods of disease detection are: (i) Gross symptoms (ii) Post mortem examination (iii) Wet mount (iv) Electron microscopy (v) Histopathology (vi) Microbiology (vii) Immunodiagnosics (viii) DNA based diagnostics. The need of the hour is to have rapid, on-site immunodiagnosics which do not need sophisticated equipments and trained manpower. Antibodies are host proteins produced in response to the presence of foreign molecules in the body by the immune system. Functionally they are characterised by their ability to bind to specific antigens. These antibodies are specific for each antigen and are used for the development of immunodiagnostic kits like 1.ELISA test 2. Dot-ELISA test 3. Latex agglutination test The paper describes the development of these rapid techniques for identification of bacteria (*Vibrio alginolyticus*) and viruses (Yellow head virus), the advantages of each method and their stability.

NOTES

Plasmid profile of bacterial isolates from white spot affected shrimps

Mulloorpeedikayil Rosalind George^{1}, Riji John¹, Jeyaseelan M J Prince¹, Prasad P S Sathish¹, Iyappan T¹, Lidwin Anna Mary¹, Sundararaj V¹*

1. Fisheries College and Research Institute

Samples of white spot infected shrimps (*Penaeus monodon*) were collected from the East coast of India. The samples were analysed and 84 presumptive vibrio isolates were collected from shrimps exhibiting various degree of infection. These isolates were examined for their antibiotic resistance. Presence of plasmids was found in strains exhibiting multiple drug resistance. Restriction analysis of the plasmids indicated that isolates contained plasmids belonging to different groups. The study demonstrated that the plasmid profiles of the isolates could be used as a useful indicator to investigate the strain variability of the *Vibrio* strains. Plasmid curing studies conducted revealed that some of the strains have antibiotic resistance associated with R factor connected with plasmids. This could imply that the resistance transfer could happen from the farm isolates to microbes of public health significance.

NOTES



Experimental Bacteriophage-mediated virulence in strains of *Vibrio harveyi*

*Munro James*¹, *Oakey Jane*¹, *Bromage Erin*¹, *Owens Leigh*¹

1. James Cook University

Vibriosis is a major disease of prawn aquaculture. Until now there has been no clear explanation why some strains of *Vibrio* are pathogenic, while others are not. This study demonstrated that the presence of the bacteriophage, *Vibrio harveyi* Myo-Like virus (VHML) may confer virulence in *Vibrio harveyi* strain 642. This was demonstrated by infecting naive avirulent *V. harveyi* strains 12, 20, 45 and 645 with the bacteriophage and converting them into virulent strains. The previously naive strains of *Vibriosis* infected with the bacteriophage, VHML, from *V. harveyi* strain 642 demonstrated up-regulation of haemolysin, up-regulation of protein excretion, additional proteins which were recognised as toxic proteins from strain 642 by monoclonal antibodies specific to the exotoxin sub-units and a significant increase in mortality of larval *Penaeus monodon*. It was concluded that the bacteriophage VHML conferred virulence to *V. harveyi* strain 12, 20, 45 and 645 and that the bacteriophage VHML either fully or partly confers virulence in *V. harveyi* strain 642.

NOTES

Development and characterization of a monoclonal antibody against white-spot syndrome virus in Penaeid shrimp

Nomura Nakao¹, Tanaka Michiaki¹, Hagio Maiko¹, Matsumura Masatoski¹

1. Institute Of Applied Biochemistry, University Of Tsukuba

White spot syndrome virus (WSSV) is a major viral disease agent of penaeid shrimp. Number of studies have been done for basic research and development of a diagnostic kit for WSSV. One of the promising diagnostic tools to detect viral pathogens in shrimp farms must be the immunological methods by using monoclonal antibody because it doesn't need any expensive equipment and time for the analysis can be very short. However, since WSSV is a virus encapsulated with envelope protein and the envelope protein are very fragile and easily destroyed by ordinary virus purification procedure including the steps that gives physical stress to WSSV. Purification of pathogen is an essential step for a manipulation of monoclonal antibody to develop a diagnostic kit. This work will focus on the development of a new method to purify the virion of WSSV and to obtain IgG monoclonal antibodies against envelope proteins. The monoclonal antibodies that recognize WSSV envelope proteins, VP29 and VP19 specifically were successfully isolated from this work. The antibodies obtained here can be used for detection of WSSV in infected *Penaeus japonicus*. After confirmation of the antibodies specificity against WSSV from different countries, these antibodies would be a great help to develop a diagnostic kit for detection of WSSV in shrimp farms in several countries. Moreover, some of the antibodies obtained in this study must have the activity to block the viral infection because it is known that envelope proteins play an important role in WSSV infection. The investigation on the blocking activity of the obtained antibodies is now undergoing. The antibody that can block viral attachment to the host shrimp cell will be a powerful tool for the farther study on the infection mechanism of the WSSV.

NOTES



Detection and comparison of lymphocystis virus in flounder (*Paralichthys olivaceus*) and sea bass (*Lateolabrax japonicus*)

Park Myoung-Ae^{1, 2 *}, *Do Jeong-Wan*¹, *Lee Ju-Seol*¹, *Park Mi-Seon*¹ *Park Jeong-Woo*¹

- 1. The Korean Society of Fish Pathology
- 2. The Japanese Society of Fish Pathology

Lymphocystis disease has occurred in various kinds of marine fin fish in the world. This study was performed both to explore the host spectrum of lymphocystis virus (LV) among cultured marine fishes and to examine the phylogenic position of the LV in Korea. LV-infection was confirmed on the basis of histopathological examination, analysis of structural protein and nucleotide sequence analysis of partial gene, then LV was identified from fishes including flounder (*Paralichthys olivaceus*) and sea bass (*Lateolabrax japonicus*). LV-Infected fishes developed lymphocystis on the body surface, fin, gill and intestine. Electron microscopic observation revealed that the virus was large icosahedral structure, 150-200nm in diameter that assembled in cytoplasm of infected cell. Infected cell was encapsulated by hyaline extracellular matrix and contained basophilic intracytoplasmic inclusion and condensed nucleus. Viral proteins of the two LVs isolated from flounder and sea bass were analyzed by SDS-PAGE, and the resulting electrophoretic profiles were different from each other. The result of nucleotide sequencing for a partial gene of LVs from olive flounder and sea bass showed 98% and 88% homology respectively with those of LCDV-1 in GenBank. There was phylogenetic difference between LV from seabass and LV from flounder.

NOTES

Identification and diagnosis of *Aphanomyces piscicida* by PCR

Phadee Panarat^{1*}, *Kurata Osamu*¹, *Hirono Ikuno*², *Aoki Takashi*², *Hatai Kishio*¹

1. Division of Fish Disease, Nippon Veterinary and Animal Sciences University

2. Department of Aquatic-bioscience, Tokyo University of Fisheries

Some primers were designed from the sequence data of *Aphanomyces* sp. 84-1240 in Gene Bank to determine the sequence of ITS1, 5.8S and ITS2 genes of *Aphanomyces piscicida*, which is pathogen of mycotic granulomatosis in fishes. The sequence identity of this region was 100% among samples from diseased fishes in several countries. The specific primers were designed from sequence data of fish pathogenic *Aphanomyces piscicida*. The PCR using the specific primers detected only *Aphanomyces piscicida*, but not *Saprolegnia* spp., *Achlya* spp., the other *Aphanomyces* spp. *Lagenidium* spp. and *Dictyuchus* sp. The PCR method was also useful for the detection of the pathogen in the lesion of goldfish, *Carassius auratus*, artificially infected with *A. piscicida*. It was demonstrated that the primer set designed in this study was effective for identification of *A. piscicida* and diagnosis of mycotic granulomatosis in fishes.

NOTES

Development of a monoclonal antibody to hybrid catfish (*Clarias macrocephalus* x *C. gariepinus*) immunoglobulin

*Puttinaowarat S*¹, *Panyawachira V*¹, *Saengthong P*¹, *Polchana J*¹, *Thompson K*², *Adams A*²

1. Aquatic Animal Health Research Institute, Department Of Fisheries

2. Institute Of Aquaculture, University Of Stirling

Monoclonal antibodies (Mab) are commonly used to examine the immune response of fish. A Mab was developed here against the immunoglobulin of hybrid catfish (*Clarias macrocephalus* X *C. gariepinus*) for use as a probe to monitor the antibody response of this fish species against different pathogens. The Mab, referred to as B6 was of an IgG 2a subclass with k light chain. The Mab reacted strongly with the immunoglobulin of hybrid catfish when used in both enzyme linked immunosorbent assay (ELISA) and Western blot (WB). The Mab was found to react with the heavy chain of the hybrid catfish immunoglobulin in WB analysis, identifying a band at 66 kDa. The cross reactivity of the MAb was examined by ELISA against eight different fish species, including snakehead (*Channa striata*), giant snakehead (*Channa micropeltes*), Nile tilapia (*Oreochromis niloticus*), common carp (*Cyprinus carpio*), silver barb (*Babodes gonionotus*), rohu (*Labeo rohita*), grouper (*Epinephelus malabaricus*) and seabass (*Lates calcarifer*). The Mab was found to react with the immunoglobulin of common carp (*Cyprinus carpio*) and Silver barb (*Babodes gonionotus*) at 61% and 45% respectively, when compared to the hybrid catfish immunoglobulin.

NOTES

Cloning and expression of variable region of nucleocapsid gene of aquatic morbilliviruses for serological diagnosis

Sahoo Pramoda ^{1 *}, Parida S ¹, Hiscock J ², Barrett T ¹

1. Institute For Animal Health

2. University Of Reading

Morbilliviruses constitute a major threat to the health of animals and man; they group in the Morbillivirus genus in the family Paramyxoviridae. The more commonly known members are measles virus (MV), canine distemper virus (CDV), rinderpest virus (RPV), and peste-des-petits-ruminants virus (PPRV). In the last 14 years, newly recognised members of the morbillivirus family have caused many deaths among marine mammals, specifically pinipeds and cetaceans. These viruses are phocine distemper virus (PDV) found in pinnipeds and two closely related viruses isolated from cetaceans: dolphin morbillivirus (DMV) from striped dolphins and porpoise morbillivirus (PMV) from harbour porpoises. Serological and molecular studies have shown that cetaceans of many species have been exposed to these latter two viruses. The cetacean morbilliviruses are thought to spread to other species by means of the pilot whale which is considered to be the main vector of the virus. Altered migration patterns in animals caused by environmental changes is thought to be the origin of the epizootic of PDV in European seal in 1988. In order to determine the prevalence and understand the epidemiology and of these marine morbilliviruses it is necessary to develop simple, inexpensive and rapid differential diagnostic tests. The nucleocapsid (N) protein of the virus has a highly variable C-terminal region, which is also highly immunogenic. In the present study we have cloned and expressed the variable region of N protein gene from aquatic morbilliviruses (DMV and PDV), as well as from all other species of morbillivirus (RPV, PPRV, CDV and MV), to produce virus-specific antigens for developing specific serological diagnostic tests.

NOTES



Ultrastructural changes in the sperm and eggs of the black tiger shrimp, *Penaeus monodon*, before and after fertilization

Thaweepreda Pattira^{1,2*}, *Withyachumnarnkul Boonsirm*^{1,2,,}, *Plodpai Porntep*³, *Chawadej Jittiphphan*¹, *Pratoomchart Boonyarath*⁴, *Sobhon Prasert*¹

- 1. Faculty of Science, Mahidon University
- 2. Centex Shrimp, Mahidol University
- 3. Shrimp Culture Research Center, Charoen Pokphand Foods Public Company Limited
- 4. Department of Marine Biology, Faculty of Science, Burapha University

This series of studies were aimed at revealing the changes of the sperm and eggs before and after fertilization of the black tiger shrimp *Penaeus monodon*. The first study was to investigate the morphological events of the sperm at the time of acrosomal reaction. The sperm were incubated with egg water and the acrosomal reaction was observed under SEM and TEM. The sperm of *P. monodon* were nonmotile and composed of posterior spherical main body, central cap and anterior single spike extending from the cap. In the acrosomal reaction, the sperm underwent two phases of ultrastructural changes, acrosomal exocytosis and acrosomal process formation. It began with a gradual degenerative change of the spike. While the spike was degenerating, the cap region became enlarged; the outer and inner membrane swell and burst. Finally, the subacrosomal region was polymerized to form an acrosomal process. The second study was to compare the acrosomal reaction of the sperm between those from the male spermatophore and those from the female thelycum, following contact with egg water. It was found that 95% of the sperm from the female thelycum had an acrosomal reaction, while only 30% of sperm from the male spermatophore did. The third experiment was aimed at revealing the morphological changes of the eggs upon contact with seawater, the process known as egg activation. As soon as the oocyte was released from the gonopore into seawater, the cortical rods that appeared in the peripheral cytoplasm were released out from the oocyte surface. At the early stages of egg activation, the rods began to emerge from the cortical crypts of the oocyte and elevated the thin investment coat that encompassed the oocyte from its surface. Several sperm with the first phase of acrosomal reaction were observed on both the oocyte and the surface of the investment coat. The rods protruded from the surface and were completely expelled, within 45 sec after leaving the gonopore. Immediately after the completion of cortical rods extrusion, the rods began to dissipate and formed the jelly layer around the egg. By this time, the interaction between the sperm with second phase of acrosomal reaction and egg took place. Then the hatching envelope was formed at 1 min post-spawning, and the hatching envelope elevation was completed within 13-15 min post-spawning. The first and second polar body were extruded from the egg at 2-5 and 5-8 min post-spawning, respectively. During egg activation, exocytosis of three types of cortical vesicles; high-density vesicles, low-density vesicles, and granular vesicles occurred.

NOTES

A rapid multiplex real time PCR for the early detection of double targets: white spot syndrome virus (WSSV) and yellow head virus (YHV), in the black tiger shrimp *Penaeus monodon*

Taweetungtragoon Attaporn^{1,5*}, *Withyachumnarkul Boonsirm*^{2,5}, *Flegel Timothy*^{3,5}, *Boonsaeng Vichai*^{4,5}

1. National Center for Genetic Engineering and Biotechnology (BIOTEC)
2. Dept. of Anatomy, Faculty of Science, Mahidol University
3. Dept. of Biotechnology, Faculty of Science, Mahidol University
4. Dept. of Biochemistry, Faculty of Science, Mahidol University
5. Center for Excellence for Shrimp Molecular Biology and Biotechnology

WSSV and YHV cause high and rapid mortality in shrimp (*Penaeus monodon*). Recently, PCR has been commonly used for screening specific pathogen-free (SPF) shrimp in many hatcheries. However, performing PCR and RT-PCR assays take time; new quicker, multipurpose detection methods are being demanded by shrimp producers. The multiplex real time PCR is one route to classify quality of shrimp. Briefly, total nucleic acid of the viral and shrimp genetic materials were extracted, preserved and amplified with specific primer pairs for DNA virus, WSSV and RNA virus, YHV. The designed primers were optimized by annealing to two targets in one tube with fluorescent dye. The RT-PCR and PCR reaction were performed in one tube. Comparison between conventional and real time PCR was carried out. The results demonstrated that sensitivity was 10 times higher and 12 times faster than that of conventional PCR. By using the advantage of melting curve analysis, the optimized conditions could simultaneously generate two peaks of amplification products showing the different curve of both viruses based on their T_m . Analysis of the melting curve could demonstrate WSSV, YHV infection and no peak from uninfected organisms. Moreover, this method was tested with naturally infected with WSSV and YHV infected shrimp samples collected in Thailand. It works well with no gross sign shrimp from field. This detection system is rapid and sensitive for simultaneous dual detection. It should be a useful early detection system for shrimp culturists, especially for screening SPF broodstock and larvae in the hatchery.

NOTES



Theme 6

Infectious hypodermal and hematopoietic necrosis virus infection in Domesticated *P. monodon* broodstock

Chayaburakul Kanokporn^{1*}, *Withyachumnarnkul Boonsim*¹

1. Department of Anatomy and Centex Shrimp, Mahidol University

About 30% of a domesticated *Penaeus monodon* broodstock line of the fourth generation (F4) were infected with infectious hypodermal and hematopoietic necrosis virus (IHHNV), as detected by polymerase chain reaction (PCR). The growth rate of these broodstock was normal, compared to non-infected cohorts. Tissues from female broodstock were prepared for examination by routine histology using H&E staining and by in situ hybridization using a DNA probe specific for IHHNV. The tissues examined included hematopoietic tissue, lymphoid organ tissue, connective tissue, muscle tissue, ovarian tissue, subcuticular epithelium and neural tissue. In the H&E stained sections, Cowdry A Type inclusion (CAI) bodies frequently observed in IHHNV-infected cells of *P. vannamei* and *P. stylirostris*, were not observed in any of these tissues. By contrast, in situ hybridization with IHHNV-PCR positive broodstock gave positive hybridization reactions for all these tissue types except neural tissue while IHHNV-PCR negative brooders gave no positive hybridization reactions. The most abundant staining reaction was observed in hematopoietic tissue and lymphoid organ tissue. In the ovary, positive hybridization reactions were absent in oocytes and confined to the ovarian capsule and some follicular cells. Several degenerated/resorptive oocyte were observed. The results suggested some difference in the tissue tropism of IHHNV infection in *P. monodon* when compared *P. vannamei* and *P. stylirostris*. They also indicated possible detrimental effects of IHHNV infection on reproductive performance of the domesticated *P. monodon* broodstock.

NOTES



Potential indicators of stress response and their relation to survival in *Penaeus monodon*

de la Vega Enrique^{1,2,*}, *Hall Michael*¹, *Downs Craig*³, *Degnan Bernard*², *Wilson Kate*¹

- 1. Australian Institute of Marine Science
- 2. Department of Zoology and Entomology. The University of Queensland
- 3. Envirtue Biotechnologies Inc.

There is growing evidence describing the links between environmental stressors and the on-set of disease in many marine organisms including crustaceans. However, there is very limited data on the relationship between environmental conditions or stressors and variation of the immune system in marine invertebrates. Among the multiple environmental variables, temperature is potentially of great importance in ectothermic organisms since it directly affects their metabolism. A stress condition leads to an onset of molecular and physiological modifications, some which can be measured and used as molecular biomarkers. These have been used as an indication of cellular status, and hence of a stressed condition. Certain biomarkers, such as heat shock proteins (HSPs), have also been linked to the immune response in invertebrates. An analysis of these molecular biomarkers should provide information not only on the general health status of the organism but might be used as an alert system to help prawn farmers identify any environmental stressor which might induce the outbreak of a disease in the farm. As a test for the impact of temperature on prawns, four groups of 15 animals were exposed to 25, 27.5, 30 and 35 °C water temperatures, respectively. Survival ranged from 20% to 100% with highest mortalities in 35 C. Changes in expression levels of different molecular biomarkers in *P. monodon* were examined by Western Blotting and ELISA using invertebrate specific antibodies. The relationship between expression patterns in these molecular biomarkers will be discussed.

NOTES

A novel antimicrobial peptide isolated from the shrimp *Fenneropenaeus chinensis* after bacterial challenge

Guo Zhenyu ^{1*}, Xiang Jianhai ¹, Wu Changgong ¹

1. Institute of Oceanology, Chinese Academy of Sciences

Antimicrobial peptides are important for non-specific host defence in many animals. Here we report the presence of a novel antimicrobial peptide, active against both gram-positive and gram-negative bacterial, in the shrimp *Fenneropenaeus chinensis*. CZE (capillary zone electrophoresis) was applied to analyse hemolymph samples from shrimp before and after an immune challenge with the bacteria strains *Vibrio anguillarum*. The analysis of these spectra led to the following observation: the peptides relative concentration was found to increase by the first 3 hours post-challenge; and reached a maximum at 6 hours; after 24 hours, the plasma content of peptides appeared to be similar to that observed in unchallenged animals. On the basis of CZE and antimicrobial assay, the antimicrobial peptide was purified to homogeneity by Sep-Pak C18 extraction and reverse-phase HPLC. The partial N-terminal amino acid sequence obtained via Edman degradation revealed that it was proline rich and shared more than 60% identity in a 15-amino-acid overlap with the penaeidins, a family of shrimp *Litopenaeus vannamei*. The classification and differential count of circulating hemocytes have also been studied at different time after microbial challenge. The results showed that bacterial challenge triggered a plasmatic increase of the antimicrobial peptide concentration and gave implication of the simultaneous release of the peptide from the shrimp granulocytes.

NOTES



White Spot Disease in *Penaeus monodon*: case definition, accuracy of clinical diagnosis and description of an epidemic

Hao Nguyen Van^{1*}, *Corsin F*², *Phi T T*¹, *Phuoc L H*¹, *Tinh N T N*¹, *Turnbull J F*³, *Mohan CV*⁴, *Morgan K L*²

- 1. Research Institute For Aquaculture
- 2. The University Of Liverpool
- 3. University Of Stirling
- 4. College Of Fisheries India

This paper presents a case definition for WSD. It is developed from a detailed longitudinal study of 24 ponds growing *P. monodon* in a rice-shrimp farming system in Vietnam. At pond level WSD is defined by the occurrence of any dead shrimp in which WSSV is detected. Using this definition the sensitivity and specificity of white spots on the carapace as a diagnostic sign for WSD was 84.6% and 81.8% respectively. The sensitivity of mortality was by definition 100% but the specificity was 45.5%. This low sensitivity was reflected by the fact that a third of 'emergency harvested' crops in this study were negative for WSSV. This case definition highlights the importance of farmer observation and rapid detection of WSSV in limiting the economic impact of disease. We suggest that, in open farming systems, WSSV PCR resources would be better targeted at rapid diagnosis than at screening post-larvae. WSD was detected in 54% (13/24) ponds in this study. The pattern of disease suggested size, development stage or density of *P. monodon* as important in initial disease occurrence and subsequently an incubation period of 7 days. There was evidence of clustering in space and time. The temporal clustering appeared to be associated with a fall in temperature and water exchange at spring tides. Cases were clustered spatially in the area nearest the sea and we propose that water exchange may be important not just in lowering the temperature but also in introducing infection into a pond. These results suggest that manipulating water exchange may be a strategy for preventing disease. In addition to its effect on spring tides, the lunar cycle may also have a direct effect on the susceptibility of shrimp by synchronising moulting. These hypotheses need to be tested in larger epidemiological studies. UK government DFID research project 7051.

NOTES

The PK1 protein of the white spot syndrome virus (WSSV) is a nuclear kinase with autophosphorylation activity

Huang Chiu-Jung^{1}, Lo Chu-Fang¹, Kou Guang-Hsiung¹*

1. Department of Zoology, National Taiwan University

White spot syndrome virus (WSSV) is a virulent pathogen causing high mortality in cultured shrimp, and its genome encodes a serine/threonine protein kinase (PK1) that contains the major conserved subdomains. To determine the cellular localization of PK1, EGFP-PK1 fusion protein was expressed in Sf9 cells. Full-length PK1 protein was found to localize in the nucleus of transfected cells suggesting that PK1 contains a nuclear localization signal (NLS). To assay for PK1 activity, the pk1 gene was expressed as a GST-fusion in *E. coli* and purified by glutathione-agarose beads. The fusion PK1 catalyzed an autophosphorylation activity when incubated with gamma-32P [ATP]. When domains VIB and VII were deleted, the autophosphorylation and the MBP phosphorylation activity were abolished. Subdomain VII of PK1 contains the highly conserved DFG triplet. When D586 was replaced with an R residue, the autophosphorylation and MBP phosphorylation activity were abolished. Subdomain VII includes an Mg²⁺-binding loop, and it was experimentally confirmed that PK1 phosphorylation activity was Mg²⁺-dependent; Mg²⁺ was not replaceable by Mn²⁺ or Ca²⁺. Taken together, the results show that the PK1 protein of WSSV is a nuclear kinase with autophosphorylation activity.

NOTES



Evaluation of pathogenicity of bacterial strains in crustacean larvae by static bath: significance of monitoring bacterial counts.

Lavilla-Pitogo Celia R ^{1}, Catedral Demy D ¹, De Le Pena Leobert D ¹, Inui Yasuo ¹*

1. Fish Health Section, SEAFDEC Aqauculture Department

Although pathogenicity test has become the accepted method to explain the course of disease, differences in protocol under which tests are performed may result in strong variations and cause misinterpretations. Bacterial challenge in crustacean larvae by means of static bioassay may be complicated by their associated microflora or feed, if it is given. Therefore, in studies where only the initial inoculum dose was determined, effects of possible growth and replication of bacterial pathogen in the test chamber were not accurately measured and reported. In the course of our study on microbial diseases of hatchery-reared crab larvae, several tests were simultaneously conducted to prove the pathogenicity of luminescent *Vibrio harveyi* or the benign effect of potential probiotic bacteria associated with the alga *Chlorella* sp. In both tests, inoculation of 10² bacterial colony-forming-units (cfu)/ml in UV-sterilized seawater always resulted in increase of bacterial numbers by at least 2 logs after 24 h, proving that in static bath bacterial bioassays, growth and replication of test bacteria occurs in the test chamber. However, when the bacterial inoculum is 10⁵ to 10⁶ cfu/ml, no significant change in bacterial numbers occur. Furthermore, control chambers with no bacteria inoculated, were found to harbor between 10⁴ to 10⁵ cfu/ml of mixed bacterial population after 24 h, part of which is luminescent *Vibrio*. As these tests were conducted without feeding, the results prove that bacteria associated with the test larvae also proliferated in significant numbers inside test chambers. Thus, continued monitoring of bacterial population in the test chambers needs to be done to show its interaction with the already established microbiota in the larvae. Meaningful results of static bioassay of bacteria on crustacean larvae need to present a correlation of bacterial flora in tests chambers with survival of larvae.

NOTES



Electro-chemical processes for phytoplankton control and shrimp disease disinfection

Matsumura Masatoshi^{1*}, *Whangchai Niwooti*¹, *Alfajara Catalino G*¹, *Nomura Nakao*¹, *Migo Veronica P*², *Young Henry K*³

1. Institute Of Applied Biochemistry, University Of Tsukuba
2. National Institute Of Molecular Biology And Biotechnology, University Of The Philippines Los Banos
3. Aqua-Ecotechnology Research Inc

Some species of phytoplankton produce compounds that are extremely toxic to aquatic animals. Mass mortality of shrimp was observed to coincide with blooms of some species of *Oscillatoria*. Even in the case of beneficial phytoplankton massive algal die-off triggered the increase in pathogenic bacteria resulting in the mass mortality of shrimp. The control of phytoplankton over-blooming remains to be one of the serious problems in intensive shrimp culture. Electro-chemical processes offer several promising approaches for the prevention and remedy of pollution problems, and it has been widely applied in the drinking water industry and wastewater treatment. However, its application in the field of aquaculture is still very limited. We found that electro-chemical processes like electro-flotation and electro-oxidation are effective not only for controlling phytoplankton over-bloom but also for ammonia removal and inactivation of pathogenic bacteria.

NOTES



Prevalence of white spot syndrome virus (WSSV) and Monodon baculovirus (MBV) infection in Penaeus monodon postlarvae in Vietnam

Oanh Dang Thi Hoang ^{1*}, *Phuong Nguyen Thanh* ¹, *Walker Peter J* ², *Hodgson Richard AJ* ², *Preston Nigel* ³

1. College Of Aquaculture And Fisheries, Cantho University
2. CSIRO Livestock Industries
3. CSIRO Division Of Marine Research

A survey on the nature and prevalence of white spot syndrome virus (WSSV) and Mondodon Baculovirus (MBV) infection in Penaeus monodon postlarvae collected in hatcheries and nurseries in the centre and the south of Vietnam was conducted from the period between mid October 2001 to mid May 2002. Four hundred seventy-one samples were collected and subjected to two-step nested ploymerase chain reaction (PCR) analysis for WSSV using the IQ 2000 WSSV kit (Farming IntelliGene Technology Corporation, Taiwan). Of these, three hundred and eighty-eight samples were also tested for MBV using rapid staining method with Malachite green (lightner et al. 1996). The prevalence of WSSV infection in postlarvae collected from hatcheries and nurseries ws 20.6% whereas 46.4% prevalence was found in samples tested for MBV. In addition, it was found that prevalence infection of WSSV fluctuated according to sampling month and was the highest in February (37.7%) while prevalence infection of MBV remained the same throughout sampling months. Postlarvae samples from the central region, where hatcheries supply approximately 80% postlarvae for grow-out ponds in southern provinces, displayed a significantly higher prevalence of WSSV infection (21.1%) than those from the south (15.7%) (P<0.05). However , prevalence of MBV infection on postlarvae from the central part was not significantly different compared with samples from the south (44.3% and 46%, respectively) (P<0.05). The statistical significance of the differences in prevalence indicated that quality of postlarvae varies by production points and sources of supply. Therefore, the implications of the findings for shrimp health management in Vietnam will be discussed in this paper.

NOTES

Comparison of penaeid shrimp and insect parvoviruses suggests that viral transfers may occur between two distantly related arthropod groups

Roekring Songsak^{1*}, *Flegel TW*¹, *Nielsen Linda*¹, *Owens Leigh*², *Pattanakitsakul Sa-nga*³,
*Malasit Prida*³

1. Dept. Biotechnology, Faculty of Science, Mahidol University
2. Dept. Microbiology and Immunology, James Cook University
3. Division of Medical Molecular Biology, Faculty of Medicine, Siriraj Hospital

The DNA and putative amino acid sequences of representative insect and shrimp parvoviruses (subfamily Densovirinae) were analyzed using computer programs. Shrimp viruses included hepatopancreatic parvovirus (HPV) of *Penaeus monodon* (HPVmon) and *P. chinensis* (HPVchin), spawner-isolated mortality virus from *P. monodon* (SMVmon) and infectious hypodermal and hematopoietic necrosis virus (IHNV) from *P. vannamei*. Insect viruses included *Aedes aegypti* densovirus (AaeDNV), *Aedes albopictus* densovirus (AalDNV), *Junonia coenia* densovirus (JcDNV), *Galleria mellonella* densovirus (GmDNV), *Bombyx mori* densovirus 5 (BmDNV), *Diatraea saccharalis* densovirus (DsDNV) and *Periplaneta fuliginosa* densovirus (PfDNV). Virion size for all these viruses ranged between 18 and 30 nm diameter and ssDNA genome length was between 4-6 kb. Using BLAST or Clustal W with the sequence fragments available, no significant DNA homology was found except for 77% DNA identity between HPVmon and HPVchin. However, phylogenetic trees constructed by comparing DNA genome sequences for putative viral polypeptides, capsid proteins and nonstructural proteins placed the parvoviruses into two Clades: Clade 1 with SMVmon, PfDNV, DsDNV, GmDNV, JcDNV, and BmDNV; and Clade 2 with HPVmon, HPVchin, IHNV, AalDNV and AaeDNV. The 4 shrimp parvoviruses fell into two different groups that grouped in different insect parvovirus clusters.

NOTES



The white spot syndrome virus (WSSV) infects specific hemocytes of the shrimp *Penaeus merguensis*

Wang Y T^{1,2}, *Liu W*¹, *Seah J N*¹, *Lam C S*¹, *Xiang J H*², *Korzh V*¹, *Kwang J*¹

- 1. Institute of Molecular Agrobiolgy, The National University of Singapore
- 2. Institute of Oceanology, Chinese Academy of Science

The white spot syndrome virus (WSSV) was specifically detected by PCR in *Penaeus merguensis* hemocytes, hemolymph and plasma. This suggested a close association between the shrimp hemolymph and the viruses. Three types of hemocytes from shrimp were isolated using flow cytometry. Dynamic changes of the hemocyte subpopulations in *Penaeus merguensis* at different times after infection were observed, indicating that the WSSV infection selectively affected specific subpopulations. Immunofluorescence assay (IFA) and Wright-Giemsa double staining were performed to confirm this observation and to identify the hemocyte types susceptible to WSSV infection. These results revealed the cellular localization of the virus in the infected hemocytes. Electron microscopy showed that virus particles were found in both vacuoles and the nucleus of the semigranular cells (SGC) and vacuoles of the granular cells (GC). However, no virus could be detected in the hyaline cells (HC). Our results indicated that the virus infected two types of shrimp hemocytes: SCGs and GCs. Since SGCs express higher virus loads and exhibit faster infection rates, these cells are apparently more susceptible to WSSV infection.

NOTES

Effect of Tilapia *Tilapia hornorum* on luminous bacteria *Vibrio harveyi*

Tendencia Eleonor ^{1*}

1. Southeast Asian Fisheries Development Center, Aquaculture Department

In this study, the effect of tilapia *Tilapia hornorum*, stocked at different biomass, on *Vibrio harveyi* was investigated using 3-ton concrete tanks filled with micro-filtered water (5 micron) with salinity of 24 ppt, covered with black plywood to prevent sunlight penetration. All tanks were stocked with *Penaeus monodon* (20g) at 4 pcs/ton. Tanks for treatment 1 were stocked with 1 kg/10 m³ tilapia, treatment 2 at 3kg/10m³, and 5 kg/m³ for treatment 3. No tilapia was stocked in tanks that served as the control. Tanks were inoculated with luminous bacteria, *V. harveyi* to a final concentration of 10³ cfu/ml 6 hours after stocking with tilapia. There were two replicates per treatment. Luminous bacteria, total bacteria, fungal, and algal count were monitored regularly. Results showed that no luminous bacteria could be detected in treatments 2 and 3 from day 6 until termination on day 28. Luminous bacteria can still be observed in treatment 1 and the control on day 28. Total bacterial count and fungal count were comparable in all tanks. No algal growth was observed in all treatments by microscopic examination using a haemocytometer. Results suggest that the presence of *Tilapia hornorum*, in a shrimp culture system, at a density not lower than 3kg/10 m³ can control the occurrence of luminous bacteria. This culture system is sustainable, easy to manage, and cost less and therefore could be an alternative method to prevent the occurrence of luminous bacterial disease in shrimp

NOTES

Identification of *Penaeus monodon* baculovirus (MBV) in cultured *P. semisulcatus* in Islamic Republic of IRAN

Tokhmafshan M^{1}, Shariff Mohamed¹, Davoud Mohamed Hassan¹, Wang Yang Ging¹*

1. University Putra Malaysia

Shrimp aquaculture in I. R. Iran has a short history. Farmed shrimp production grow slowly from 1992 until 2000 when a number of large farm started to come production. In the 2000 the production was 4,500 MT and estimated that more than 7,000 MT will be produce by the 2001. During the period from August 1997 to March 1998, two thousand samples of cultured *Penaeus semisulcatus* postlarvae and subadults were collected from 5 hatcheries and 20 growout farms distributed in 3 province along the cost of Persian Gulf and Oman Sea. Based on grows sign, Histopathology, LM and TEM, *Penaeus monodon* baculovirus (MBV) have been recorded from the samples. The MBV is a rod-shaped baculovirus, DsDNA virus and the virion size is 300 ± 75 nm in diameter. The target organ of virus is hepatopancreas and midgut epithelium

NOTES

Does spawning stress trigger the replication of white spot syndrome virus in shrimp

Wang Han-Ching ^{1*}, Lo Chu-Fang ¹, Kou Guang-Hsiung ¹

1. Department of Zoology, National Taiwan University

In shrimp, a persistent viral infection can be converted to productive viral infection by stressors such as poor water quality and overcrowding. Here we investigate whether the stress associated with spawning can also trigger virus replication in shrimp. Quantitative real-time PCR was used for a temporal analysis of WSSV loads in a batch of 14 wild-caught black tiger shrimp (*Penaeus monodon*) brooders. For these specimens, four basic patterns emerge: Group 1 was comprised of 4 specimens which had a relatively high initial virus load (approximately $8 \times 10^2 \sim 8 \times 10^3$ viral DNA copies/per microgram total DNA) and in these specimens, the virus replicated rapidly up to the time of spawning. After spawning the virus levels remained high and all of these shrimp died within a few hours. Two other groups (II and III) both had similar initial virus loads (approximately $1 \times 10^1 \sim 6 \times 10^2$ viral DNA copies/per microgram total DNA), but in group II (5 specimens), as in group I, the virus replicated rapidly up to and beyond spawning and all the shrimp died soon after spawning, whereas in Group III (3 specimens) the virus load increased only relatively slightly (approximately a 10-fold increase) and the shrimp survived well beyond spawning. The group IV (2 specimens) shrimps did not spawn at all during the observation period, and no viral replication was triggered. At this time, it seems clear that spawning stress can trigger WSSV replication, but it is not known why or how this does not occur in some (25%) specimens.

NOTES

Phylogenetic analysis of replicase (ORF1b) amplicon sequences reveals a fourth genotype of yellow head complex virus in *P. monodon* from India

Wijegoonawardena Priyanjali ^{1*}, *Hodgson Richard* ¹, *Bhat Vishnu* ², *Walker Peter* ¹

1. CSIRO Livestock Industries

2. Marine Products Export Development Authority, India

Yellow head virus (YHV) and gill-associated virus (GAV) are closely related pathogens of farmed shrimp in the Asia-Pacific region. Yellow head disease was first reported in Thailand in 1990. Although there have since been reports of yellow head disease in other Asian countries, YHV has been positively identified only in diseased *P. monodon* from Thailand and Taiwan. GAV was first isolated from diseased shrimp from northern Australia in 1996 but the virus is also known to occur commonly as a chronic infection in healthy Australian *P. monodon*. We have recently reported that a third genotype in the yellow head complex occurs commonly in healthy *P. monodon* broodstock from Thailand and in postlarvae from Vietnam. The third genotype is more closely related to GAV than YHV and has not yet been detected in diseased shrimp. In this paper, we describe the identification of a fourth virus of the yellow head complex in healthy postlarvae from Andhra Pradesh in India. The new virus was detected by phylogenetic analysis of nucleotide sequences amplified from a 612 nucleotide region of the replicase (ORF1b) gene. The Indian virus is a new genetic lineage that is distant from all 3 previously known genotypes in the complex and is not detected by diagnostic PCR tests currently in use. In the amplified region, the new virus shares approximately 81-82 % nucleotide sequence identity with YHV, GAV and the third genotype from Thailand and Vietnam. By comparison, GAV and YHV share approximately 83% identity. The third genotype shares 82-83% identity with YHV and approximately 92% identity with GAV. We are continuing to investigate the distribution of all 4 genotypes in the Asian region and seeking evidence of the association of the new genotypes with outbreaks of disease.

NOTES



Cloning and characterization of the White spot syndrome virus (WSSV) p25 gene and its promoter region

Yu Li ^{1}, Sivan Subbu ¹, Kwang Jimmy ¹*

1. Temasek Life Sciences Laboratory, national University of Singapore

A complement DNA (cDNA) encompassing a 672bp open-reading-frame (ORF) was isolated from the pooled heads of WSSV-infected shrimp by RT-PCR. The ORF encodes 224 amino acids with estimated pI of 4.69 and calculated molecular mass of 25.0 kDa, bearing no significant homology to any known genes and thereby being called p25 gene. The deduced amino acid sequence contains a putative EF-hand calcium-binding domain and a C2H2 Zinc finger domain, predicting a possible involvement in DNA-protein interaction and transcription regulation. The transcript of the gene was determined using 5' RLM-RACE and 3' RACE. The transcription appeared to be initiated from a conserved CAGT motif consistent to that of early genes of many baculovirus and insects. 5' non-coding region of the gene was inserted before the luciferase gene in pHRG-b vector to investigate the promoter efficiency. Transient transfection assay performed on insect sf9 and monkey Marc145 cells revealed that this non-coding region has a remarkable promoter activity. The minimal sequence required for basal promoter efficiency was mapped with a panel of 5' end-truncated fragments of the non-coding region, and the promoter activity was also investigated in various cells including human TK143, monkey Marc145 and Vero, porcine PK15, chicken embryo-fibroblasts, carp EPC, insect sf9 and shrimp haemocytes. In addition, p25-coding protein was expressed in sf9 and Marc145, and its localization was characterized.

NOTES

Theme 7

Immunostimulatory effect of CpG oligodeoxynucleotides on the innate immune response of common carp, *Cyprinus carpio* L

A.R. Tassakka Asmi Citra Malina ^{1*}, Masahiro Sakai Masahiro ²

1. United Graduate School of Agricultural Sciences, Kagoshima University

2. Faculty of Agriculture, Miyazaki University

Bacterial DNA and synthetic oligodeoxynucleotide (ODN) containing unmethylated CpG motifs within certain flanking base pairs are recognized as a danger signal by the innate immune system of vertebrates. However, such effects in fish are poorly described. Using phagocytic activity, production of superoxide anion and lymphocyte proliferation response (LPR) assays, a panel of synthetic oligodeoxynucleotide was screened for immunostimulatory activity on the innate immune response of common carp (*Cyprinus carpio* L) kidney cells. In vitro treatment of fish by CpG-ODNs, resulted in increased responses of phagocytic activity and production of superoxide anion in kidney phagocytic cells. The CpG-ODNs also stimulated lymphocyte proliferation in the fish kidney cells. Intraperitoneal injection of CpG-ODNs into fish, resulted in enhanced responses of phagocytic activity and production of superoxide anion in kidney phagocytic cells. The serum lysozyme activity also increased in fish treated with CpG-ODNs. ODN composed of multiple CpGs were highly stimulatory in all assays, and CpGs located at the terminus of an ODN were ineffective. These results show that CpG-ODNs are effective immune stimulators for fish immune cells, suggesting that CpG-ODN may be useful in enhancing the innate immune response in veterinary applications for fish.

NOTES



Industry's Need of Standard Challenge Tests for Shrimp

Alday-Sanz Victoria^{1*}, *Decamp O*¹

1. INVE Technologies, Belgium

There are many products available in the shrimp market such as vaccines, immunostimulants or anti-viral components that claim to have an effect on the better survival or disease resistance of shrimp to viral and bacterial pathogens. However, no standard methodology has been available till now (to perform the challenge tests required in order to prove it). Results from bacterial and viral challenges commonly performed in shrimp lack confidence due to the difficulty to repeat and reproduce them consistently. In order to service the industry with reliable products, standard methodology should be developed. In this poster, we refer to the standardization already in place for fish and terrestrial animals and discuss the different variables that affect the challenges proposing alternatives to solve them.

NOTES

Dietary intake of levamisole enhances the immune response and disease resistance of the Asian catfish, *Clarias batrachus*

*Kumari Jaya*¹, *Sahoo Pramodo*^{2*}

1. Central Institute Of Freshwater Aquaculture
2. Institute For Animal Health

In order to determine the immunomodulatory effect of the dietary intake of levamisole in Asian catfish (*Clarias batrachus*), specimens were fed four different diets for 10 days: a commercial diet as control and the same diet supplemented with 50, 150 and 450 mg levamisole kg⁻¹ feed. The serum bacterial agglutination titre against *A. hydrophila* as a measure of specific immunity, serum haemolysin titre, lysozyme activity, globulin level and oxidative radical production by neutrophils as a measure of non-specific immunity as well as disease resistance against *A. hydrophila* challenge to vaccinated and non-vaccinated fish were evaluated at 0, 1, 2 and 3 weeks after last administration of levamisole. The results demonstrate that levamisole supplements at lower level (50 mg/kg), significantly ($p<0.05$) enhanced oxidative radical production, serum lysozyme and globulin levels to peak immediately after 10 days of feeding except for haemolysin titre which was enhanced after 3 weeks of feeding. The bacterial agglutination titre was raised at 50 and 150 mg/kg of levamisole feeding after 0 and 1 weeks. To examine the disease resistance capacity, fish were challenged with a virulent strain of *A. hydrophila* on 0, 1, 2 and 3 weeks after levamisole feeding and the percent mortality was recorded up to 10 days after challenge. Feeding of levamisole at 50 mg/kg significantly reduced the mortality in non-vaccinated fish up to 3 weeks than any other groups. There was no difference in mortality pattern in vaccinated groups of fish. On the contrary, higher level of levamisole feeding (450 mg/kg) caused immunosuppression in catfish as noticed from the reduction in most of the immune parameters along with increase rate of mortality. The present result supports the possible use of levamisole as immunostimulant in catfish farming.

NOTES

Effects of B-glucan on non-specific immune response in grouper: hematological analysis

Tavarró Rover John ¹, Torres James ^{1*}, Amar Edgar ²

1. University of the Philippines in the Visayas
2. Southeast Asian Fisheries Development Center - AQD

Groupers, *Epinephelus coioides* were injected intraperitoneally with curdlan, a B-1,3 glucan from bacterium *Alcaligenes faecalis*, at different doses of 0.5 mg, 5 mg, 10 mg, 20 mg, and 50 mg/kg body weight to determine the effect of immunostimulants on the non-specific immune response. Various hematological and non-specific immune response parameters were determined. A significant increase in the total leukocyte count was noted in immunostimulated fishes. It was observed that the percentage of granulocytes and monocytes increased while the lymphocyte numbers decreased in the differential leukocyte count in fishes treated with curdlan. These changes in the leukocyte composition was significant when compared with unimmunostimulated fish. It was only hematocrit determination which showed no significant difference between immunostimulated and unimmunostimulated fish. The production of superoxide anion, determined by NBT reduction and plasma lysozyme levels showed significantly increased values in immunostimulated fishes. Groupers injected with curdlan at a dose of 5 mg/kg body weight exhibited the most significant changes in the *in vitro* determinations. Doses higher than 5 mg/kg body weight have no more significant effect. This shows that curdlan at this dose may be used to enhance immunocompetency and disease resistance in grouper culture.

NOTES



Bacterial disease prevention in high value marine fish culture by vaccination

Vinitnantharat Somsak ¹, Gravningen Kjersti ^{2*}

- 1. Alpharma Inc. USA
- 2. Alpharma Norway

This presentation describes vaccination in high value marine fish culture; such as sea bass (*Dicentrarchus labrax* and *Lates calcarifer*), great amberjack (*Seriola dumerili*), yellowtail (*Seriola quinqueradiata*) etc., those become economically important for aquaculture lately. In the first experiment, sea bass (*D. labrax*) are immunized with *Vi-brio anguillarum* and *Photobacterium damsela* vaccine either by double immersion (Imm-Imm) or immersion followed by intraperitoneal injection (Imm-IP) at 15 weeks post primary vaccination. When fish are challenged with *V. anguillarum*, the mortality are 0.0 and 1.4 % for Imm-Imm and 0.0 and 0.0 % in Imm-IP vaccination compare to 62.3 and 46.7 % in non-immunized group at 7 and 26 weeks post primary vaccination respectively. The mortality, when challenge with *Ph. damsela*, are 1.4 and 8.7 % for Imm-Imm and are 0.0 and 10.0 % in Imm-IP vaccination compare to 55.1 and 50.0 % in non-immunized group at 7 and 26 weeks post primary vaccination respectively. The second experiment, great amberjack are immunized by intraperitoneal injection (IP) with *Ph. Damsela* and *Lactococcus garvieae* vaccine and are challenged 37 days later. At challenge, the mortality are 36.2 and 38.8 % in immunized group compare to 93.5 and 89.2 % in non-immunized for *Ph. damsela* and *L. garvieae* challenge respectively. In third experiment yellowtail are immunized by IP with *Ph. damsela* and *L. garvieae* vaccine but challenged with *L. garvieae* only. The mortality are 0.0 and 9.8 % in immunized group compare to 92.0 and 92.5 % in control when challenge 14 and 60 days post vaccination respectively.

NOTES

Theme 8**Orally administered praziquantel as a treatment for monogeneans infecting *Seriola quinqueradiata*: efficacy and practical considerations for a commercial fish farm**

Chambers Clinton ^{1*}, Ernst Ingo ¹

1. The Department of Environmental Biology, The University of Adelaide

Parasitic diseases are a serious concern for sea cage aquaculture and in some instances they may threaten industry viability. In Japan, cultured *Seriola* spp. are vulnerable to outbreaks of the monogenean parasites *Benedenia seriolae*, *Heteraxine heterocerca*, *Neobenedenia melleni* and *Zeuxapta seriolae*. Control of monogeneans is currently achieved by bathing fish in freshwater or hydrogen peroxide. This procedure significantly increases production costs because it is labour and infrastructure intensive, stresses fish and may result in lost growth. In-feed treatment for monogenean infections provides a desirable alternative to bathing. Presently, only one in-feed medication is licensed in Japan for use against monogenean parasites of *Seriola* spp. The active ingredient of this product is praziquantel, which has demonstrated therapeutic activity against a broad range of helminths. We assessed the efficacy of orally administered praziquantel as a treatment for *B. seriolae* and *H. heterocerca* infections in a commercial yellowtail farm. Treatment was found to reduce total abundance of *H. heterocerca* by up to 97.9% and juvenile abundance by up to 99.5%; total abundance of *B. seriolae* were reduced by up to 62.9% and juvenile abundance by up to 70.5%. Methods of application may have a significant effect on the efficacy of orally administered praziquantel. The key practical considerations for praziquantel treatment are discussed, recommendations for improving efficacy made and areas requiring further research identified.

NOTES

Studies on antiviral and antibacterial substances of *Spirulina platensis* for prevention the infectious diseases in black tiger shrimp (*Penaeus monodon*) caused by white spot syndrome virus and vibrio harveyi.

Hemtanon Piyalai^{1*}, *Deregbusarakom Sataporn*¹, *Bunyawiwat Wissanu*², *Tontithagool Opas*¹

1. Walailuk University, Nakornsri thamarat, Thailand 8060

2. Kasetsart University, Bangkok, Thailand

Experiments were designed to 3 parts; the first, the effect of crude extract from *S. platensis* against white spot syndrome virus (WSSV) and application to post larvae 15 and adult stage of *P. monodon* ; the second, the effect of crude extract against *Vibrio harveyi* and application to *P. monodon* and the final was the effect of *S. platensis* on shrimp cellular immunity. The crude extract 1, 0.1, 0.01 and 0.001 mg/ml were mixed with WSSV. After 14 days of injection, all of concentrations could inhibit WSSV infection. For post larvae 15, fed with the steamed egg containing 0, 0.5 and 5 % of *S. platensis* and challenged with WSSV, the survival rate were 73.75±10.34, 86.50±8.80 and 83.17±3.87 %. The results from PCR diagnosis, the infection were 100, 0 and 16.67 %. Application for adult, the survival rate were 61.00±38.74, 96.00±8.94, 100.00±0.00 and 92.00±10.95 %. The effect of crude extract on the growth of three strains of *V. harveyi*. The result showed that it could inhibit the growth of all strains and the minimum concentration was 2.5 mg/ml. The application for adult, fed with the diet containing 0, 0.5 and 1 % of *S. platensis*. The result showed that bacterial clearance higher in treatments than those control group, there was significant different (P<0.05). For cellular immunity, phenoloxidase activities in the haemolymph were not significant different between treatments and control group (P>0.05). This study indicated that *S. platensis* could control WSSV and *V. harveyi* infection in shrimp.

NOTES



Health status of cultured shrimp at harvest - epidemiological significance

Mohan CV ^{1*}, Sashikala G ¹, Jagannath V ¹, Ahmed I ¹, Corsin F ², Padiyar PA ¹, Madhusudan M ¹, Turnball JF ³, Hao NV ⁴ Morgan KL ²

1. Fish Pathology Laboratory, Department Of Aquaculture, College Of Fisheries
2. Department of Veterinary Clinical Science And Animal Husbandry, The University Of Liverpool
3. Institute Of Aquaculture, University Of Sirling
4. Research Institute For Aquaculture

A longitudinal observational study was undertaken in 70 shrimp farms in Kundapur, Karnataka, India from September 1999 to April 2000 to identify risk factors associated with outbreaks of White Spot Disease (WSD) in cultured *Penaeus monodon*. As a part of the larger study, 400 shrimp/pond were randomly collected from 62 ponds at the time of harvest and observed for the presence or absence of white spots on the carapace. From this, 100 shrimp were randomly selected for recording the length and weight and for detailed clinical examination. Per pond, 10 each of clinically sick and clinically healthy looking shrimp were selected and fixed in 10% neutral buffered formalin for histopathological studies. In 21 (33.9%) ponds, only white spot syndrome virus (WSSV) inclusion bodies were detected. Samples from 21 (33.9%) ponds showed evidence of only chronic inflammatory lesions (CIL) characteristic of chronic bacterial infections. In 13 (21%) ponds, both WSSV inclusion bodies and CIL were observed. Samples from 7 (11.3%) ponds did not have any pathology. In total, WSSV inclusions were recorded in 55% (34/62) and CIL in 55% (34/62) of the ponds. On a pond basis, there was no significant difference in the health status of sick and healthy looking shrimp. If WSSV ($p<0.001$) or CIL ($p=0.001$) was present in clinically sick shrimp in a pond, it was also more likely to be present in the healthy looking shrimp in the same pond. Ponds with clinical white spots on shrimp, were more likely to have WSSV inclusions in the samples ($p<0.001$), while there was no association with CIL ($p=0.120$). In 77% of the ponds (48/62), dead shrimp had been observed and collected by the farmers prior to harvest. Ponds where dead shrimp were positive for WSSV inclusions ($p=0.001$) or CIL ($p=0.008$) were also more likely to have the same condition at harvest. Ponds positive for WSSV inclusions at harvest were significantly associated with high mortalities prior to harvest ($p<0.001$) and shorter rearing period ($p<0.001$). On the other hand, ponds positive for CIL at harvest, were significantly associated with low or no mortalities prior to harvest ($p=0.01$) and longer rearing period ($p=0.002$). Monitoring pond side mortality and rapidly confirming WSD can assist shrimp farmers in proper decision making.

NOTES



Effect of methylparathion on the susceptibility of shrimp (*Litopenaeus vannamei*) on the development of vibriosis

Roque Ana^{1*}, *Labrie Lauke*², *Gomez-Gil Bruno*¹, *Turnbull James F*²

- 1. CIAD/Mazatlan Unit for Aquaculture and Env. Management
- 2. Institute of Aquaculture, University of Stirling

Under increasing pressure of environmental concerns, a replacement of persistent pesticides with more rapidly degradable products has been observed over the past two decades. However, these pesticides can still affect non-target species and can be associated with slow growth and increased susceptibility to infections. In this study juvenile shrimp (*Litopenaeus vannamei*) were exposed to combinations of methylparathion and a *Vibrio* sp. There were five treatments, each with five replicates and 10 shrimp per replicate. In one treatment shrimp were fed pellets containing 0.080 mg of methylparathion g-1 for 4 days and injected intramuscularly with *Vibrio parahaemolyticus* on day 5 (MPAR/V). The other treatments were shrimp fed with methylparathion and injected with sterile saline solution (MPAR), shrimp fed with pellets containing acetonitrile (solvent for methylparathion) and injected with *V. parahaemolyticus* (V), shrimp fed with pellets containing acetonitrile and injected with a sterile saline solution (A) and a group that only received uncontaminated pellets (C). Mortality was recorded and the surviving shrimp were sampled for histology on the fourth day after injection. Analyses of the incidence rate for mortalities (IR) in the treatments revealed that feeding methylparathion was significantly associated with an increase in IR. The injection of the *Vibrio* sp. was significantly associated with increased IR and histopathological lesions. The combination of feeding methylparathion and injection of the *Vibrio* sp. was also associated with a significant increase in IR when compared with either treatment alone. The study provides strong experimental evidence that exposure to methylparathion can increase susceptibility to *Vibrio* infection.

NOTES

Acute toxicities of deltamethrin and lambda-cyhalothrin to the fry of green grouper (*Epinephelus tauvina* Forsskål) and milkfish (*Chanos chanos* Forsskål)

Silapan Judith^{1*}, *Cadiz Geofe*¹

1. UP in the Visayas, Cebu College

A static 96-hr acute toxicity test was used to assess the potential ecological effects of deltamethrin and lambda-cyhalothrin to the fry of green grouper *Epinephelus tauvina* and milkfish *Chanos chanos*. The deltamethrin and lambda-cyhalothrin LC50 of the grouper fry (0.23 g mean body weight) were 3.6 and 8.25 µg/L, respectively. For both pesticides, the NOAEC (No Adverse Effect Concentration) value was 2.5 µg/L. The deltamethrin and lambda-cyhalothrin LC50 of the milkfish fry (0.06 g mean body weight) were 0.14 and 0.006 µg/L, respectively. The NOAEC values were .025 and .0025 µ/L for deltamethrin and lambda-cyhalothrin, respectively. The two species are the leading fin fishes cultured in cages and pens that are located in estuarine and semi-exposed bays in the Philippines. Moreover, these areas are the collecting grounds for wild grouper and milkfish fry. On the other hand, the two toxicants are pyrethroid insecticides that have gained wide use in the country for the control of major insect pests of common Philippine crops. There is a growing concern among mariculture farmers on the risk of transient agricultural run-offs. Reports of fish kills in these facilities have been recorded following first rainfall of the wet season. The basic effects data provided by this study may be used as input for risk assessment of run-off related pesticides on mariculture organisms. Key words: deltamethrin, lambda-cyhalothrin, effects assessment, acute toxicity, fry of *Epinephelus tauvina* and *Chanos chanos*

NOTES



Contamination of mycobacterium spp. in live feed

Somsiri Temdoung ^{1*}, Puttinaowalat Suppalak ¹, Soontornwit Suriyan ¹, Lacharoje Sitthichok ¹

1. Aquatic Animal Health Research Institute

The importance of moina, mosquito larvae and blood worm as live feed for tropical fish, especially carnivorous fish, is well known. These live feed always has contaminate with pathogens that cause diseases in aquarium fish. Mycobacteriosis is a chronic bacterial disease infecting aquarium fish. The detection of Mycobacterium spp contaminated in live feed in Bangkok and 4 provinces around Bangkok by isolation and polymerase chain reaction technique revealed four pathogenic bacteria, namely Mycobacterium spp., M. fortuitum, M. marinum and M. chelonae. Contamination of M. fortuitum showed high prevalence. Live feed are recognized as one of the important sources of mycobacteria that cause diseases in aquarium fish. Feeding of contaminated live feed should be avoided. The practical on producing live feed free from mycobacteria is recommended. Key word : Mycobacterium spp., Moina, Mosquito larvae, Blood worm

NOTES

Theme 10

Mycobacteriosis in rancho goldfish (*Carassius auratus*) imported from Japan

*Lawhavit Ong-Ard*¹, *Wada Shinpei*², *Hatai Kishio*^{2*}

1. Kasetsart University
2. Nippon Veterinary And Animal Science University

Rancho, Japanese goldfish, is very famous for culturing among Thai people. Each year, many company have imported rancho from the famous farm in Japan. In the past winter, January 2002, 30 percent of imported juvenile fish showed inactive swimming, anorexia and lethargy. Therefore, the six diseased fish were collected to investigate the cause of diseases. The clinical signs of the fish showed asymmetrical swelling of abdomen, pale discoloration of gills, swelling and discoloration of head and trunk kidneys. Abdominal organs were attached to peritoneal wall, indicating peritonitis, associated with numerous white nodules of various sizes. These nodules were also found on the abdominal organs, head and trunk kidneys and heart. The isolation was done with Ogawa egg medium and identified as *Mycobacterium chelonae* by the PCR method at Aquatic Animal Health Research Institute, Department of Fisheries, Thailand. The histopathological features showed that these white nodules on the affected organs were chronic proliferative lesions composed of multiple caseous epithelioid cell granulomata with surrounding granulation tissue. In the central caseous area, a number of colonies of slender, long rod, Gram positive were found to be acid-fast by Ziehl-Neelsen method. These acid-fast bacteria were also observed in the epithelioid cells and in the macrophages of the granulation tissue. From these findings, this case was diagnosed as serious systemic mycobacteriosis.

NOTES

Fish Kill of Mullet *Liza klunzingeri* in Kuwait Bay: The Role of *Streptococcus agalactiae* and the Influence of Temperature

*Al-Marzouk Ahmed*¹, *Duremdez Roselyn*¹, *Yuasa Kei*², *Al-Zenki Sameer*¹,
*Al Gharabally Hashem*¹, *Munday Barry*^{3*}

- 1. Kuwait Institute for Scientific Research, Mariculture and Fisheries Department, P.O. Box 1638, Salmiya 22107, Kuwait
- 2. Fisheries and Aquaculture International Co., Ltd., Kohji- Machi Bldg., 4-5 Kohji-Machi, Chiyoda-Ku, Tokyo 102-0083, Japan
- 3. School of Human Life Sciences, University of Tasmania, Locked Bag 1-320, Launceston, Tasmania 7250, Australia

A massive fish kill occurred in Kuwait Bay during August and early September 2001. Wild mullet *Liza klunzingeri* represented 99% of the fish kill, whereas the remaining 1% consisted mainly of silvery croaker *Otolithes argenteus*, giant sea catfish *Arius thalassinus* and striped grunt *Rhonciscus stridens*. The average seawater temperature during the incident was 32.3°C. Affected fish showed erratic swimming, hemorrhages around the mouth, abdomen, pectoral and pelvic fins. They also exhibited internal hemorrhage and exophthalmia. Bacterial isolates obtained from the four wild fish species, one cage-cultured species (sea bream *Sparus auratus*) and sewage samples from two beach locations were used to identify the causative organism. The biochemical, biophysical and API 20 Strept tests identified the bacterium as b-hemolytic, group-B *Streptococcus agalactiae*. *S. agalactiae* can grow in a temperature range from 18 to 43°C, but not at 5, 12 or 45°C. Also, it can grow in a salinity range between 0.5 to 6.0% NaCl, but not at 6.5%. To investigate the effect of temperature on the susceptibility of fish to infection, healthy mullet kept at two seawater temperatures (25 and 33°C), were intraperitoneally injected with four bacterial doses (102, 104, 106 and 108 CFU/mL). At 33°C, 100% mortality was produced after 24h, when fish were injected with 108, whereas the other doses produced the same mortality after three to four days. At 25°C, the mortality varied between 20 and 60% among the different doses. The results clearly implicate the significant role of high temperature in increasing the virulence of *S. agalactiae* causing the massive mullet mortality. The possible relationship of this outbreak to another in August and September 2000 in cultured silver pomfret *Pampus argenteus* is discussed.

NOTES



Isolation of IHN virus from imported turbot (*Scophthalmus maximus*)

Yulin Jiang ^{1*}, *Hong Liu* ¹, *Long Ying Gao* ¹, *Xiujie Shi* ¹

1. Shenzhen Exit and Entry Inspection and Quarantine Bureau

At end of May 2001, turbot fry were imported from France through Hong Kong. 5 fry were sent to Lab for routine detection. The liver, spleen, kidney and brain of fish were collected and homogenized, some of them were inoculated into EPC, CO, RTG-2, CHSE cell monolayers and cells were incubated at 15 C, and 25 C for isolation of viruses. The RNA in some organs were extracted for detection of viral RNA using PCR according to Diagnostic Manual for Aquatic Animal Diseases (OIE, 2000). CPE appeared in EPC cell culture at 15 C in 1 week, which indicated that there was a virus in fish organs. 786 bp segment and 323 bp sigment were amplified respectively after RT-PCR and nested -PCR. DNA sequence of RT_PCR products from this virus (2000-479) and IHNV strain were compared. The sequence homologies between 2000-479 and IHNV_WRAC, RB, 11290 etc. was 91.7% - 98.6% . It is suggested that the virus isolated from turbot fry a closely related with IHNV. These results give reasonable evidence that the turbot may be infected with IHNV. The susceptibility of turbot to IHNV was demonstrated experimentally (I/P inoculatin) iby a French workers some years agol However, natural infection of IHNV in a marine fish species has not been demonstrated yet. It is the first finding of natural infection of IHNV in turbot.

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